

**River Glen:
River Channel Assessment - Annex B**

An Analysis of Invertebrate Records for The River Glen, 1976 - 1991

September 1992



ENVIRONMENT AGENCY

NATIONAL LIBRARY &
INFORMATION SERVICE

ANGLIAN REGION

Kingfisher House, Goldhay Way,
Orton Goldhay,
Peterborough PE2 5ZR

**Prof. G Petts
M Bickerton**

Freshwater Environments Group

Anglian Regional Operational Investigation 447

OI/447/4/A



NRA

National Rivers Authority

**River Glen:
River Channel Assessment - Annex B**

An Analysis of Invertebrate Records for The River Glen, 1976 - 1991

September 1992

Environmental Agency

Thames Region -

Library Catalogue

Class No.

Accession Code ADEH **.....**

**Prof. G Petts
M Bickerton**

Freshwater Environments Group

Anglian-Regional Operational-Investigation 447

01/447/4/A

Environmental Agency

Thames Region

Library Catalogue

Class No.

Accession Code

ANNEX B
AN ANALYSIS OF
INVERTEBRATE RECORDS FOR THE RIVER GLEN, 1976-1991

Undertaken for the National Rivers Authority, Anglian Region

By

**FRESHWATER ENVIRONMENTS GROUP,
INTERNATIONAL CENTRE FOR LANDSCAPE ECOLOGY,
LOUGHBOROUGH UNIVERSITY,
LEICESTERSHIRE, LE11 3TU.**

Supervised by Professor G.E. Petts

Researcher: M.A. Bickerton

September 1992

TABLE OF CONTENTS

1 SUMMARY	1
1.1 The data set.....	1
1.2 Reach and site level differences in invertebrate fauna	1
1.3 Biological quality	1
1.4 Temporal changes	1
1.5 Within-site temporal changes.....	2
1.6 Seasonal changes	2
1.7 Changes in invertebrate fauna with changing biological quality	2
1.8 Recommendations	2
2 INTRODUCTION	3
3 THE DATA SET	4
3.1 Sites	4
3.2 Dates	4
3.3 Taxonomic lists	4
4 METHODS	8
4.1 Sampling.....	8
4.2 Standardisation of taxa lists	8
4.3 Biotic indices.....	8
4.4 Sample classification using TWINSpan.....	8
4.5 Sample ordination using CANOCO.....	9
5 ANALYSIS AND DISCUSSION	10
5.1 Taxonomic diversity.....	10
5.2 Biological quality	10
5.3 Ordination of all samples based upon the family data	10
5.4 Classification of all samples based upon the family data.....	14
5.4.1 The first major division.....	14
5.4.2 Subdivision of the main and East Glen sites.....	14
5.4.3 Subdivision of the West Glen and Tham sites.....	18
5.4.4 Definition of reach communities	18
5.5 Long-term changes in biological quality	21
5.5.1 Changes in scores at the catchment scale.....	21
5.5.2 Changes in scores at the site and reach scale.....	21
5.6 Site case studies.....	24
5.6.1 West Glen, upstream of Essendine	24
5.6.2 West Glen at Banthorpe Lodge.....	26
5.6.3 River Glen at Kates Bridge	28
5.6.4 River Glen at Surfleet Seas End.....	30
5.6.5 East Glen at Braceborough.....	32
5.7 Seasonality	34
5.8 Performance of biotic indices	38
5.8.1 Variation within LQI classes.....	38
5.8.2 Classification and Ordination of samples within LQI classes.....	38
5.8.3 Implications of within LQI class differences.....	47
6 CONCLUSIONS	50
6.1 Biological records	50
6.2 Site and reach level differences in Invertebrate communities.....	50
6.3 Biological scores.....	51
6.4 Long-term changes.....	51
6.5 Seasonal differences.....	52
7 RECOMMENDATIONS.....	53
7.1 Preliminary reach assessment	53
7.2 Selection of routine monitoring sites	53
7.3 Biological monitoring - frequency.....	53
7.4 Biological monitoring - field procedure.....	55
7.4.1 Invertebrate sampling.....	55
7.4.2 Invertebrate identification	55
7.5 Associated routine monitoring	55
8 REFERENCES.....	56
APPENDICES	
APPENDIX 1 TAXA LISTS FOR ALL SAMPLES	
APPENDIX 2 BIOTIC SCORES FOR ALL SAMPLES	
APPENDIX 3 TWINSpan PREFERENTIAL TAXA	

1 SUMMARY

1.1 The data set

The data set comprises invertebrate records from 44 sites on the main River Glen and its tributaries. These can be grouped into a series of physically defined reaches. Sites and reaches are shown in Figure B1. Site codes are explained in Table B1.

Records for each site vary in frequency and distribution throughout the 1976-91 period. Taxonomic level of identification varies between records, therefore the data used in the analyses have been reduced to presence / absence family level.

1.2 Reach and site level differences in invertebrate fauna

TWINSPAN classification (Figures B5 and B6) and ordination using correspondence analysis (Figure B4) revealed distinct reach and site level differences in fauna which can be related to a variety of water quality, hydrological, hydraulic and other environmental variables. Figure B7 summarises the classification of sites (emboldened site codes indicate the majority of samples from this site falling in this group; plain text indicates just two or more samples). "Key taxa" are those showing strong preference for the group indicated. Classification and ordination first separated the Tham and middle West Glen sites from the rest, on the basis of their rich fauna including many taxa associated with moderate to fast flows and stable substrates. Subsequently the East Glen sites, Lower Glen/Bourne Eau sites and a group of "intermediate" sites were distinguished; the first having a very impoverished fauna (related to intermittent flow and poor water quality), the second having a fauna typical of slow-flowing, weedy habitats, and the third having an intermediate fauna, sharing taxa found in the other two. Communities associated with three major reach-types: "Upland", "lowland" and "intermediate" were identified and described are in Table B3.

1.3 Biological quality

Biological quality varied between sites in a way consistent with the divisions described above. The highest scoring sites were around the lower Tham and the West Glen below the Tham confluence, reflecting the diverse fauna of these sites and particularly the predominance of high-scoring insect taxa characteristic of these high-quality habitats. High scores were also obtained from the lower Glen sites around the Bourne Eau confluence - water and habitat quality is also high, but due to physical habitat differences the actual taxa present are very different from the Tham / West Glen. The interrelated factors: small size, intermittent flow and poor water quality are assumed to be responsible for the poor scores in the East and upper West Glens.

1.4 Temporal changes

The overall trend in biological scores during the period 1976-1991 was upwards, the majority of sites showing a steep rise in all scores from 1976 to the mid to late 1980's, followed by a slight decline in 1991. Patterns of change did vary between sites, as illustrated in Figure B10 which shows sites showing significant trends in scores. Notably the lower main Glen sites displayed constant improvement in scores. "intermediate sites" showed a slight decline in 1990-91, and some East Glen sites steadily declined.

1.5 Within-site temporal changes

Detailed analysis of a number of sites (section 5.6) whilst confirming the temporal trends mentioned above revealed different faunal changes associated with temporal changes in biological quality. responses to detrimental changes in habitat quality are complex, sometimes involving a reversion to a poorer fauna similar to that many years earlier, while in other cases simply a reduced version of the fauna of the previous year.

1.6 Seasonal changes

Evidence was present for seasonal differences in the frequency of individual taxa and biotic scores, particularly ASPT. The greater seasonality in the life-histories of higher-scoring insect taxa may account for the changes in ASPT. Variability in the frequency of sampling in each month prevent detailed investigation of seasonal changes in this data set.

1.7 Changes in invertebrate fauna with changing biological quality

Diversity within the higher LQI classes is high (section 5.8), with reach and site scale differences as clear within LOI classes as within the whole data set. The clear implication is that site and reach specific differences other than water quality are vital in determining the fauna. This was confirmed by examining changes in fauna associated with changes from LQI class B and C to class A-A++. The "improved" fauna associated with a change from the lower to higher class was almost invariably predictable and dependent on the initial fauna, which in turn was related to specific reaches. Table B4 summarises the dominant within-class reach groups and the taxa associated with changes from the lower to higher class.

1.8 Recommendations

Recommendations are made towards an improved system of biological monitoring, involving an initial detailed survey of a river system in order to determine reach-communities, followed by coordinated invertebrate, macrophyte and hydrological monitoring. This strategy aims at enabling the magnitude, nature and causes of habitat changes to be determined and related to short and long term changes in invertebrate communities.

2 INTRODUCTION

The River Glen, incorporating the Rivers East and West Glen and a lower section below their confluence downstream to the confluence with the River Welland, supports a wide variety of invertebrate habitat types from fast flowing, gravel-bed reaches to slow flowing, fenland drains.

Monitoring of the invertebrate communities, both routine and in response to pollution events, has been regularly practiced on the Glen since 1976. During this time the frequency of sampling and level of invertebrate identification has varied considerably, but leading up to the 1990-91 situation where sampling at most sites was carried out at least twice a year and a substantial proportion of the fauna is identified to species level.

Despite its inconsistencies, the data set for the 1976-91 period contains a substantial amount of information, comprising some 250 records from 44 sites distributed throughout the River Glen system.

This report aims to assess:

- the nature of the invertebrate communities in different parts of the catchment;
- long-term changes in biological quality at sites with long records;
- the performance of indices of biological quality in both monitoring within-site change and comparing sites within the river system.

3 THE DATA SET

3.1 Sites

The distribution of sites on the East, West and main River Glen is shown in Figure B1. Sediment analysis of the West Glen and main Glen allow the river to be divided into a number of physically distinct reaches: The upper West Glen (reach A); the West Glen around Essendine (reach B); the West Glen around Banthorpe Lodge (reach C); the main Glen from the confluence of the East and West Glens to the confluence with Bourne Eau (reach D); the East Glen (E - lack of physical information prevents this from being further subdivided); the lower main Glen downstream of the confluence with Bourne Eau (G); Bourne Eau (B); and the River Tham (T).

The codes given to sites are listed in Table B1. Where a particular sample is referred to later in the report, the site code is followed by the month and year. In the one instance where two samples from the same site were taken in the same month, the full date follows the site code.

3.2 Dates

Records exist for most years in the 1976-91 period, with the exception of a gap from 1980 to 1981. The year 1988 has a particularly good site coverage due to a special survey being carried out (Extence, 1989), the results of which have been added to the NRA routine sampling data. 1990 has good seasonal coverage of the routine monitoring sites, and it is understood that current practice is to sample in spring, summer and autumn. Table B2 shows the years in which each site was sampled.

Although a few sites have regular records for the entire period (eg. WLD, GSS), the majority have a patchy record, some sampled no more recently than the early 1980's (eg. GGG), and some only in recent years (eg. ELN).

3.3 Taxonomic lists

Each record consists of a list of taxa found at the site, occasionally with an indication of abundance. Taxa recorded, and the level of identification, are undoubtedly influenced by the format of the recording sheet. In general the level of identification expected has tended towards the species level in later years; the earliest recording sheets are mainly to family level, ie. sufficient to allow calculation of the standard biological indices.

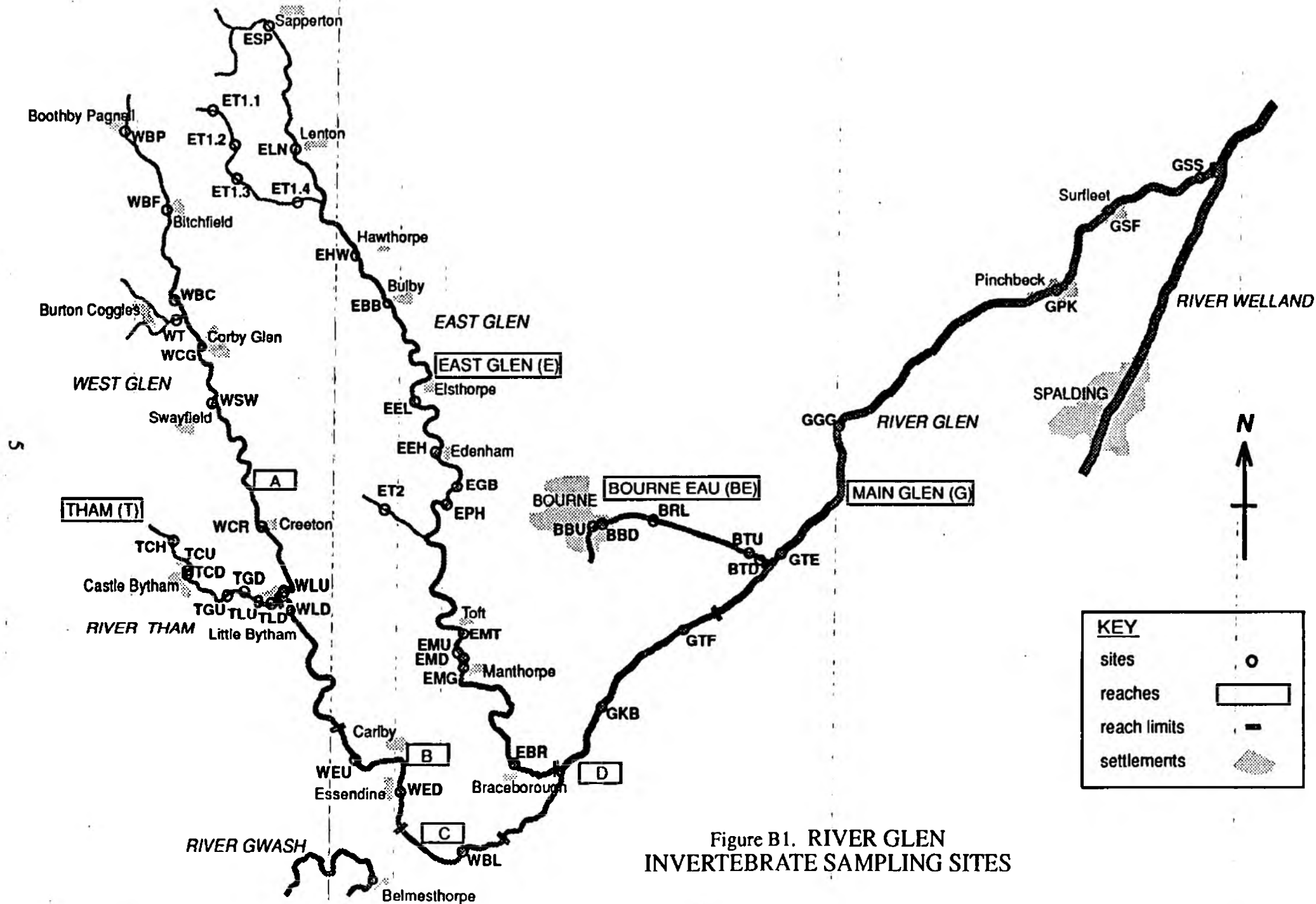


Figure B1. RIVER GLEN
INVERTEBRATE SAMPLING SITES

Table B1. River Glen site codes and locations

SITE CODE	N.G.R.	LOCATION
ELN	TF 022302	E. Glen at Lenton
EHW	TF 038272	E. Glen at Hawthorpe Rd.
EBB	TF 047260	E. Glen at Bulby - between fd. and br.
EEL	TF 054233	E. Glen at Elsthorpe
EEH	TF 061219	E. Glen at Edenham d/s A151 rd.br.
EGB	TF 067209	E. Glen at Gravel Bridge, d/s rd.br.
EPH	TF 064205	E. Glen at Pasture Hill Fm. d/s br.
EMT	TF 068169	E. Glen at Manthorpe Toft, u/s STW
EMU	TF 066164	E. Glen at Manthorpe, u/s STW outfall
EMD	TF 067163	E. Glen at Manthorpe, d/s STW outfall
EMG	TF 068160	E. Glen at Manthorpe G.S., d/s weir
EBR	TF 082135	E. Glen at Braceborough, u/s & d/s rd. br.
ET11	SK 998313	Trib.of E. Glen
ET14	TF 022288	Trib.of E. Glen
WBP	SK 973307	W. Glen at Boothby Pagnell
WBF	SK 985286	W. Glen at Bitchfield u/s br., d/s pipe
WBC	SK 987261	W. Glen, B1176, Burton Coggles G.S.
WCG	SK 995248	W. Glen at Corby Glen u/s rd. br.
WSW	SK 998233	W. Glen at Swayfield Rd., d/s Corby Glen STW
WCR	TF 011199	W. Glen at Creeton
WLU	TF 019177	W. Glen at Little Bytham, u/s confl., d/s fd. & ft. br.
WLD	TF 019177	W. Glen at Little Bytham, d/s confl., d/s rd.br.
WEU	TF 038137	W. Glen at Essendine u/s & d/s Carby-Essendine rd.br.
WED	TF 050127	W. Glen at Essendine
WBL	TF 068112	Glen at Banthorpe Lodge, d/s br.
GKB	TF 107148	Glen at Kates Bridge G.S., d/s weir & br.
GTF	TF 129170	Glen at Thurlby Fen, end of Fen rd.
GTE	TF 156190	Glen at Tongue end, d/s confl. with Bourne Eau
GGG	TF 173225	Glen at Guthram Gowl
GPK	TF 235260	Glen at Pinchbeck, u/s br. by sluice
GSF	TF 251281	Glen at Surfleet, A16
GSS	TF 277291	Glen at Surfleet Seas End, d/s Blue Gowl Drain
TCH	SK 986196	Tham at Cottage Hill, u/s bridge, d/s track
TCU	SK 992187	Tham at Castle Bytham, d/s ft.br. to motte & bailey, u/s village pond
TCD	SK 993183	Tham at Castle Bytham, d/s fd. in village, d/s pond
TGU	TF 003181	Tham at Glebe Fm., u/s STW
TGD	TF 008181	Tham, d/s STW
TLU	TF 012178	Tham at Little Bytham
TLD	TF 015179	Tham at Little Bytham
BBU	TF 102198	Bourne Eau at Bourne, u/s confl. Car Dyke; d/s rd. br.
BBD	TF 105198	Bourne Eau at Bourne, d/s rd.br., sluice +weir
BRL	TF 121199	Bourne Eau, d/s railway br., d/s pylons
BTU	TF 148190	Bourne Eau at Tongue End, end of farm track
BDP	TF 153188	Bourne Eau at Tongue End, u/s Glen confl.

abbreviations: u/s = upstream rd.br. = road bridge confl. = confluence
 d/s = downstre: ft.br. = foot bridge
 br. = bridge fd. = ford

Table B2. Dates of samples from River Glen sites

SITE	1976	1977	1978	1979	1980	1981	1982	1983	1984	1985	1986	1987	1988	1989	1990	1991
ELN													1	2	2	1
EHW													1			
EBB		1	1	1			1	2					1			
EEL													1			
EEH	1		1	1				1	1	1		1	1	2	3	1
EGB							1							1	1	
EPH		1					1						1			
EMT													1			
EMU													2			
EMD													2			
EMG	1	1					1	1					2			
EBR			1	1				2	1	1	1	2	1	2	3	1
ET1.1								1								
ET1.4								1					1			
WBP													1			
WBF			1	2				1					1			
WBC													1	2	3	1
WCG			1	2			1	1					1	1		
WSW													1	1		
WCR														1		
WLU										1						
WLD	1	1		2				1	1	2	1	1	2	2	3	1
WEU	1	1	1	1			1	2								
WED													1			
WBL	1		1	2				2	1	1	1	1	1	2	2	1
GKB	2	1	1											1	3	1
GTF	1	1	1	1				1		1	1	1				
GTE			1	2			1	1		1	1	1		2	3	1
GGG	2	1	1	1			1	1								
GPK	2	1	1	2			1	1								
GSF														2	3	1
GSS	2	1	1	1				1	1	1	1	1		2	3	1
TCH							1		1			1				
TCU									1			1				
TCD							1		1			1				
TGU							1		1	1					1	1
TGD										1						
TLU			1	1				1								
TLD		1					1		1	1		1	1	2	3	1
BBU	1		1	1												
BBD		1		1			1	1								
BRL				1			1	2								
BTU														2		
BTD															2	1

4 METHODS

4.1 Sampling

All the samples were collected using a standard 1.5-minute kick-and-sweep technique with a hand net. All micro-habitats within a site were sampled in order to maximise the range of taxa recorded, except where in very deep sites sampling was restricted to the margins. Samples were sorted "live", either in the field or laboratory, and taxa identified and recorded on a standard recording form.

4.2 Standardisation of taxa lists

Information from the recording forms was entered in full onto a Microsoft EXCEL spreadsheet, using separate spreadsheets for different formats of recording sheets. These were then edited together to form a composite file on which every taxon name used was included. Due to differences between recording sheets the same taxa may have been referred to under different names, eg. *Baetis vernus* could have been recorded variously under Baetidae, Other Baetidae, *Baetis* sp., *Baetis* indet., *Baetis vernus/tenax* or *Baetis vernus*. For the purposes of comparing sites and dates it was therefore necessary to group such records to the appropriate taxonomic level: in this example, Baetidae. A "grouped" file was therefore produced for all sites. In most instances the groupings were to family level, and a further file was constructed which recorded all taxa to family level or higher.

An indication of abundance class was given on a minority of records, and for the purposes of standardisation ignored: presence / absence data were used in all the analyses.

4.3 Biotic indices

Using the standard formulae (Extence and Ferguson, 1989) the following biotic indices were calculated for each record, using the family data file:

Biological Monitoring Working Party Score (BMWP)
number of BMWP scoring taxa
Average Score Per Taxon (ASPT)
Lincoln Quality Index (LQI)

Calculation of the LQI required information on the type of habitat sampled: Riffle, run or pool. Such information was provided on most record sheets; where it was absent the most likely habitat type was assumed.

4.4 Sample classification using TWINSpan

Two-Way Indicator Species Analysis (Hill, 1979) was used to classify samples on the basis of the family level data into groups from different regions, sites, dates and seasons sharing similar invertebrate communities.

TWINSpan uses a method of "refined ordination" to classify sites. A hierarchical pattern of divisions is produced, and with each division "preferential" taxa identified which are significantly more frequent in one sub-group or the other. Subsequently an "indicator ordination" is performed which identifies taxa on the basis of the presence or absence of which alone the same classification could be produced. For this reason the "indicator" taxa chosen are frequently the rarer taxa associated with each group, rather than more common taxa with large differences in frequency, even though these may be more ecologically significant. For this reason the discussion focuses on the preferential taxa of the main TWINSpan ordination,

although indicator taxa are shown in the dendrograms. In some instances "key" taxa are referred to - these are the most strongly preferential taxa whose presence or absence is believed to be particularly significant in group-separation.

It should be noted that TWINSpan places greater weight on similarities in taxa present than on the absence of taxa, with the result that samples with very reduced communities tend to be grouped with others of which their faunas are subsets, at least in the earlier divisions. For this reason the TWINSpan classifications should be viewed in conjunction with the correspondence analysis ordinations which are more effective at identifying sites with an absence of commoner taxa.

TWINSpan sub-divides groups of samples repeatedly until the requested level of divisions has been reached. The lowest level of division which still represents ecologically meaningful end-groups, ie. there is an ecological basis for the differences in taxa between groups rather than chance sampling effects, has to be subjectively assessed. In making this assessment reference was made to the preferential taxa associated with the divisions, and to the correspondence analysis (Section 4.5) which is extremely useful in determining the actual amount of faunal variation between end-groups.

4.5 Sample ordination using CANOCO

Ordination was performed using the correspondence analysis option of the CANOCO package (Ter Braak, 1988). Correspondence analysis allows samples to be grouped according to similarities in their taxonomic composition. As such it is a useful tool both on its own for grouping samples and for confirming the groupings produced by complementary analyses such as TWINSpan. Correspondence analysis was used in preference to any form of detrended correspondence analysis (eg. DECORANA). In this particular (Glen) data set where major differences in groups of samples were being investigated, rather than small between-sample changes in relation to linear environmental variables, the small benefits of detrending in eliminating some mathematical errors were considered to be outweighed by the problems that exist with the detrending process which invalidate the interpretation of the distance between points in the ordination diagram.

5 ANALYSIS AND DISCUSSION

5.1 Taxonomic diversity

From the combined list of taxa, in which the grouping of taxa under a family or higher group name was kept to the minimum necessary to standardise across the 15 year period, a total of 112 types were recorded. Lists for all the samples are given in Appendix 1.

5.2 Biological quality

BMWP scores, ASPTs and numbers of scoring taxa for all samples are given in Appendix 2.

Mean site scores for BMWP, no. scoring taxa and ASPT are shown in Figure B2. One standard deviation error bars give an indication of the range of scores at each site over the 15 year period, although these have little true significance due to variable sample size. It is apparent that the lower West Glen and the River Tham have the most diverse and sensitive communities as indicated by all three scores. High BMWP scores are a result of both the number and "quality" of taxa. Rather surprisingly the other reaches have very similar score ranges despite the diverse nature of the habitats, with the smaller E. Glen riffle sites scoring similarly to the slow flowing lower Glen and Bourne Eau reaches, possibly due to water quality problems. Bourne Eau is notable in the steady increase in scores in progressively more downstream sites.

When reorganised and plotted against distance from the confluence with the River Welland there is a predictable relationship between scores and distance (Figure B3), with the highest scores occurring in the middle regions. The top of the tributaries have reduced communities due to their intermittent nature, and the worsening effect this has on organic pollution problems, whilst the lowest sites are deep, slow flowing and have rather specialised faunas. The middle reaches show the greatest diversity, with more predictable, moderate, flows and better water quality. LQI shows improved scores in the lower reaches due to the compensation for "restricted riffles".

5.3 Ordination of all samples based upon the family data

The ordination diagram constructed from the results of a correspondence analysis of the family data is shown in Figure B4. Axes one and two of the ordination accounted for the majority of the variation in the data and are therefore the only two used. Two points are omitted from the diagram: two samples from the West Glen at Corby Glen which possessed very few taxa and plotted very distantly from the rest of the points. The remainder of the sites plotted within a relatively small range on axis one of the ordination, indicating the overall similarity of their faunas. However, certain between-reach differences can be seen.

There are two noticeable trends: Along axis two, the River Tham and reach A of the (upper) West Glen overlap considerably towards the top of the diagram, with the other reaches plotting lower down. The lower West Glen (reaches B, C and D) plot slightly closer to the middle than the other sites but with no clear separation. Secondly, along axis one, the sites separated from the Tham and reach A on axis two (which overlap on axis one) are further separated into the main Glen, Bourne Eau and East Glen in a left to right direction. These trends are summarised in the inset of Figure B4. The two trends seem logical: The West Glen and Tham, being physically close and probably of similar habitat type, share a similar fauna. The lower Glen and Bourne Eau separate from the other sites being of a very different physical nature, but also possess differences from each other. The East Glen is surprisingly different from the West Glen - without knowledge of the physical nature of this reach it is difficult to explain, but pollution and/or drought may be factors. Reaches B, C and D which fall in the centre of the ordination probably possess taxa common to all the other reaches.

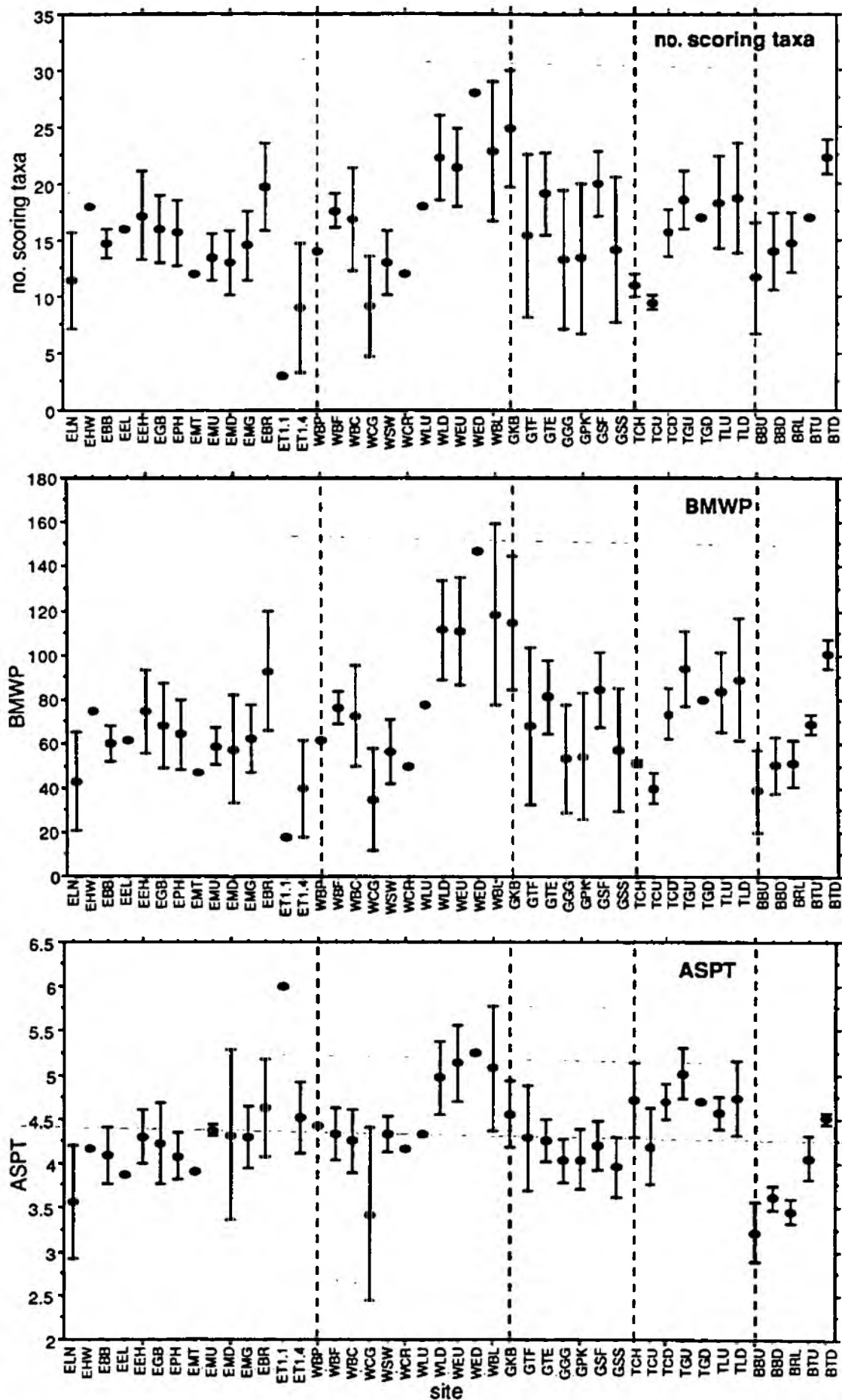


Figure B2. Mean number of scoring taxa, BMWP and ASPT for each site, with 1 S.D. error bars.

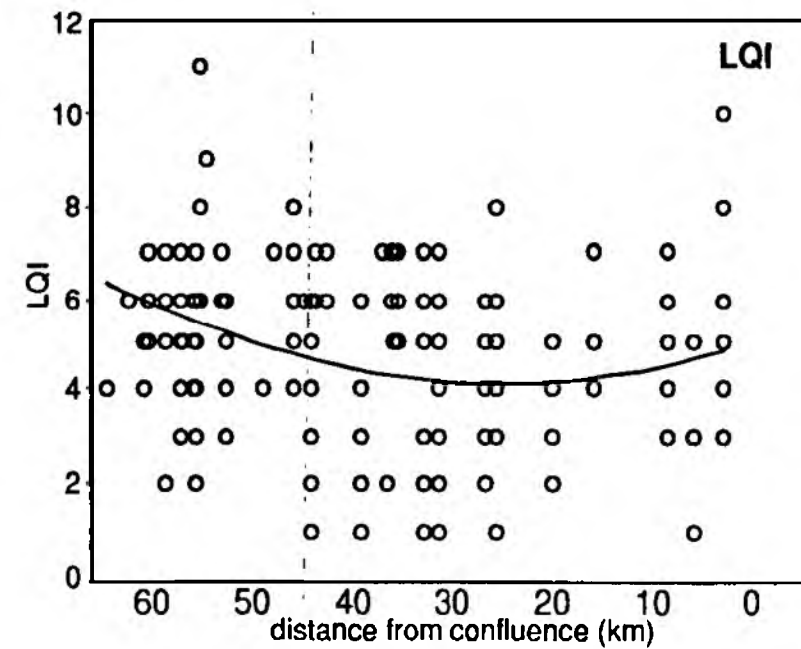
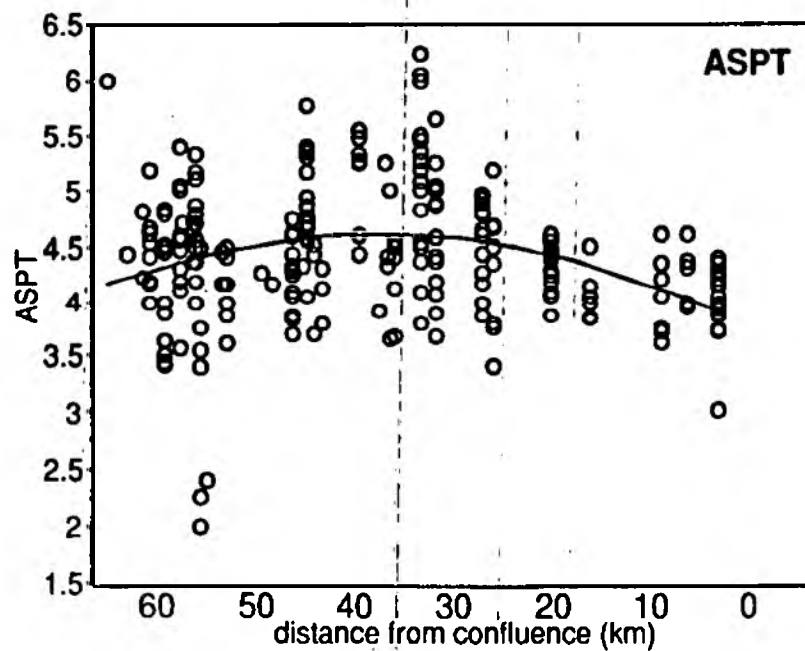
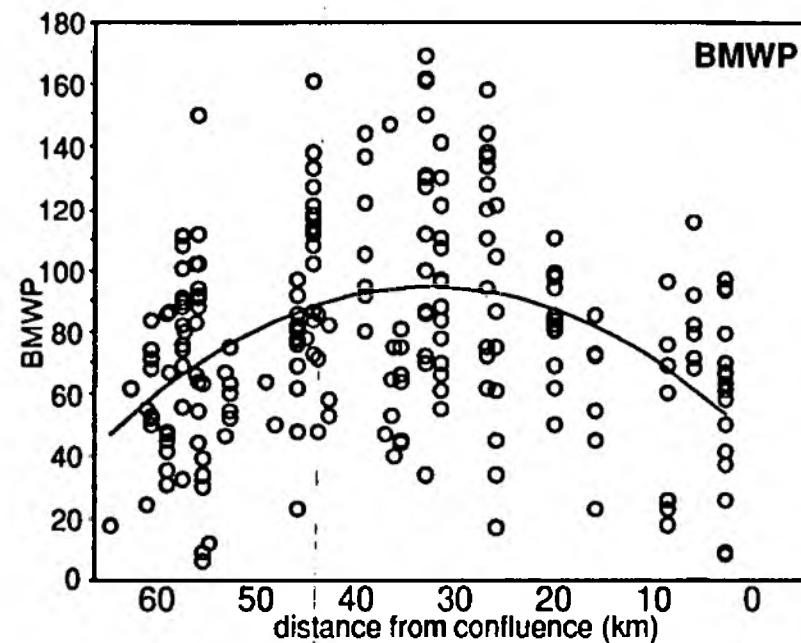
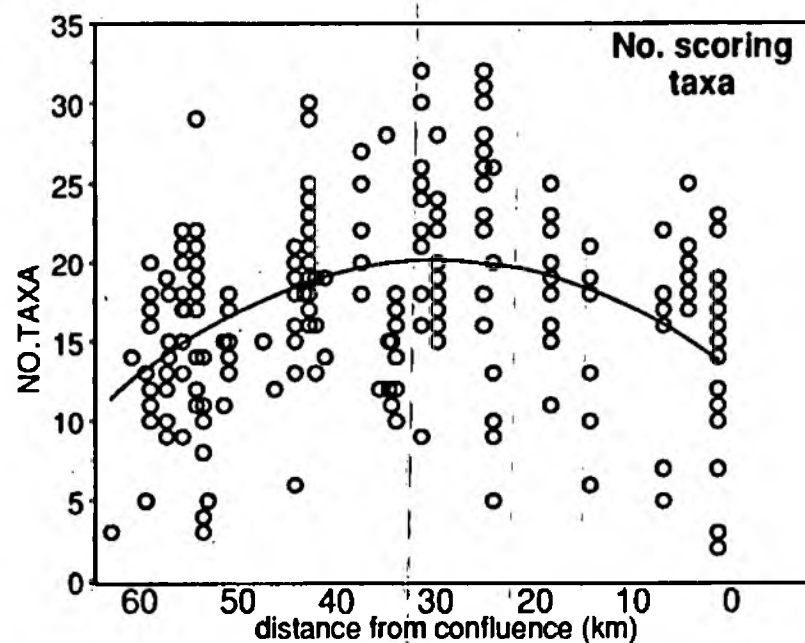


Figure B3. Number of scoring taxa, BMWP, ASPT and LQI for all samples, against distance from source

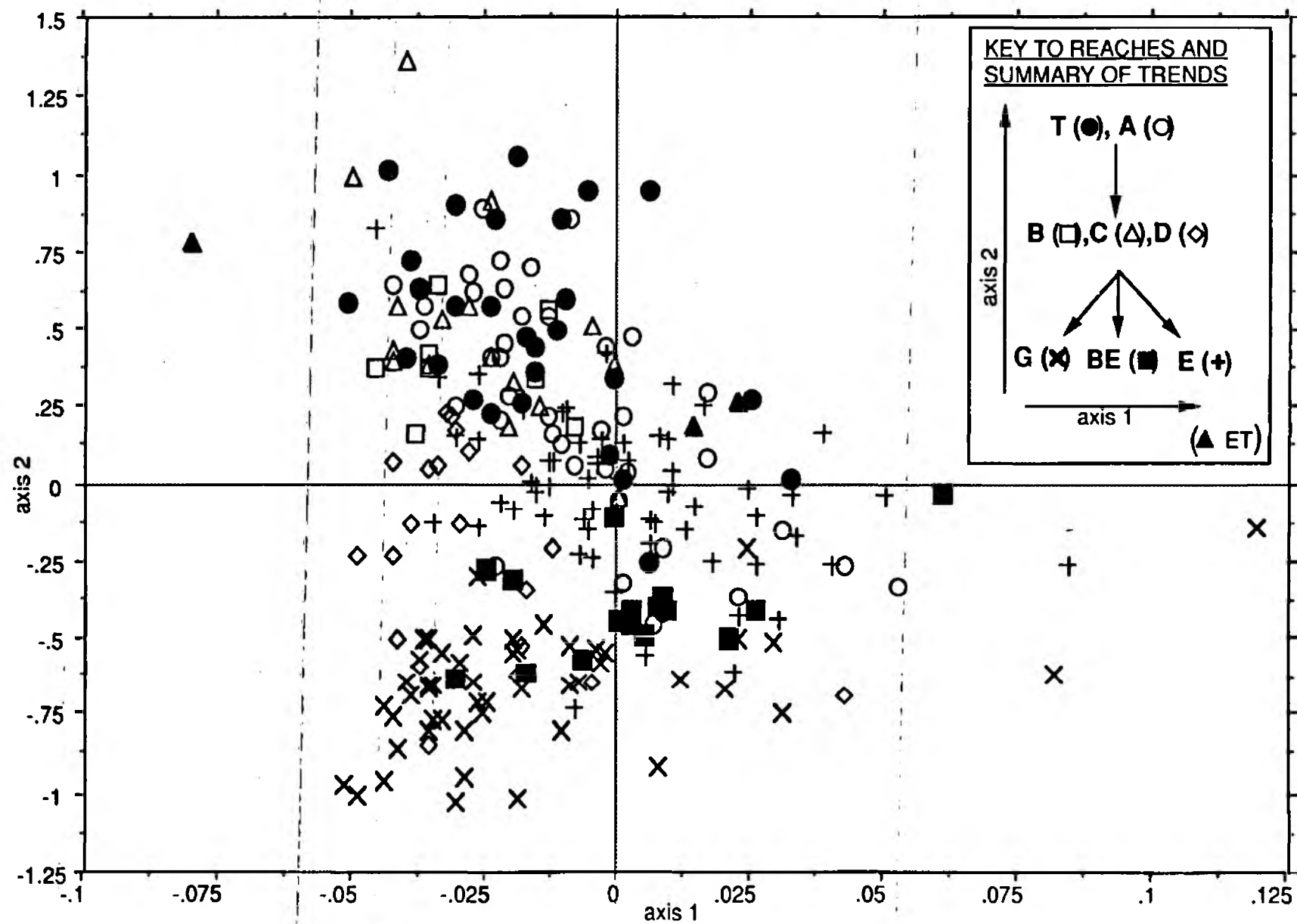


Figure B4. Correspondence analysis of River Glen family data

5.4 Classification of all samples based upon the family data

A classification of the whole data set was made using TWINSpan. Figure B5 shows the dendrogram produced from the output, with taxa derived from the indicator ordination shown at each division. End group numbering is for convenient reference and is not the same as the TWINSpan division numbering. Figure B6 shows the diagonal samples-and-taxa matrix with groups of sites numbered at the bottom using the same notation as in the dendrogram (Figure B5). Preferential taxa for each division are listed in Appendix 3.1. A summary of the sites, physical and biotic characteristics and key taxa associated with the eight significant divisions of the classification is illustrated in Figure B7 (emboldened site codes indicate the majority of samples from this site falling in this group; plain text indicates just two or more samples). "Key taxa" are those showing strong preference for the group indicated.

5.4.1 The first major division

The first major division separates (with some exceptions) the main river Glen samples, Bourne Eau and the bulk of the East Glen samples from the West Glen and Tham samples. Some sites have samples in both groups, notably GKB and EBR. These two are physically on the borderline between the two groups and so this seems logical. Taxa associated with this division suggest that the separation is based on physical rather than water quality differences: The taxa found significantly more frequently in the "East and lower Glen" group are the Ostracoda, Cladocera, Coenagriidae and Valvatidae; while those preferring the "West Glen and Tham" group include a number of Ephemeroptera: Ephemeridae, Ephemerellidae, and Leptophlebiidae; several Trichoptera: Rhyacophilidae, Polycentropodidae, Hydropsychidae, Hydroptilidae, Limnephilidae, and Goeridae; and also the Elmidae, Tipulidae, Ancyliidae, Simuliidae and Dendrocoelidae. In Figure B6 these taxa are arrayed on the top left (East and lower Glen group) and bottom right (West Glen and Tham group). It is apparent from this matrix that by far the majority of taxa occur in both groups, i.e. the obvious site distinctions are based on more subtle differences in frequencies of rather than the presence or absence of large groups of taxa. This might well be expected as all sites are within one catchment with many physical similarities. Also, recolonisation of any site is likely to be from others in the same catchment, which may significantly influence faunal composition of that site.

5.4.2 Subdivision of the main and East Glen sites: groups 1-16

The second TWINSpan division separates end-groups 1-8 from 9-16, within the East and lower Glen group. Groups 1-8 encompass most of the lower main Glen samples and all but two of the Bourne Eau samples. Groups 9-16 contain most of the East Glen samples but also a mixture of others from the main and West Glen and Tham. Only one family - the Hydropsychidae - is given as preferential for the groups 1-8, while those taxa strongly preferential for the groups 9-16 include the Piscicolidae, Cladocera, Coenagriidae, Hydroptilidae, Valvatidae and Physidae. The preferential taxa probably reflect the physical difference between the smaller, faster-flowing nature of the sites in groups 9-16 as opposed to the slower-flowing lowland main Glen sites.

The subdivision of groups 1-8 into groups 1-4 and 5-8 does not appear to be site-based but identifies a group samples (in end groups 1-4) which possess a number of taxa including the Gerridae, Notonectidae and Unionidae at relatively high frequency but the Cladocera, Copepoda and Hydroptilidae less frequently. The subsequent division identifies groups 1 and 2 as being less diverse than groups 3 and 4, lacking a number of taxa including a number of Coleopteran families. Groups 5-8 are separated fairly clearly into groups 5-6 and 7-8 based upon the frequency of taxa which again suggest differences in flow conditions and/or possibly macrophyte development: groups 5 and 6, which mainly consist of samples from the lower main Glen sites, have higher frequencies of Coenagriidae, Caenidae and Corixidae; whereas groups 7 and 8, which include samples from Bourne Eau and the two sites close to the confluence of the East and main Glen - EBR and GKB - possess greater frequencies of taxa including Polycentropodidae and Simuliidae.

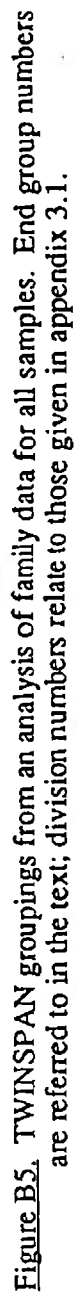


Figure B6. TWINSPAN samples and taxa matrix. End groups numbers are as in figure 5.

[illegible]

TWINSpan division	8	9	10	11	12	13	14	15
TWINSpan end groups	1-4	5-8	9-12	13-16	17-19	20-23	24-27	28-31
sites	BTD GTE GTF GTE	GTF GTE GGG GPK GSS GSF BBU BBD BRL BTU GKB EBR	TCH GTF GSS GPK WCG	ELN EBB EEH EGB EPH EMG EBR WBF WBC WCG TCU GKB WEU WBL	GKB	EBR WEU WBL GKB WLD	WBF EBR EEH WLD WBL	WLD TCD TGU TLD TLU EBR WBL
group description	lowland, slow-flowing, abundant macrophytes, good water quality, rich fauna	lowland, slow-flowing, macrophytes, reasonable water quality and fauna	intermittent, or poor water quality, very reduced fauna	poor habitat, poor fauna	good, diverse habitat, rich fauna	moderate flow, some macrophytes, moderate water quality, intermediate fauna	moderate flow, clean substrates, rich fauna	moderate/fast flow, stable, clean substrates, macrophytes, rich fauna
key taxa	Leptoceridae, Hydrometridae, Gerridae, Notonectidae, Unionidae	Cladocera, Copepoda, Hydroptilidae	Cladocera, Copepoda	Glossiphoniidae, Erpobdellidae, Asellidae, Slafidae, Caenidae, Elminthidae, Hydrobiidae, Planorbidae	Coenagriidae, Valvatidae, Helodidae, Psychodidae	Piscicolidae, Copepoda, Ephemeridae, Polycentropodidae, Psychomyidae, Gyrinidae		Dendrocoelidae, Piscicolidae, Rhyacophilidae, Hydroptilidae, Ancylidae, Goeridae, Glossosomatidae
	Piscicolidae, Cladocera, Coenagriidae, Hydroptilidae, Valvatidae, Physidae				Ephemerellidae, Notonectidae Physidae, Sericostomatidae, Gyrinidae		Rhyacophilidae, Ancytidae	
	Ostracoda, Cladocera, Coenagriidae, Valvatidae				Leptophlebiidae, Ephemeridae, Ephemerellidae, Rhyacophiliidae, Hydropsychidae, Limnephilidae, Goeridae, Ancylidae, Simuliidae, Polycentropodidae			

Figure B7. Summary of site groupings based upon TWINSpan classification of samples. Key taxa are those showing a strong preference for the group indicated.

Groups 9-16 are subdivided into 9-12 and 13-16 on the basis of a range of taxa absent or at very low frequency in the former group. This particularly apparent in the samples - and - taxa matrix (fig.6), where many of the common taxa located in the middle of the diagram are visibly less frequent in these groups. In particular the Hydropsychidae and Leptoceridae are absent from this group altogether. Samples comprising groups 9-12 are drawn from a wide range of sites, and appear to represent occasions when the fauna at those sites has been severely impacted (drought or pollution?). Many of these samples are from the late 70's period. Further separation of these groups (9-12) appears to reflect the severity of the faunal reduction, with group 9 the worst affected. Groups 13-16 subdivide into groups 13-14: a range of samples from the upper East Glen sites, the lower East Glen sites below Manthorpe STW and a few "odd" sites from the West and main Glen; and groups 15-16: The larger East Glen sites, a significant number of upper West Glen sites and a few from the Tham and lower Glen. All of these samples appear to be from "poor quality" sites or from dates when "better" sites were under stress (eg. WEU, WLD and WBL in 1976). The taxa separating groups 13-14 from 15-16 include the Simuliidae and Physidae which are more frequent in the former group and the Coenagriidae, Sialidae, Hydropsychidae, Leptoceridae and Sphaeriidae in the latter. It could be that the former group is the more severely impacted than the latter.

5.4.3 Subdivision of the West Glen and Tham sites: groups 17-31

Within this set of groups the first subdivision into groups 17-23 and 24-31 separates the majority of the lower West Glen and "intermediate" sites (EBR and GKB) from the upper West Glen and Tham sites. The taxa associated with the division reflect this: the Rhyacophilidae and Ancyliidae which prefer faster flow and more stable substrates show a strong preference for the West Glen and Tham sites, whereas taxa such as the Ephemeridae, Polycentropodidae, Gyrinidae, Corixidae, Notonectidae and Sericostomatidae which tolerate or prefer slower flowing conditions are more frequent in the lower West and East Glens and Kates Bridge.

Within groups 17-23 the ecological basis of further subdivisions are difficult to interpret, these being based on substitutions of taxa particularly from the Trichoptera, Coleoptera and Diptera.

Groups 24-31 subdivide into groups 24-27 and 28-31 mainly on the basis of a collection of more "sensitive" taxa which are more frequent in the latter group. Sites dominating this group are from the Tham and middle West Glen, and are those which scored highest using the various biotic indices. Taxa preferring this group include the Piscicolidae, Rhyacophilidae, Hydroptilidae, Goeridae, Glossosomatidae and Ancyliidae. Taxa preferring the other group (24-27) include the Asellidae, Sialidae, Leptoceridae and Planorbiidae. Further subdivisions within groups 24-27 and 28-31 are again very difficult to interpret, being based mainly on the substitution of a few taxa at each division.

5.4.4 Definition of reach communities

The TWINSpan classification produced 31 end groups between which both meaningful between-site and temporal (between-sample, within site) differences and some chance between-sample differences could be discerned (Sections 5.4.2 and 5.4.3). At the higher level of division eight major groupings based mainly upon reach-scale differences were apparent. From these it is possible to define three major reach-types:

- "lowland" - divisions 8 and 9 (end groups 1-8) contain most of the lowland-type sites, division 8 containing the most faunistically rich samples.
- "upland" - division 15 (end groups 28-31) contain the upland-type sites.
- "intermediate" - divisions 12, 13 and 14 (end groups 17-27) contain sites possessing taxa in common with both the "upland" and "lowland" sites, the three divisions representing slightly different communities, all faunistically rich.

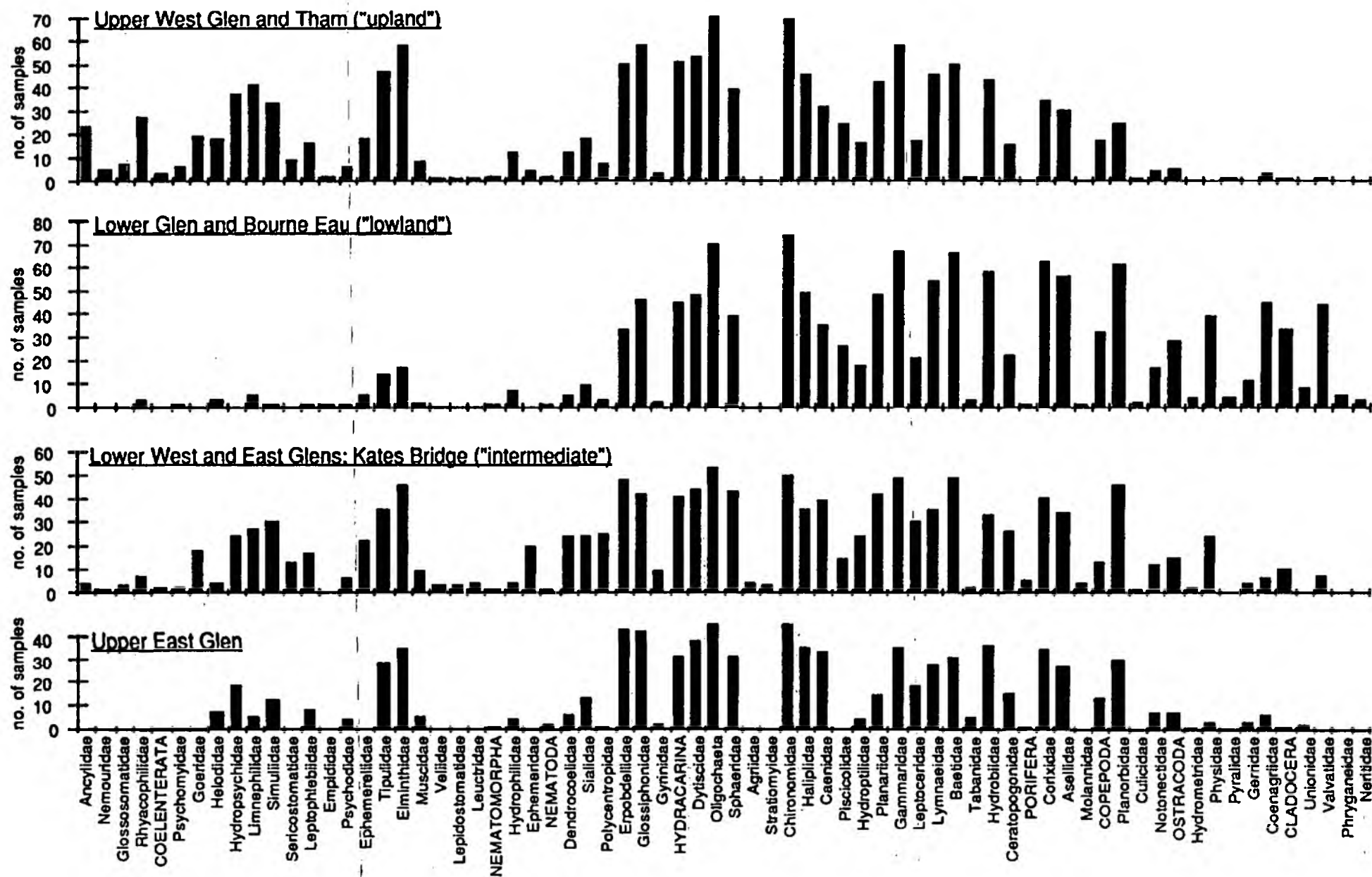


Figure B8. Frequencies of taxa within reach-types.

"UPLAND"	"INTERMEDIATE"	"LOWLAND"
Chironomidae	Chironomidae	Chironomidae
Oligochaeta	Oligochaeta	Oligochaeta
Gammaridae	Gammaridae	Gammaridae
Baetidae	Baetidae	Baetidae
Corixidae	Corixidae	Corixidae
Planorbidae	Planorbidae	Planorbidae
Hydrobiidae	Hydrobiidae	Hydrobiidae
Asellidae	Asellidae	Asellidae
Lymnaeidae	Lymnaeidae	Lymnaeidae
Halplidae	Halplidae	Halplidae
Dytiscidae	Dytiscidae	Dytiscidae
Planariidae	Planariidae	Planariidae
Glossiphonidae	Glossiphonidae	Glossiphonidae
HYDRACARINA	HYDRACARINA	HYDRACARINA
Coenagriidae	Coenagriidae	Coenagriidae
Sphaeridae	Sphaeridae	Sphaeridae
Caenidae	Caenidae	Caenidae
Erpobdellidae	Erpobdellidae	Erpobdellidae
COPEPODA	COPEPODA	COPEPODA
OSTRACODA	OSTRACODA	OSTRACODA
Piscicolidae	Piscicolidae	Piscicolidae
Ceratopogonidae	Ceratopogonidae	Ceratopogonidae
Leptoceridae	Leptoceridae	Leptoceridae
Hydroptilidae	Hydroptilidae	Hydroptilidae
Elminthidae	Elminthidae	Elminthidae
Notonectidae	Notonectidae	Notonectidae
Tipulidae	Tipulidae	Tipulidae
Sialidae	Sialidae	Sialidae
Hydrophilidae	Hydrophilidae	Hydrophilidae
Limnephilidae	Limnephilidae	Limnephilidae
Ephemerellidae	Ephemerellidae	Ephemerellidae
Dendrocoelidae	Dendrocoelidae	Dendrocoelidae
Rhyacophilidae	Rhyacophilidae	Rhyacophilidae
Helodidae	Helodidae	Helodidae
Polycentropidae	Polycentropidae	Polycentropidae
Tabanidae	Tabanidae	Tabanidae
Muscidae	Muscidae	Muscidae
Gyrinidae	Gyrinidae	Gyrinidae
Hydropsychidae	Hydropsychidae	
Simuliidae	Simuliidae	
Ancylidae	Ancylidae	
Goeridae	Goeridae	
Leptophlebiidae	Leptophlebiidae	
Sericostomatidae	Sericostomatidae	
Glossosomatidae	Glossosomatidae	
Psychodidae	Psychodidae	
Psychomyidae	Psychomyidae	
Ephemeridae	Ephemeridae	
COELENTERATA	COELENTERATA	
	Physidae	Physidae
	CLADOCERA	CLADOCERA
	Valvatidae	Valvatidae
	Gerridae	Gerridae
	Hydrometridae	Hydrometridae
Nemouridae		Unionidae
NEMATODA		Phryganeidae
NEMATOMORPHA		Pyrallidae
Empididae		Neritidae
		Culicidae

Table B3. Taxa lists for the three reach-types.
Taxa occurring in less than 1% of samples in the reach omitted.

Divisions 10 and 11 (end groups 9-16), which include the East Glen sites, represent samples with impoverished faunas rather than any distinct reach community.

Figure B8 displays histograms showing the frequency of occurrence of taxa within each reach-type. The taxa have been ordered according to their proportional occurrence in the two major reach-types: Taxa occurring more frequently in the "upland" sites are ranged to the left; those more frequent in the "lowland" sites ranged to the right. It can be seen that although there are a large number of ubiquitous taxa, the majority of taxa do show a preference for one or other reach-type. The "intermediate" sites possess taxa of both community-types, plus two taxa unique to these sites (Agriidae and Stratiomyidae - at very low frequency so probably chance occurrences). Interestingly, the upper East Glen sites still record taxa from both reach-types as well as the ubiquitous ones. This suggests that the poorer faunas are due to a reduced habitat diversity at individual sites rather than a general degradation at all sites.

On the basis of the TWINSPAN divisions alone it is possible to predict the potential "best" community which each of the three major reach-types could support: For the "lowland" sites the combination of taxa associated with division 8 represents the most diverse community of this reach-type. The "upland" sites potentially possess those taxa recorded in the division 15 samples. The "intermediate" sites which fall into divisions 12-14 may possess the larger range of taxa defined by these divisions, although sites within this reach type are more variable, some tending to the "upland" and some to the "lowland" end of this intermediate spectrum. Table BB3 lists the taxa occurring in the divisions associated with the major reach types, which could potentially be found in any site within that reach. Taxa occurring in less than 1% of samples have been omitted as these may be chance rarities. It is notable that the "specialist" taxa (restricted to one or two reach-types) are not necessarily those having the highest BMWP scores, ie. specialism and sensitivity are not synonymous.

5.5 Long-term changes in biological quality

5.5.1 Changes in scores at the catchment scale.

For the three indices: BMWP, ASPT and number of scoring taxa per sample, significant changes were found over the 15 year period. When all the sample scores for each year were plotted against year of sampling (Figure B9.) it was found that all three scores increased between 1976 and the late 1980's, but then declined in 1989-91. It appears that the number of invertebrate taxa have increased and then declined again, and also that the taxa which have increased and then decreased in frequency are those most sensitive to organic pollution as measured by these indices. It must be remembered, however, that the range of sites surveyed has changed during this period, and it is quite possible that the earliest samples tended to be taken in response to reported pollution incidents, whereas in later years more routine sampling of "unpolluted" sites was taking place. Such a change in sampling strategy may produce the observed increase in biological quality.

5.5.2 Changes in scores at the site and reach scale

A similar analysis of the records for individual sites where at least six samples had been taken over the 1976-91 period confirmed the overall trends found in the whole data set, but also revealed sites with different temporal changes in scores. Figure B10 shows those sites where a significant trend in score over time was seen when scores were regressed against year of sampling, and indicates the type of change - from steady increase, through initial increase followed by a slight decrease; slight increase followed by major decrease, to steady decrease in score. (Sites where the records only covered part of the 1976-91 period are shown overlapping the groups to which they may belong.)

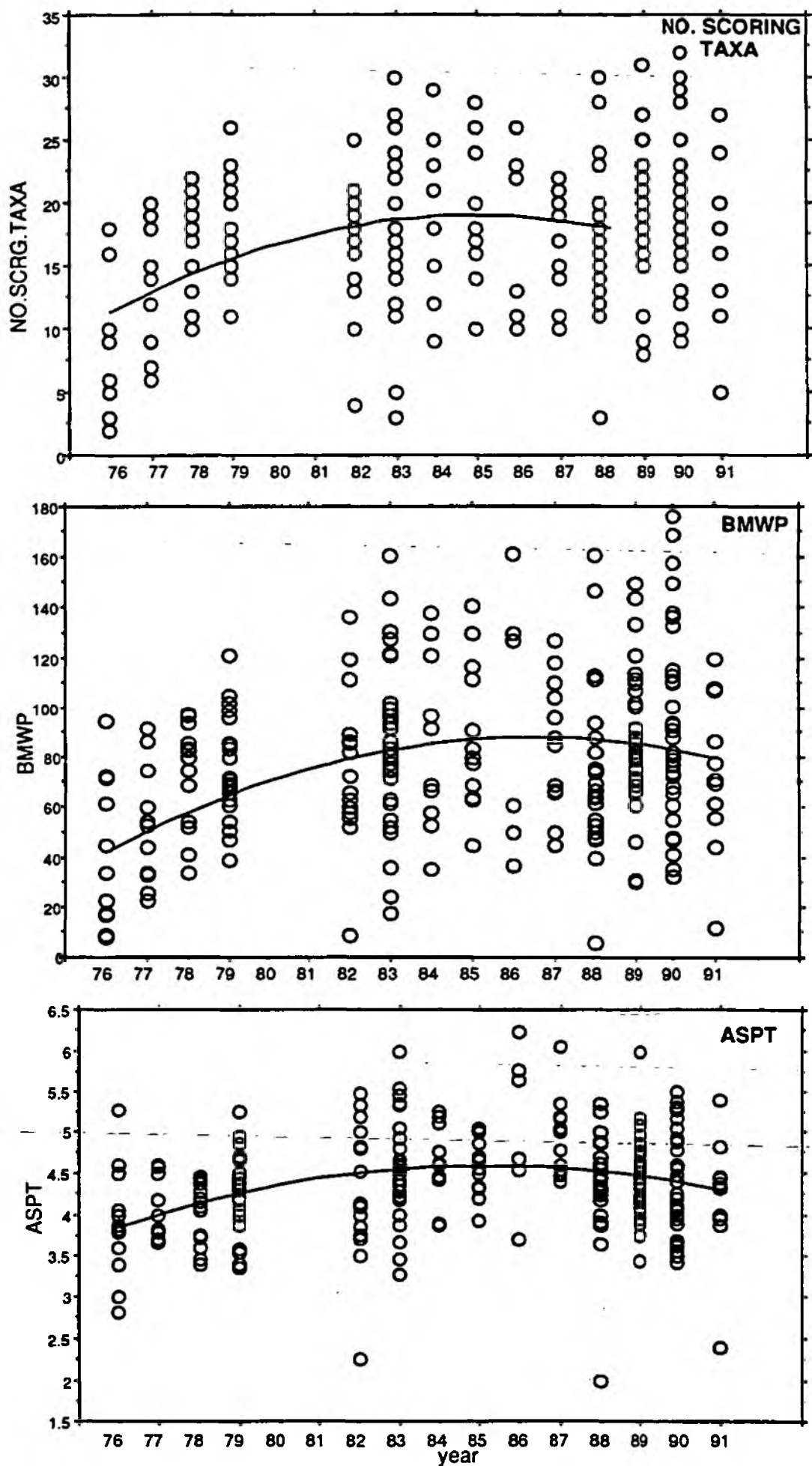


Figure B9. Regressions of number of scoring taxa, BMWP and ASPT against year of sample for all River Glen invertebrate samples.

				
	1976 1991	1976 1991	1976 1991	1976 1991
no. taxa	GKB GSS	WLD WBL ← WEU → EMG GPK	EBR	
BMWP	GSS	EEH WLD WBL GKB ← WEU → GGG	EBR	
ASPT	WLD GSS	EEH WBL GKB ← EBB → GTF	EBR	ELN

Figure B10. Patterns of 1976-1991 changes in number of scoring taxa, BMWP and ASPT for sites where the trend was significant.

The majority of sites followed the second trend, as seen in the total data set, but a few - the lower main river sites and one lower West Glen site - showed steady increases in score over the whole period. In contrast, a number of the East Glen sites showed pronounced decreases in scores. This strong regional pattern suggests an increase in quality of the main river despite major decreases in quality (due to pollution or drought?) in the East Glen.

Not all sites showed the same trend in all three scores, suggesting very different community changes. For instance, the site GSS showed a steady increase in BMWP as a result of increases in both the number and "quality" of taxa (ASPT). Sites EBR and WBL displayed similar changes but in the reverse direction, with recent decreases in BMWP, number and Average Score Per Taxon. GKB, however, showed a rise then fall in BMWP despite continually rising numbers of taxa, due to the ASPT declining. This site may in recent years be experiencing a decrease in water quality, but due to physical (perhaps hydrological?) changes is able to support a greater diversity of the more pollution-tolerant taxa. Site ELN may be responding in a similar way, as the only significant change in score was a steady decrease in ASPT.

5.6 Site case studies

5.6.1 West Glen, upstream of Essendine

Figure B11 displays the results of TWINSpan classification, and ordination using PP correspondence analysis of the sample data for the site WEU. TWINSpan indicator taxa, which could be used alone to produce similar groupings, are shown on the dendrogram and ordination diagram. The dates of samples and their LQI scores are indicated. Preferential taxa associated with each TWINSpan division are listed in Appendix 3.2.

The records for WEU span the period 1976 to 1983. During this time the BMWP scores and number of scoring taxa increased significantly, but without a corresponding increase in ASPT, suggesting that the additional taxa in later years are not significantly more pollution-sensitive. Environmental changes other than an improvement in water quality may have taken place to account for the increased diversity of taxa.

Both classification and ordination of the data identify three distinct groups of samples:

- 1: 9/77
- 2: 10/79; 11/76
- 3: 2/11/83; 3/11/83; 11/82; 1078

Sample 9/77 is most clearly separated from the other samples by both analyses. It is very probable that seasonal factors are responsible for this separation, as taxa absent from this September sample but frequent in the others include the mayflies Leptophlebiidae, Ephemerellidae and Ephemeridae as well as a number of other insect taxa that would be absent or extremely small in this month.

Samples in group 2 - 10/79 and 11/76 possess four taxa absent from group 3: Hydropsychidae, Sialidae, Gyrinidae and Copepoda. The latter three of these in particular can tolerate, or even require, standing water conditions. This together with the fact that taxa absent from group 2 but present in samples later than 1979 include the Simuliidae and Rhyacophilidae - taxa requiring flowing water - suggest that the hydrology of the site may have changed over this period.

Sample 3/11/83 was taken in addition to the sample on 2/11/83 as dredging was in progress on the first occasion. A number of taxa are present in one sample but not the other, but with no clear pattern attributable to the disturbance. It is notable that despite recent dredging these samples score higher than previous ones.

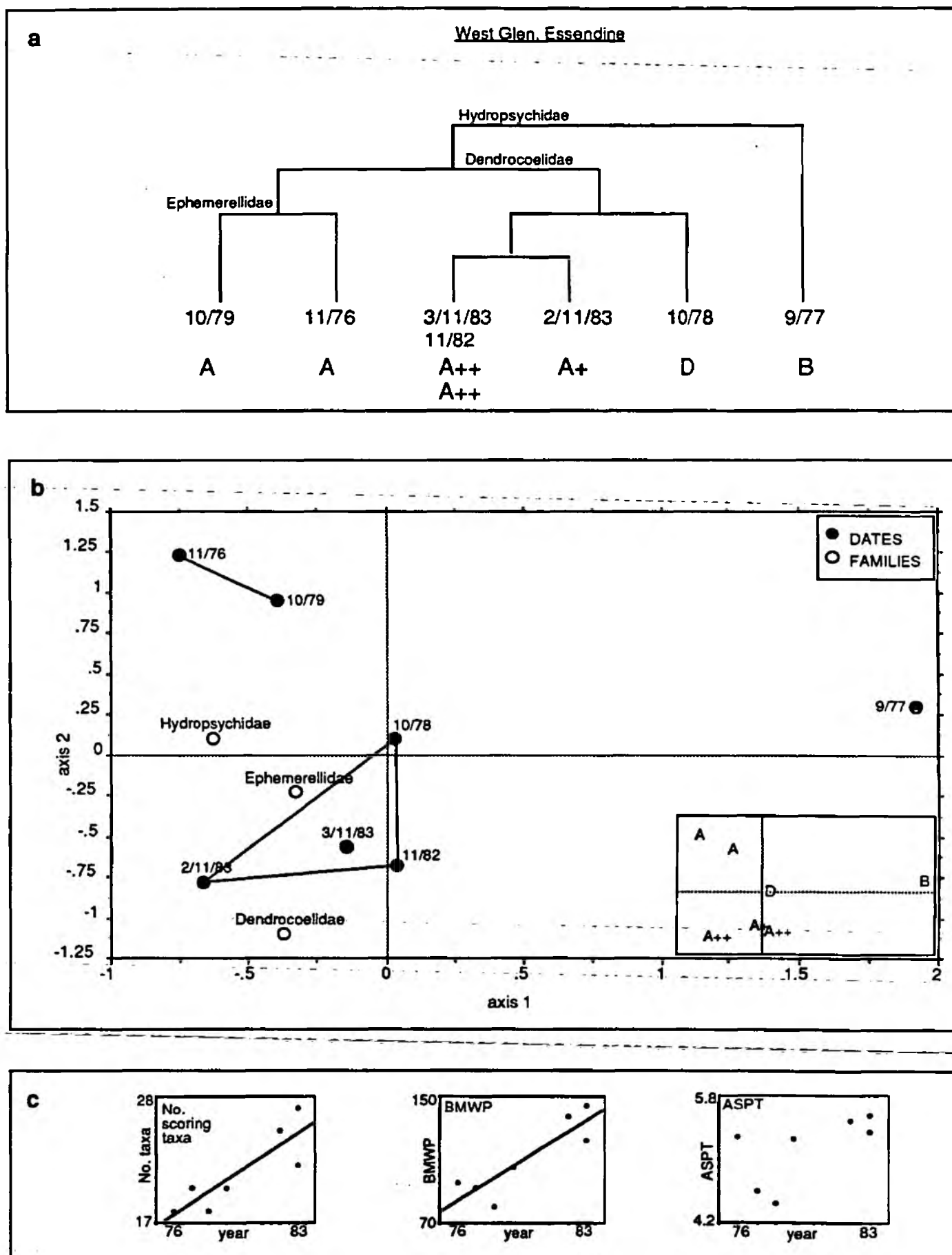


Figure B11. a) TWINSpan classification, b) ordination and c) changes in biotic scores based on family data for the West Glen at Essendine

5.6.2 West Glen at Banthorpe Lodge

Summary diagrams of the analyses are shown in Figure B12, and TWINSpan preferential taxa listed in Appendix 3.2.

Sixteen samples spanning the period 1976-91 were collected from this site. Biological quality varied considerably during this period, with a clear pattern of increasing quality until 1991, when a marked deterioration occurred.

On the basis of the TWINSpan analysis four main groups can be distinguished:

- 1: 7/90, 2/83, 2/90, 11/88, 6/89, 11/89;
- 2: 7/85, 6/86, 6/87, 8/84, 11/83;
- 3: 8/79, 5/79, 5/91, 10/78;
- 4: 11/76.

The first division separates the low-scoring, pre-1980 samples plus the 1991 sample from the rest. The only taxa showing a preference for this group are three families of Diptera: Ceratopogonidae, Stratiomyidae and Muscidae, and Copepoda. All are tolerant of moderate pollution and stagnant conditions.

Within the low scoring groups 3 and 4, the group 4 sample, the earliest in the recorded period, has a markedly reduced fauna and clearly separates in the classification and ordination. Preferential taxa for this sample include the Dytiscidae, Copepoda, Ostracoda, Lymnaeidae and Dendrocoelidae. These taxa, more notable as tolerant of standing water conditions rather than extreme pollution, suggest that the 1976 sample has an unusual and reduced community due to drought conditions rather than pollution, although the two may very well be acting together.

It is notable that the poor quality 1991 sample is associated with the pre-1980 samples of similar low score, both in the classification and the ordination, showing that not only has the fauna changed, but that the new community is very similar to that 12 years earlier. Presumably the taxa gained or replacing other taxa during the 1980's are the ones first lost as conditions deteriorate. It would also suggest that the physical change in habitat is towards similar conditions to those in the late '70's, which could suggest that there has been a recent deterioration in the habitat (due to drought) which has otherwise remained fairly constant.

The invertebrate communities of samples comprising TWINSpan groups 1 and 2 are characterised by a wide variety of relatively pollution-intolerant taxa, especially Ephemeroptera and Trichoptera. The Ephemeropteran families showing preference for this group are those associated with clean, fine substrates and slow to moderate flow velocities (Caenidae, Ephemeridae and Leptophlebiidae). The presence of the Trichopteran families Hydropsychidae and Polycentropodidae, and the molluscs Ancyliidae suggest more stable but again un-silted substrates. The site is probably a clean gravel and sand riffle or run under normal flow conditions, becoming silted or ponded under very low flows or organic pollution. Other taxa associated with weedy margins, such as the Corixidae and Agriidae are also associated with this group.

The samples separated by TWINSpan into groups 1 and 2 appear to be largely overlapping in the ordination, implying that there are relatively insignificant differences in the communities of the two groups. According to the preferential taxa offered by TWINSpan the species characterising group 2 include several associated with weedy conditions - several molluscs (Lymnaeidae, Planorbidae, Hydrobiidae) Corixidae, Agriidae and Asellidae.

5.6.3 River Glen at Kates Bridge

Summary diagrams of the analyses are shown in Figure B13, and TWINSpan preferential taxa listed in Appendix 3.2.

The changes in the invertebrate community at Kates Bridge between 1976 and 1991 are more complex than those shown for sites WEU and WBL (above). Biological quality as measured by BMWP and LQI has risen between 1976 and mid-1990 but then fallen in late-1990-91. The ASPT showed greatest change between the late 1970's and early 1980's with a quite dramatic increase. ASPT then fell in 1991. BMWP and number of scoring taxa showed greater proportional increases between the early 1980's and late 1980's/1990 than did ASPT, suggesting some other habitat improvement than water quality may have been involved to account for the increasing diversity.

The four major TWINSpan groups contain the following samples:

- 1: 4/89, 3/91, 10/89, 2/90;
- 2: 10/78, 11/82, 9/77, 11/83, 6/76, 11/76;
- 3: 11/89, 7/90;
- 4: 10/90.

The first division separates groups 3 and 4 from 1 and 2 on the basis of a large number of taxa confined to, or more frequent in, the former two groups. These include a surprisingly large number of Hemipterans: Notonectidae, Gerridae, Hydrometridae and Veliidae. These indicate slow-flowing conditions (or at least slow-flowing margins), probably with macrophytes. Being relatively mobile these animals may indicate habitats where there has been drought and subsequent recolonisation, but the presence of other taxa such as Ephemeridae, Ephemerellidae and Coenagriidae with one or two year life-cycles makes this less likely.

Within the high-scoring groups 3 and 4, group 4 (10/90) is separated from group 3 on the presence of a number of taxa associated with more lentic conditions: Cladocera, Coenagriidae and a number of Diptera, and the absence of several taxa more associated with flowing water and/or clean substrates eg. Caenidae, Sphaeriidae. However, this distinction is not very clear with group 3 also containing several taxa which suggest slow-flowing, weedy conditions.

There is very little meaningful difference between TWINSpan groups 1 and 2. Only the Sialidae are present in all the group 1 but none of the group 2 samples, and no taxon is present in all group 2 but no group 1 samples. A number of taxa are more frequent in one or other group, but without any obvious pattern related to habitat preference or . The ordination also shows these two groups as largely overlapping, but separated from groups 3 and 4. Only sample - 3/91 - is pulled out in the ordination as significantly different from all the others.

Reference to the ordination diagram reveals a major difference from the situation at Banthorpe Lodge: Whereas at WBL the 1991 sample showed a community with much in common with the 1970's situation, at GKB a different fauna is present in the most recent sample. With falling biological quality the community has not reverted to the earlier type but to a very different assemblage. Sample 3/91 possesses three taxa absent from all other samples: Stratiomyidae, Psychodidae and Helodidae; while lacking two taxa present in all others: Baetidae and Corixidae. It also shares with the 1990 samples several taxa absent from or infrequent in the low-scoring, pre-1980 samples eg. Sialidae, Caenidae and Leptoceridae; while lacking Copepoda which were common pre-1980 but absent in 1990. The combination of some unique features, plus similarities with temporally close samples rather than samples of similar score, leads to the 3/91 sample plotting separately in the ordination. The fact that TWINSpan failed to separate this sample whereas the ordination did may be due to the weight given by the latter method to unique species rather than general similarities. The similarity of the 3/91 sample to the other later samples and contrast with the early samples suggests that a permanent change in habitat has occurred at Kates Bridge.

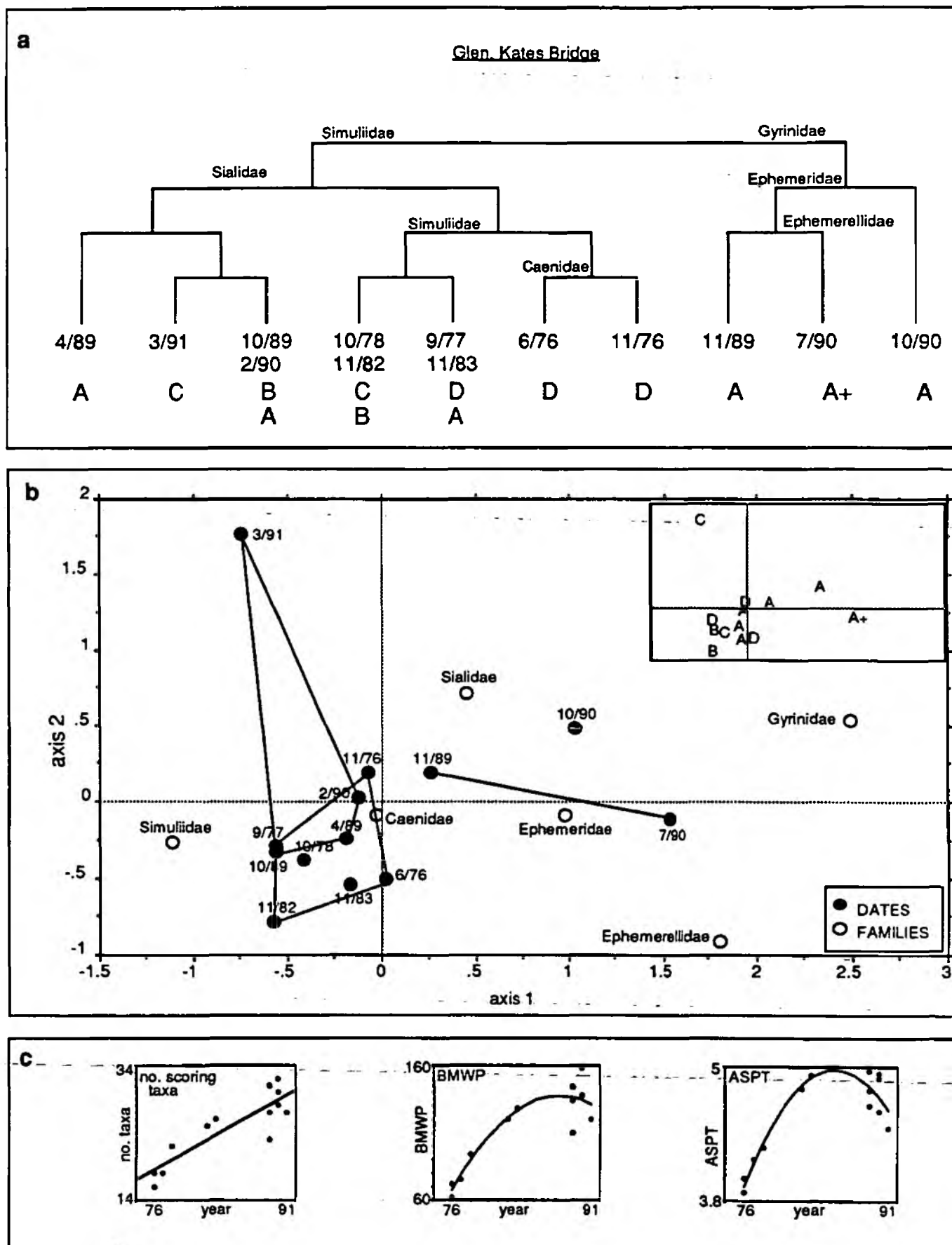


Figure B13. a) TWINSpan classification, b) ordination and c) changes in biotic score based on family data for the River Glen at Kates Bridge.

5.6.4 River Glen at Surfleet Seas End

Summary diagrams of the analyses are shown in Figure B14, and TWINSPAN preferential taxa listed in Appendix 3.2.

This site, the furthest downstream in the system and close to the confluence with the River Welland, has 16 records of invertebrate samples spanning the 1976-91 period. During this time a biological quality as measured by BMWP, number of taxa, ASPT and LQI, has shown a general increase from very poor quality in 1976 to excellent (LQI A) in late 1990, with the 1991 sample being lower (LQI C - good quality) but not as significantly lower as in the previous examples. The range of biological quality from LQI H to A is notably lower than in the more upstream examples (LQI E to A++), as might be expected given the physical nature of this site.

The TWINSPAN classification produces the following four major groupings:

- 1: 8/90, 4/91, 5/89, 3/90;
- 2: 10/90, 8/85, 6/87, 7/84, 10/89;
- 3: 10/78, 6/76, 11/76, 9/77, 8/86;
- 4: 11/83, 10/79.

The first major division (groups 1 and 2 from 3 and 4) is generally supported by the ordination, although this shows a broad trend along axis 1 rather than a distinct split into two groups. This first division appears to combine differences in score and year of sampling: Groups 3 and 4 are generally the earliest and lowest scoring samples, with the exceptions of 10/79 which fits in with the temporal grouping but has a higher score (B) than other samples in the group; and 8/86, which is a more recent sample but shares a low score (D) with the other group members. Two explanations could account for this interdependency of date and score: First, the range of taxa which could possibly exist at the site, in terms of their sensitivity to pollution or other habitat degradation, could be limited by water and habitat quality, and this determines the biological score observed. The actual community present, however, as well as being limited to the range so described, must be a modification of the community previously living at that site, with gains or losses of a few taxa in response to habitat changes. Secondly, changes in one aspect of habitat quality at the site, such as water quality, could produce the observed changes in biological score, but other features of the habitat, eg. hydrology, management, which may have changed more subtly over time, determine the actual community within the limits set by (in this example) water quality. Either of these explanations would account for samples sharing more in common with other samples from a similar time period than those of similar score from a different time period.

The community differences associated with the first TWINSPAN division are largely based on a wide variety of taxa which occur in groups 1 and 2 (containing the later and higher scoring samples) which are absent from the other two groups, including many mollusc families: Unionidae, Sphaeriidae, Valvatidae, Lymnaeidae and Physidae; several beetles: Haliplidae, Dytiscidae and Elmidae; and two relatively pollution-sensitive Trichopteran families: Leptoceridae and Rhyacophilidae. There is a notable lack of Ephemeroptera - only Baetidae and Caenidae are recorded, these being present in all the groups of samples. The only taxa found more frequently in groups 3 and 4 are the Cladocera and Copepoda, suggesting fairly stagnant conditions at the time of these samples. An oddity occurring in samples 6/87 and 8/90 is the family Mysidae, suggesting some saline input at this lowland site.

The division between groups 1 and 2 is based upon a number of taxa, including the Rhyacophilidae and Physidae, which are restricted to or more frequent in group 1. Only a few taxa are limited to group 2, including the Cladocera and Gerridae. Various habitat changes such as weed cutting or dredging, could affect the distribution of many of the taxa characterising these groups, perhaps more so than water quality.

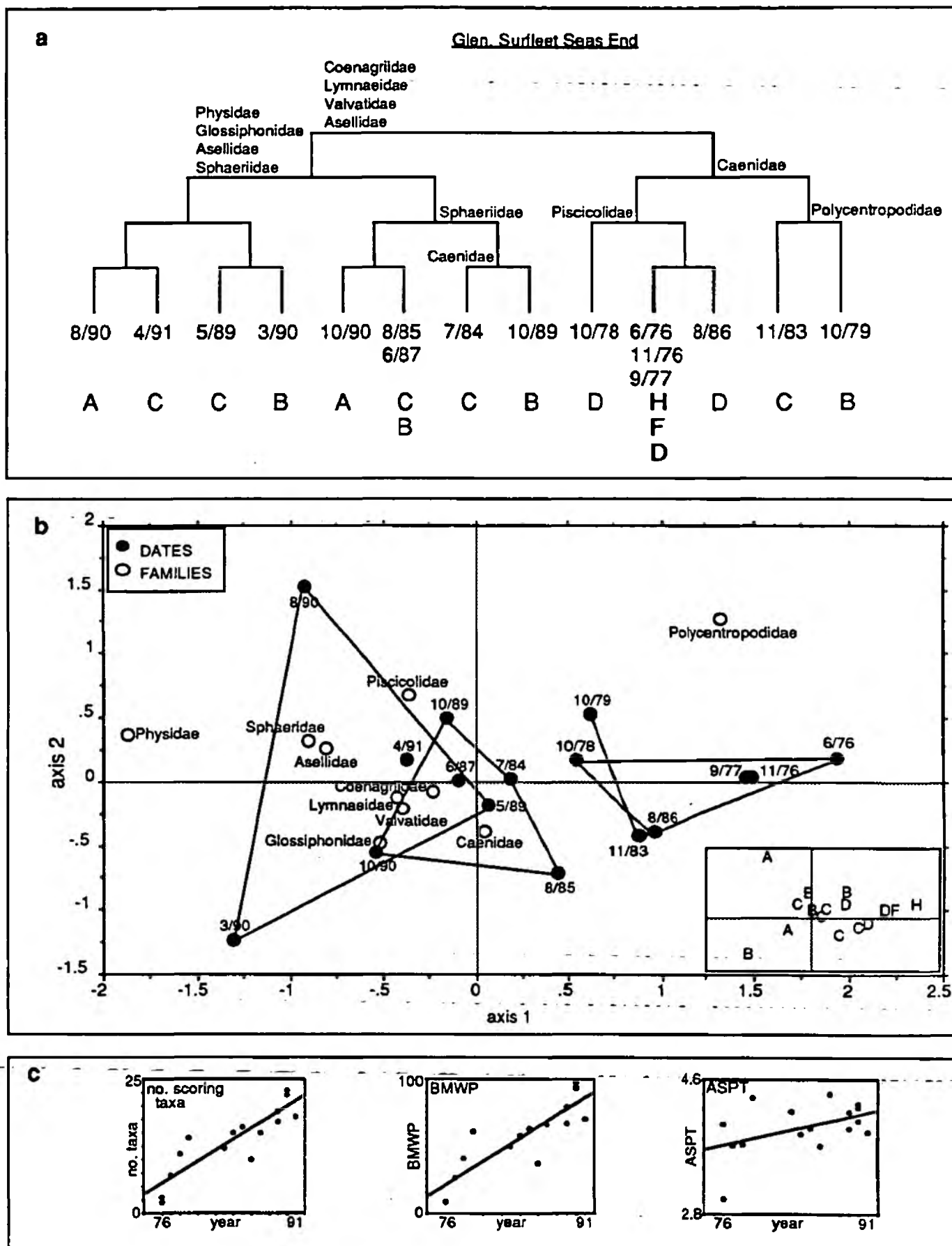


Figure B14. a) TWINSpan classification, b) ordination and changes in biotic score based on family data for the River Glen at Surfleet Seas End.

The ordination separates samples 8/90 and 3/90 far more clearly than the classification. Sample 3/90 possesses three taxa absent from all other: Dendrocoelidae, Helodidae and Corophidae; and lacks three taxa common in the majority of other higher-scoring samples: Piscicolidae, Hydracarina and Ostracoda. Sample 8/90 again has a few unique taxa: Psychomyidae, Pyralidae; a few relatively rare ones: Unionidae, Mysidae; and lacks the beetles Elmidae which occur in the other high scoring samples. Thus the ordination again appears more sensitive to unique taxa than the classification.

Within the TWINSpan groups 3 and 4 there is considerable variation in the numbers of taxa within samples rather than type. Samples 6/76 and 11/76 have severely restricted fauna, containing only Oligochaetes, Gammaridae, Chironomidae, Hydracarina and Cladocera. Saline intrusion is the probable cause, due to drought. Because they share all of the taxa present in the 1976 samples TWINSpan groups these and the 1977 sample together. However, this later sample has a considerably improved fauna with the addition of Dytiscidae, Baetidae, Planorbiidae, Corixidae and Copepoda. The other two samples in group 3 possess taxa including Haliplidae and Hydrobiidae, in addition to those in the other samples of the group.

Group 4 samples share most of the taxa present in group 3, but in addition have a number of others including two families of Trichoptera (Hydroptilidae and Polycentropodidae) - an Order completely absent from group 3. It is the presence of taxa such as the Cladocera, Gerridae and Copepoda in these group 4 samples in common with group 3 which places them on this side of the first classification division rather than with the other samples of similar score.

It seems probable that there has been an overall gradual improvement in habitat at this site, subsequent to (but not related to) a dramatic improvement following the drought of 1976 which resulted in a period of saline intrusion.

5.6.5 East Glen at Braceborough

Summary diagrams of the analyses are shown in Figure B15, and TWINSpan preferential taxa listed in Appendix 3.2.

The River East Glen at Braceborough differs from the previous examples in that biological quality peaked in 1986 and declined subsequently, whereas in the majority of sites the peak was in 1989-90. Sixteen biological records exist for Braceborough, over the period 1978-91.

The TWINSpan classification produces the following main groups:

- 1: 2/83, 6/86, 6/87, 8/87;
- 2: 4/89, 2/90, 3/91, 11/88, 11/89, 7/90;
- 3: 8/84, 7/85, 11/83, 10/90;
- 4: 10/78, 10/79.

These groups coincide with differences in both biological quality of samples and years of sampling. The first TWINSpan division separates the later samples (groups 1 and 2) from the earlier ones (groups 3 and 4), with two exceptions (see later). Within each of these two major groups the second split appears to divide samples of high (groups 1 and 3) from low (groups 2 and 4) biological quality.

The first TWINSpan division shows the "early" samples (groups 3 and 4) to have a number of taxa absent or less frequent in the "late" group, including relatively "sensitive" families such as the Leuctridae, Ephemerellidae, Hydroptilidae and Polycentropodidae; the molluscs Physidae and Valvatidae; a number of Coleopteran families: Haliplidae, Hydrophilidae and Gyrinidae; and the micro-crustaceans Copepoda, Cladocera and Ostracoda. The "late" group has relatively few preferential taxa: Limnephilidae, Hydropsychidae, Leptoceridae, Sericostomatidae and Psychodidae. Bearing in mind that this first split seems to be based on

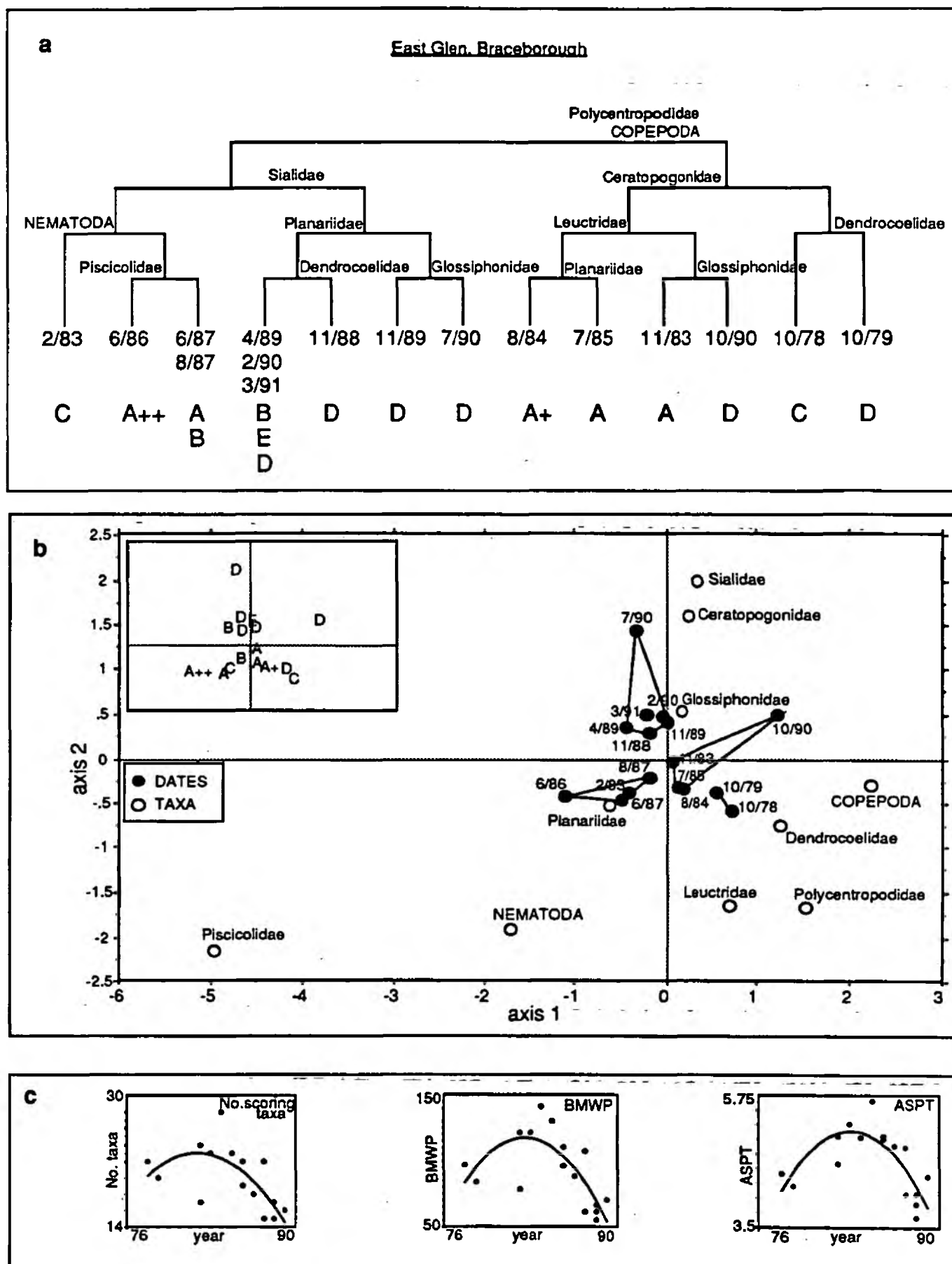


Figure B15. a) TWINSpan classification, b) ordination and c) changes in biotic scores based on family data for the East Glen at Braceborough.

temporal rather than water quality differences a possible explanation for the community changes seen might be that there has been a reduction in macrophyte cover or marginal habitat in more recent years.

Within the "late" group, the high-scoring group 1 separates from group 2 on the basis of a number of Ephemeroptera: Leptophlebitidae, Ephemerellidae, Ephemeridae and Caenidae; and Trichoptera: Hydropsychidae, Hydroptilidae, Limnephilidae and Glossosomatidae. Similarly, within the "early" group, high-scoring group 3 separates from group 4 because of the preference of several Ephemeroptera and Trichoptera: Leptophlebitidae, Ephemeridae, Caenidae, Limnephilidae and Goeridae; but also other insect orders: Odonata (Coenagriidae), Plecoptera (Leuctridae) and a large number of Coleoptera and Hemiptera: Gyrinidae, Hydrophilidae, Corixidae, Notonectidae and Gerridae. These reinforce the idea that good marginal habitat may be sustaining these rich communities.

Two samples do not appear to fit in with the early/late, high-scoring/low-scoring pattern outlined above. Sample 2/83 has a low score for group 1. It lacks many taxa present in the other members of this group, but has more in common with it than with group 4. Sample 10/90 also has a low score compared to the rest of its group, but most of those taxa it has it shares with other members that group. However, these are not all the most common taxa in the group, which may account for the ordination separating this sample more clearly than the classification. In other respects the ordination confirms the TWINSPLAN classification.

Disregarding the two "odd" samples that could be due to short-term events, the pattern suggests that there has been a permanent habitat change, possibly linked to improved marginal habitat, superimposed upon which are short term changes due to a range of stresses such as pollution or drought.

5.7 Seasonality

The life history patterns of many invertebrates, particularly univoltine insect species, create seasonal peaks and troughs in abundance of many taxa. It might therefore be expected that this element of seasonality in individual taxon abundance may lead to changes in numbers of taxa collected at a site and biotic scores depending on the time of year, particularly where a large proportion of the fauna share similar life histories. Insect dominated sites may show fewer taxa in early autumn, for instance, between emergence of one generation and hatching of the next.

Comparison of samples collected during different months using classification or ordination of the whole data set proved unsuitable for examining seasonal differences due to the overwhelming influence of site-specific variation. Applying similar methods to individual sites was also inappropriate as no single site had sufficient number of records.

Investigating the seasonal distribution of individual taxa was problematical due to the uneven distribution of samples between months. In an attempt to overcome this "weighted frequencies" were calculated for the commoner taxa by first separating the samples into the three major reach types ("upland", "intermediate" and "lowland" - see Section 5.4.4), then dividing the number of occurrences of each taxon within each month by the proportion of samples taken in each month within the reach type. The results for the three reach types were then totalled to give a single "weighted frequency" for each taxon for each month. These were then plotted against months to reveal any seasonal differences. It should be noted that the above procedure is not statistically reliable and was used only to look for general patterns. For several taxa seasonal patterns were apparent. Figure B16 shows the distribution of the Caenidae and Ephemerellidae and relates these to the known life-history curves for the common species in these families (Elliott et al 1988). The species chosen as representative of the families (*Ephemerella ignita* and *Caenis luctuosa* and *C. horaria*) have been found to be common in the River Glen (Milan, pers. comm.). For the Caenidae, a sharp decrease in frequency of collection occurs in July, corresponding to the emergence of the adults, followed by an increase during the period when

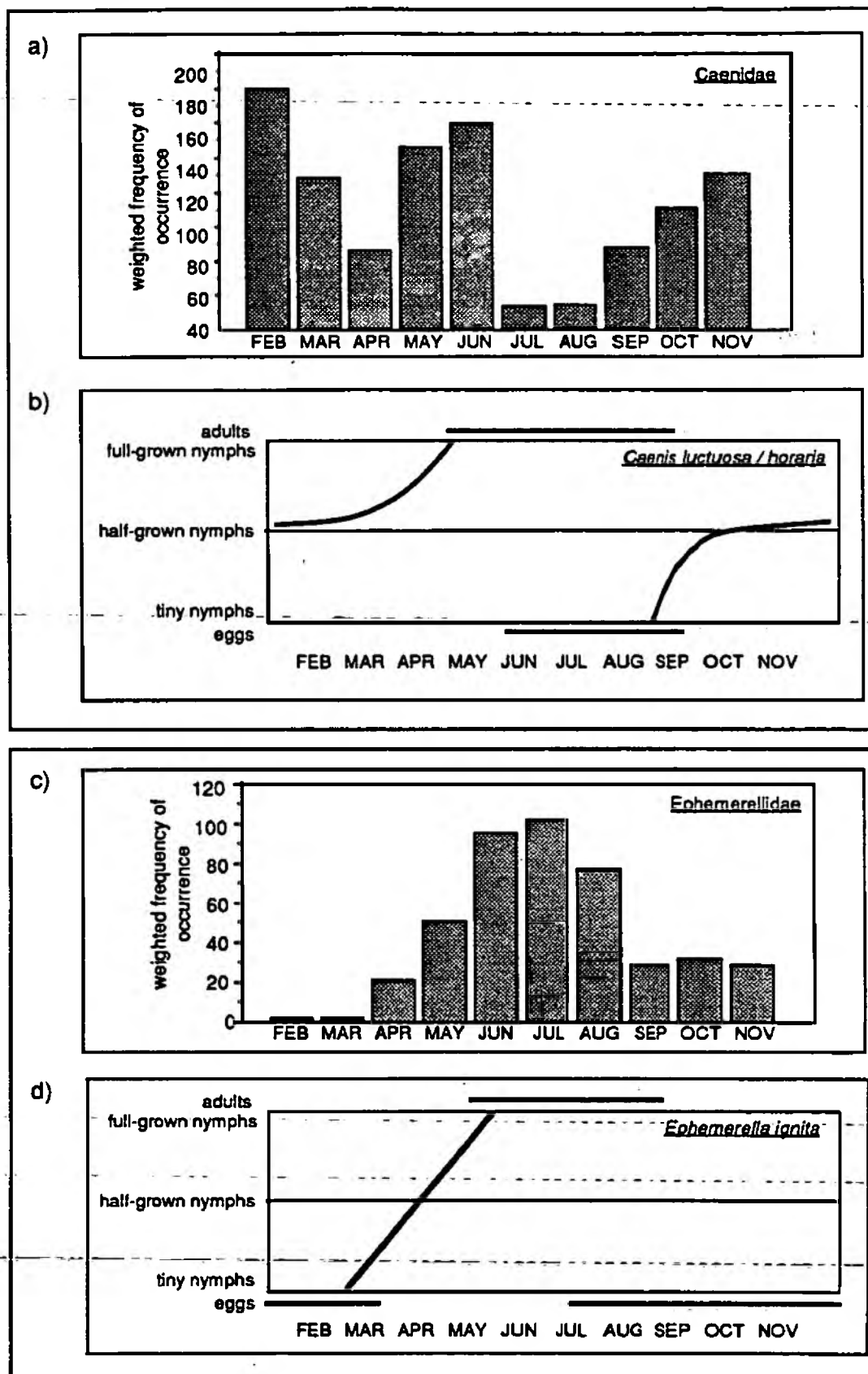


Figure 16. Frequency of occurrence of the families Ephemerellidae and Caenidae in each month. Frequencies are weighted to compensate for unequal sampling intensity within each month and reach type (units are arbitrary). Observed weighted frequencies: a) Caenidae and c) Ephemerellidae; temporal distribution of life-stages: b) *Caenis luctuosa* and *C. horaria* and d) *Ephemerella ignita* (adapted from Elliott *et al* 1988).

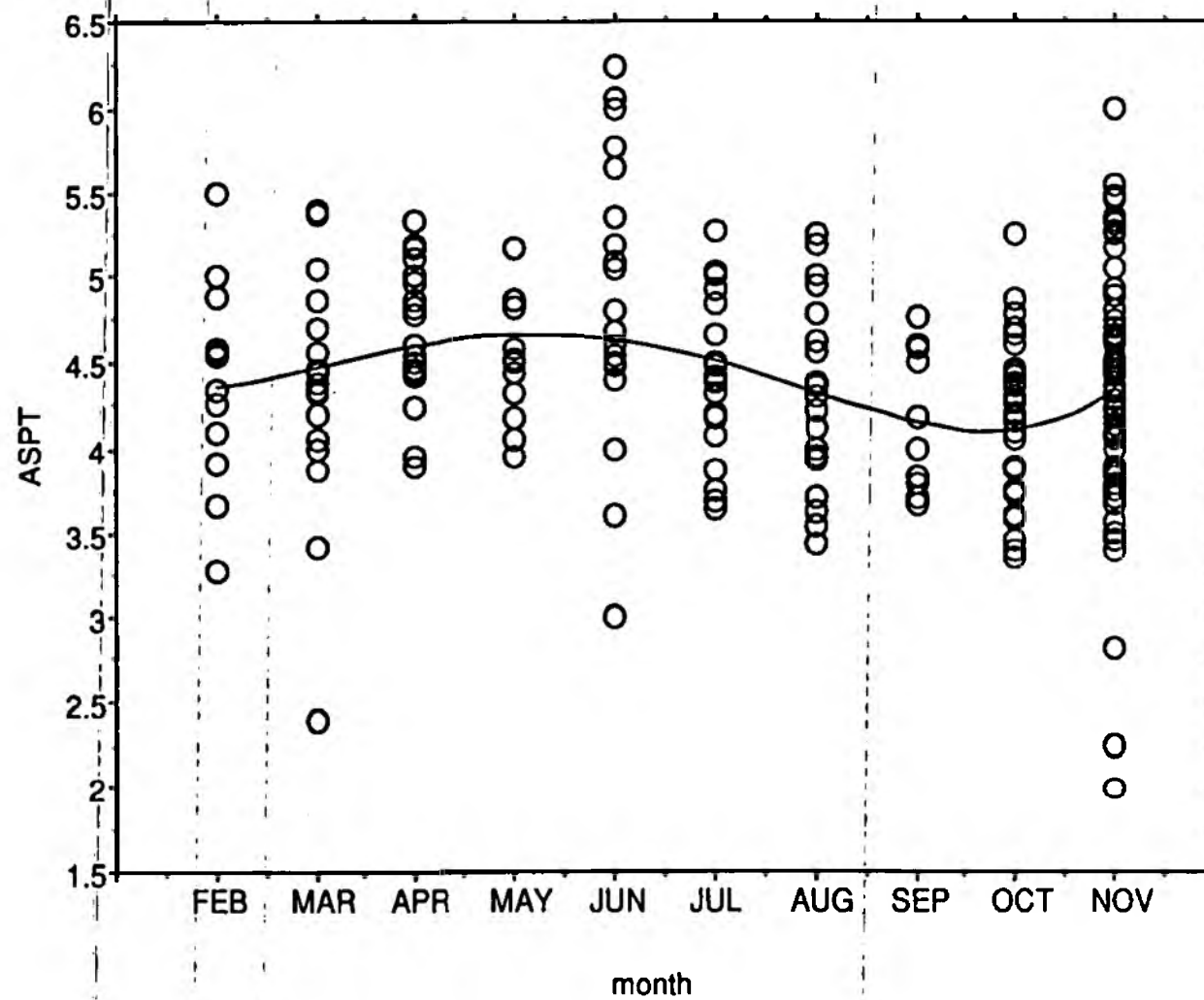


Figure B17. Regression of ASPT against month, all samples

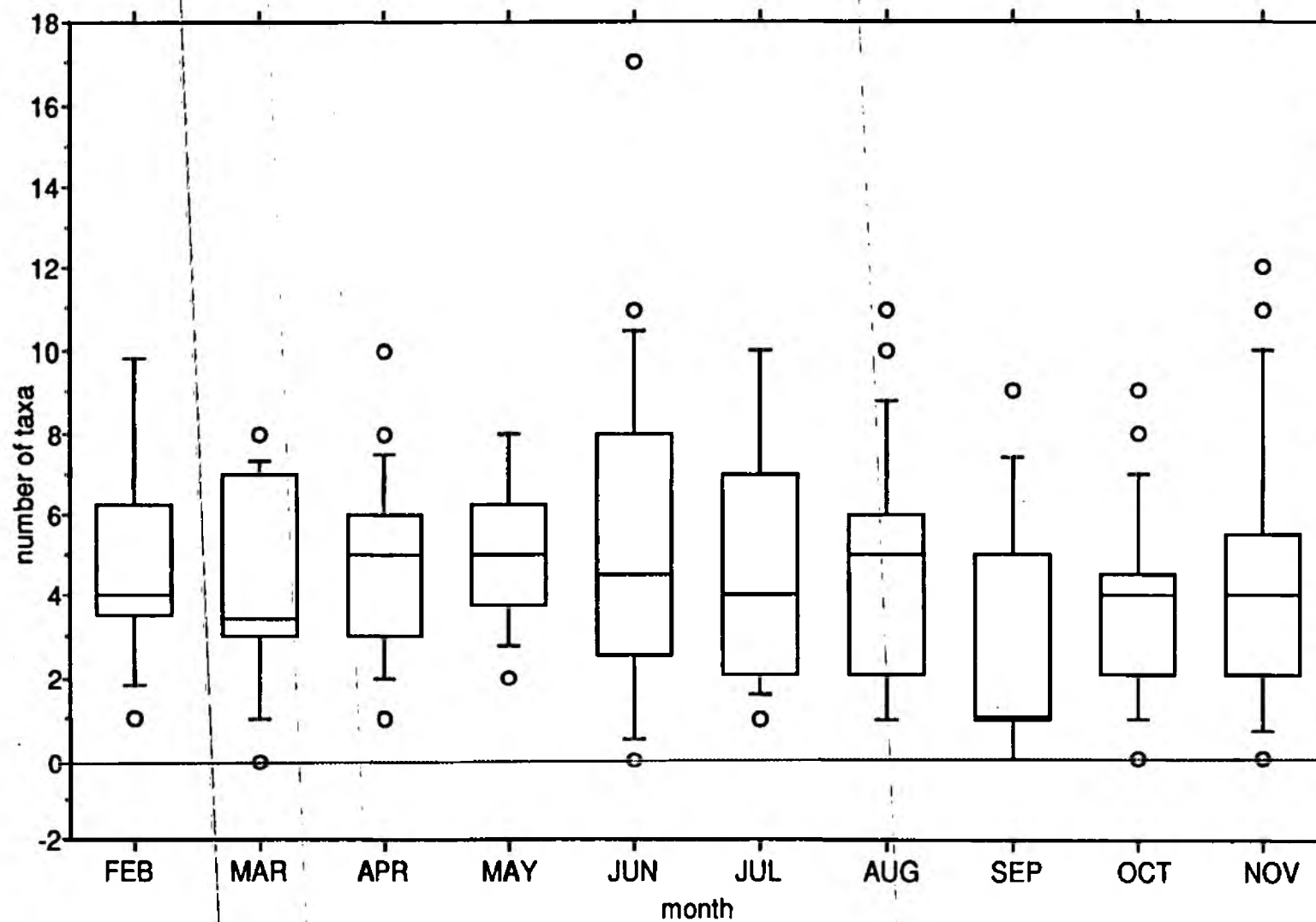


Figure B18. Numbers of Ephemeroptera, Plecoptera and Trichoptera taxa collected each month. Box plots indicate means, one standard deviation error and 95% confidence limits.

the next generation of nymphs would be hatching from eggs. In the Ephemerellidae the decrease in frequency of collection (which occurs during August-September) is later than that predicted from the known life-cycle - possibly due to a regional difference in the time of emergence.

No significant seasonal differences were found in the total number of taxa or BMWP. However, a significant relationship (NB Again the problem of the uneven distribution of samples between months makes this analysis unreliable) was found between ASPT and month of collection (Figure B17). Lowest scores were found in autumn, as might be predicted as high scoring taxa are predominantly insects with aquatic larvae and nymphs, the majority of which emerge in summer. Other explanations may be possible, linked to this season being the one of lowest flows.

Based on the assumption that the taxa contributing to the seasonality of ASPT scores would be predominantly the Ephemeroptera, Trichoptera and Plecoptera, numbers of these taxa alone were plotted against month (Figure B18). Contrary to expectations no significant seasonality was found in numbers of these taxa recorded, although the seasonal trends coincided with those of ASPT (Figure B17).

5.8 Performance of biotic indices

The use of biotic scores to evaluate the diversity and quality of invertebrate fauna at a site potentially masks differences in communities due to the grouping of taxa with quite varied behaviours, life histories and other ecological traits into score groups using the sole criterion of sensitivity to organic pollution. Classification and ordination of samples based on individual taxa can be used to look at variability within score groups, in order to identify how much between-sample variation is being ignored by consideration of biotic scores alone.

5.8.1 Variation within LQI classes

Figure B19 shows the correspondence analysis ordination of the family data from all sites with the area of the plot covered by samples from each LQI class shaded. This demonstrates that the area covered by samples within the A classes (A, A+ and A++) is greater, ie. the variety of communities is greater, than that of the lower classes. Thus grouping sites as "A" class disguises a greater range of faunas than E or F classes. Samples in the lower score classes have a greater proportion of taxa in common than those in the highest classes.

5.8.2 Classification and Ordination of samples within LQI classes

LQI A, A+ and A++ class samples

Classification of the samples which scored A, A+ or A++ on the LQI scale using TWINSpan revealed that the within class diversity could be accounted for largely by between site differences (Figures B20 and B21). A very distinct split was apparent between the lower main Glen and Bourne Eau sites on the left, and the Tham and West Glen on the right. The few East Glen samples achieving A class status split between the two main groups, with the more upstream sites on the left and downstream ones (EBR) on the right. This division mirrors that observed for the whole data set. Preferential taxa associated with this division in the A class samples (listed in Appendix 3.3) are the same as for the whole set, with the exceptions that the families Hydrophilidae, Gerridae and Physidae are also preferential for the left-hand group, and Sericostomatidae for the right-hand group in the A class samples, whereas the

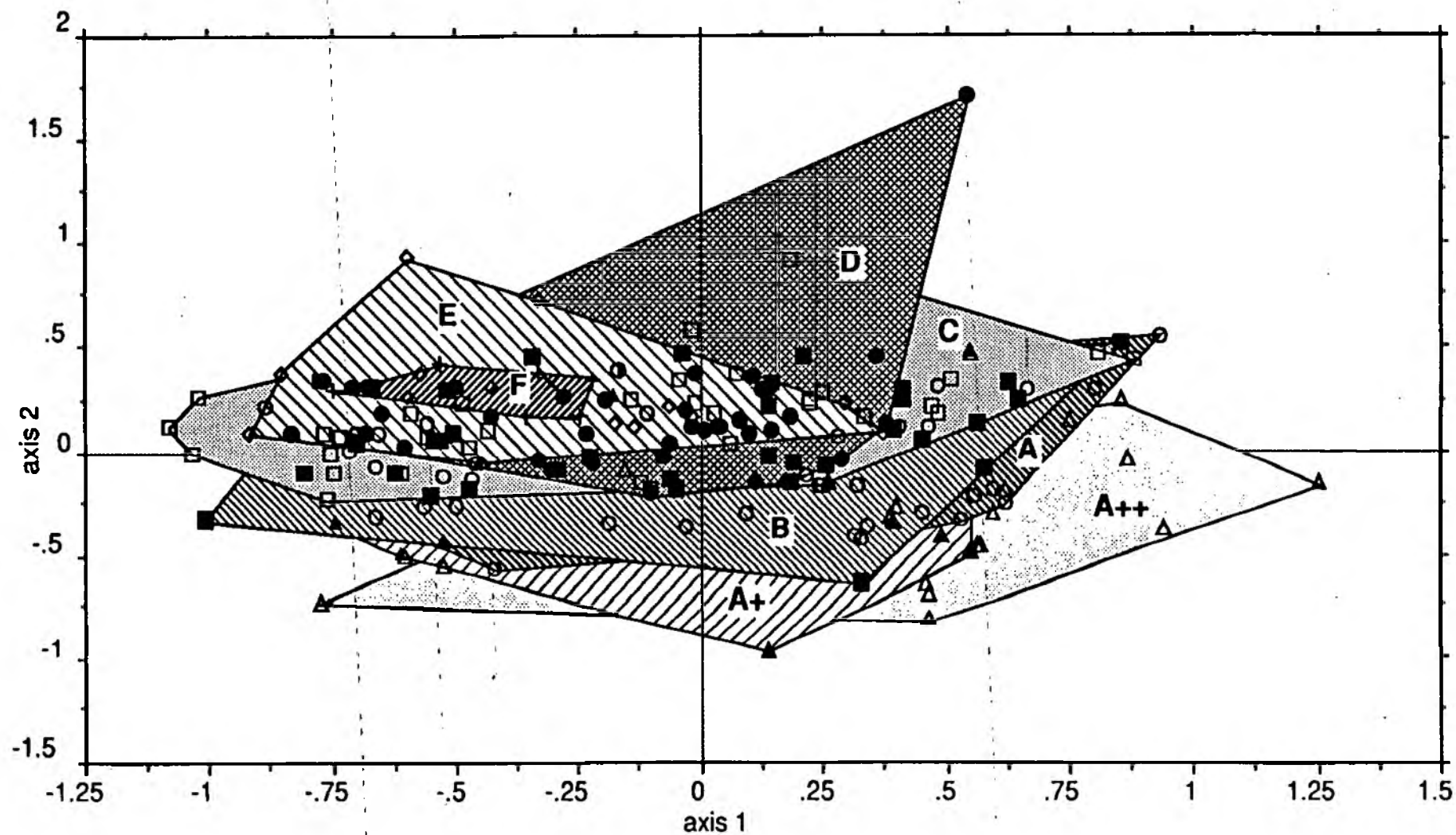


Figure B19. Ordination based on a correspondence analysis of family data for all samples; LQI classes are highlighted.

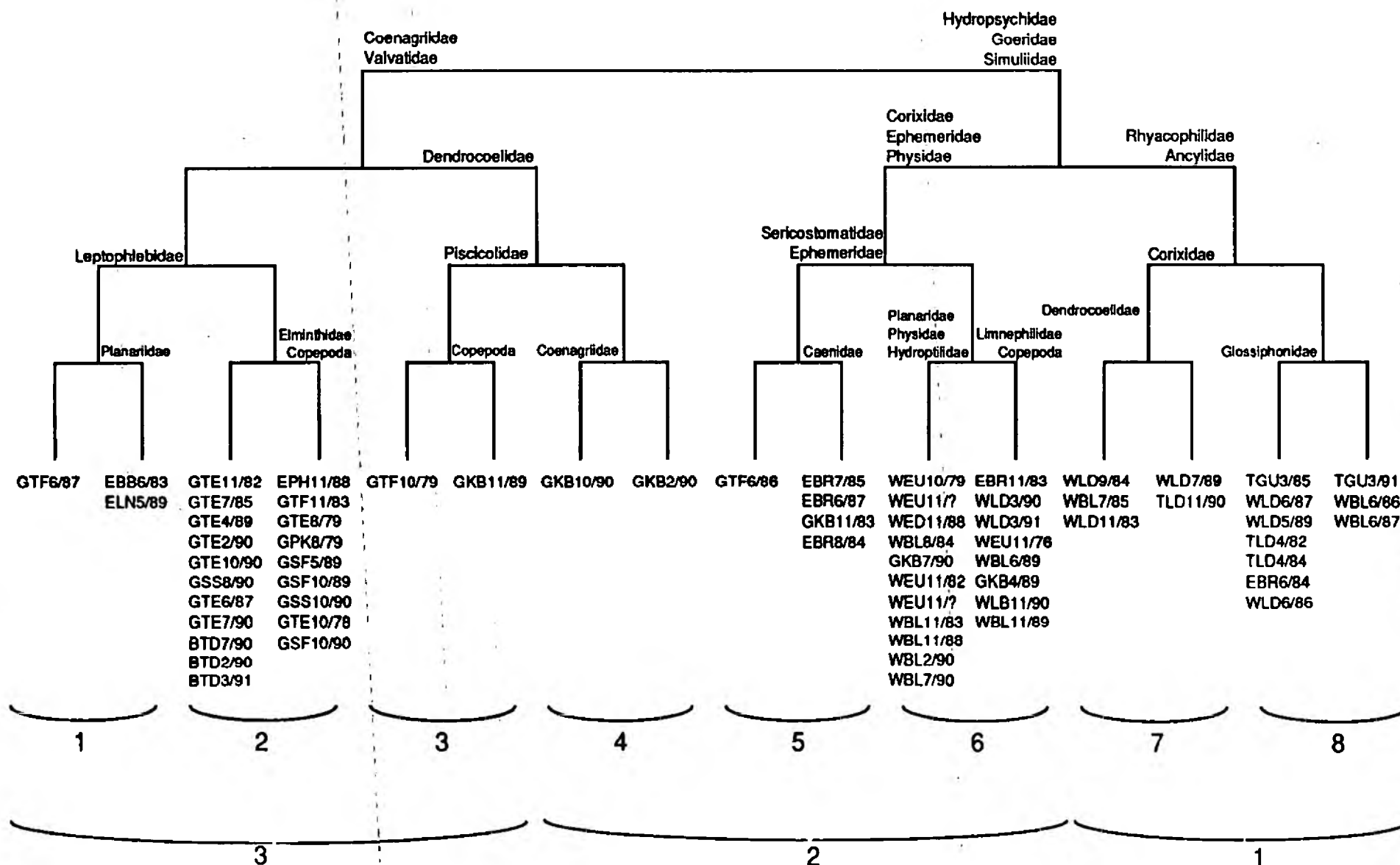


Figure B20. TWINSpan classification of LQI class A, A+ and A++ samples. Groups 1-8 are referred to in figures 20 and 21, and groups 1-3 in figure 26.

73	Valv	atid	-----22222222-22-222222222222-----2-----	1111
76	Neri	tida	-----22222222-22-222222222222-----2-----	1111
74	Unio	nida	-----2222-2-----2-----2-----2-----	1111
73	Empi	dida	-----2-----2-----2-----2-----	1111
66	Cull	cida	-----2-----2-----2-----2-----	1111
52	Hydr	ophi	-----2-----22-22-2-----2-----2-----	1111
47	Pyra	lida	-----22-----22-----22-----22-----	1111
39	Phry	gane	-----2-2-----22-----22-----22-----	1111
24	Coen	agri	2222-2222-22-222222222222-----2-----	1111
23	ANIS	OPTE	-----2-----2-----2-----2-----	1111
22	Mysi	dae	-----2-----2-----2-----2-----	1111
85	Succ	inei	-----2-----2-----2-----2-----	1110
58	Hydr	omet	-----2-2-----2-2-----2-2-----2-----	1110
17	CLAD	OCER	-----22-----222-----2222-----22-----	1110
80	Phys	idae	2-2-2-2-222222-222-222222-2-22-2-2222-----2-2-----	1101
70	Taba	nida	-----2-----2-----2-----2-----	1101
61	Noto	nect	2-2-2-22-22-2-2-2-2-22-22-2-2-2-2-2-2-2-----2-----	1101
60	Gerr	idae	-----2-----222-2-----2-2-2-22-----2-----2-----	1101
16	OSTR	ACOD	-----2-222-----2-2222-2-2222-----2-----222-----	1101
79	Lymn	aeid	22222-2222222222-22222222222-2222-2-22-2-22-----2-22222222-----	1100
19	Asel	lida	2-222222222222-2222222222222-2-2-2-2-22-222-222-2-22-2-2-----	1100
71	Musc	idae	-----22-----2-----2-----2-----2-----	1011
81	Plan	orbi	-----22222222-22222222222222222-22-222222-222222-2212222222-2-2-----	1010
78	Hydr	obi	2222222222222222222222222222-2222-22-22222-22-2-22222-2-22-2-2-----	1010
59	Cori	xida	2-222222222222222222-222222222222-22222222-2222222222-----	1010
49	Hali	plid	2-2222222222-2-22222222222-22-2-2222-22222-2-22-22222-2-2-----2-----	1010
42	Lept	ocer	2-2-2-2-222222222222222222-2-222222-22-222-222-2-2-22-22-22-----	1010
33	Sial	idae	-----22-2-----222-2-----222-----2-22-2222-2-22-2-2-----2-----	1010
65	Cera	topo	-----2-----22-2-22-2-22-2-22-2-2-22-2-2-2-----2-----2-----	1001
32	Caen	idae	2222222-2-22222-222-2222-222222222222-22222222-2-2-22-22222-----	1001
75	Spha	erid	-2222-222-22-2222222-----222-222222222222222222222222-2-2-22-2-----	1000
64	Chir	onom	2-222-222222-2222-22222222222222-----	1000
14	Glos	siph	-2222-222-2222222222-2222222-2-222222-22-22-2222222222222222-----	1000
8	Plan	arii	-222-222-2222-222222222222-222-2222222222-2-2-2-2-22-2222222-----	1000
53	Helo	dida	-----2-----2-----2-----2-----22-----22-----22-----2-----	0111
86	HYDR	ACAR	2-2-2-22-22-2222222-2222222222-222-22222-2-22-22-2-2222222222-2-----	0110
50	Dyti	scid	22222-2222222-2222222222222-22222222222-222222-2222-22-22222-2222-----	0110
28	Baet	idae	222-222222222222222222-----	0110
20	Gamm	arid	2-22-----	0110
18	COPE	PODA	2-2-----2-2222-----2-----2-----2-----2-2-2-22-22-----2-----	0110
13	Pisc	icol	2-22-2-2-2-2-2-2-222222-----2-2-2-2222-2-22-2-2-2-22222-22-----	0110
12	Olig	ocha	22-----	0110
2			22-----	0110
48	Elmi	nthi	22-----22-22-2222222-222-----	0101
15	Erpo	bdel	-22-----2-2222222-222-2222-2-2222222222222222222222222222222222-----	0101
51	Gyri	nida	-----2-----2-----22-----2-2-2-2-2-----22-----	0100
41	Mola	nnid	-----2-----2-----2-----2-----2-22-----	0100
38	Hydr	opti	-----22-----22-----2-2222-2-22-22-2-2-2-----22-----2-2-----2-----	0100
62	Tipu	lida	-----2-----22-----2-----222-2-22-2-2-2-2222-2-222222222222222222-----	0011
40	Limn	ephi	2-22-----22-----22222-2-22-2-2-2-222-2222222222222222222222-----	0011
36	Psyc	homy	-----2-----2-----2-----2-----22-----2-----2-----2-----	0011
34	Rhya	coph	-----2-----2-----2-----2-----22222-222-2222-----	0011
29	Lept	ophi	222-----2-22-2-22-22222-----222-----22-222222-2-----	0011

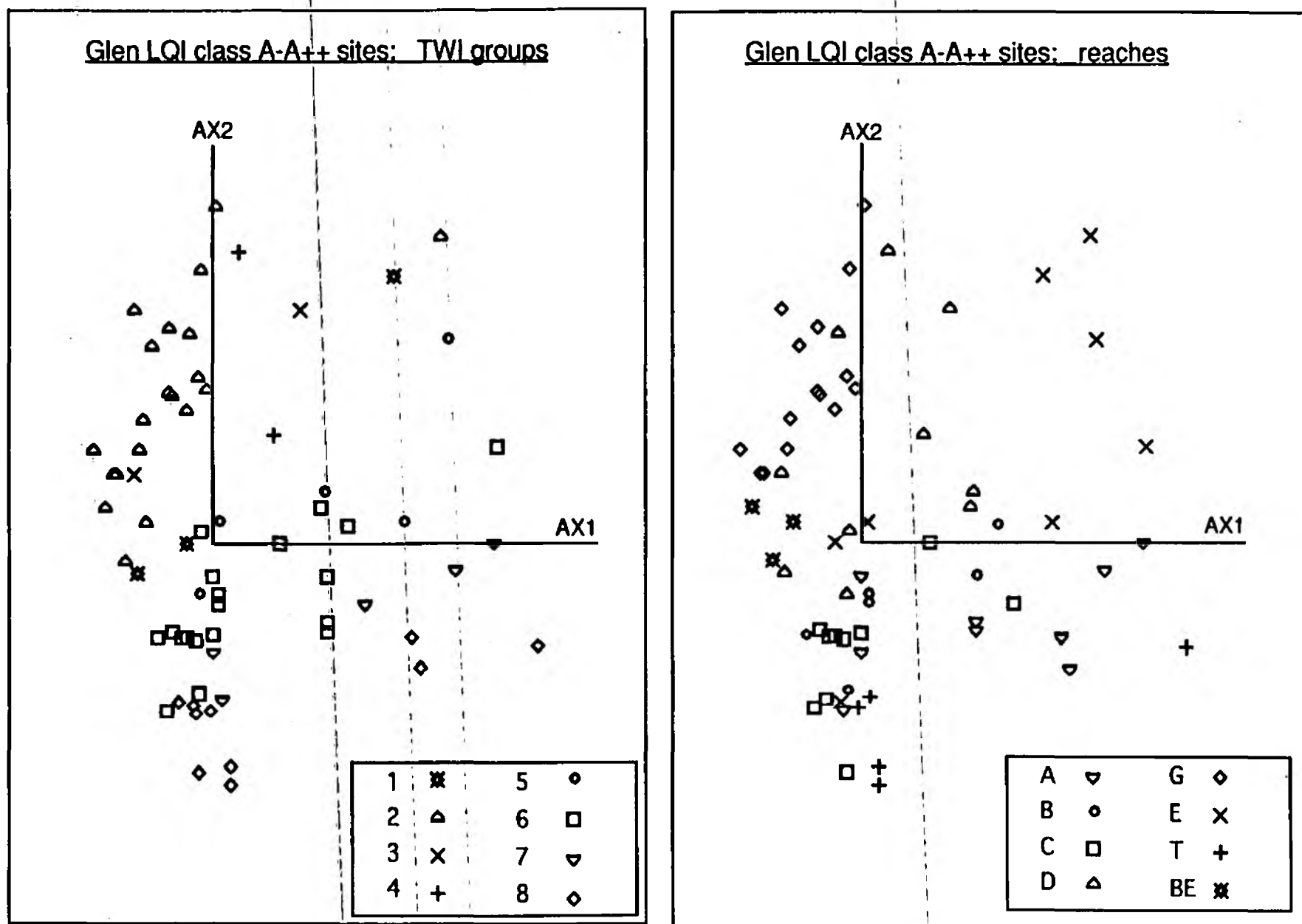


Figure B22. Ordination based on correspondence analysis of family data for samples in classes A-A++, with reaches and TWINSpan end groups (see Figure 20) highlighted.

Hydroptilidae are no longer more frequent in the right-hand group in this classification. Thus the site-based division is generally sharper in the A class group alone.

The next division within the lower Glen samples separates a small number which are relatively taxa-rich (one GTF and three GKB samples) from the rest, while in the West Glen and Tham samples split roughly between the lower West Glen and the few East and main Glen samples on the left, and the upper West Glen and Tham samples on the right. This again mirrors the classification of the whole data set.

Ordination of the A class data (see Figure B22, which shows the same ordination with reaches and TWINSPAN groups separately highlighted) confirmed the higher groupings produced by TWINSPAN, but showed the lower divisions to be somewhat arbitrary. Highlighting the samples by reach showed again that the TWINSPAN groups broadly coincided with the different reaches. The implications of the classification and ordination are that class A - A++ samples from different reaches contain communities characteristic of those reaches, with particular characteristic taxa, not just a greater diversity.

Classification and Ordination of LQI B and C class samples

Repeating the TWINSPAN classification (Figures B23 and B24) and correspondence analysis (Figure B23 - axis 3 appears to be as significant as axis 2 in this analysis, so plots using axes 1 and 2 and 1 and 3 are shown) on the samples scoring either B or C on the LQI scale produced as, if not more, positive a reach-separation as for the A class data. This was rather unexpected, as a greater diversity was predicted between the higher scoring samples. Again the lower, main river and Bourne Eau sites are separated from the rest on the basis of a rich mollusc fauna and certain other taxa characteristic of slow flowing or still-water sites with abundant macrophytes, eg. Cladocera, Coenagriidae and Notonectidae. (Preferential species for this classification are listed in Appendix 3.3.)

Figure B23. TWINSpan classification of LQI class B and C samples. Groups 1-8 are referred to in figures 23 and 24, and groups 1-3 in figure 26.

Figure B24. TWINSpan samples and taxa matrix for LQI class B and C samples

[illegible]

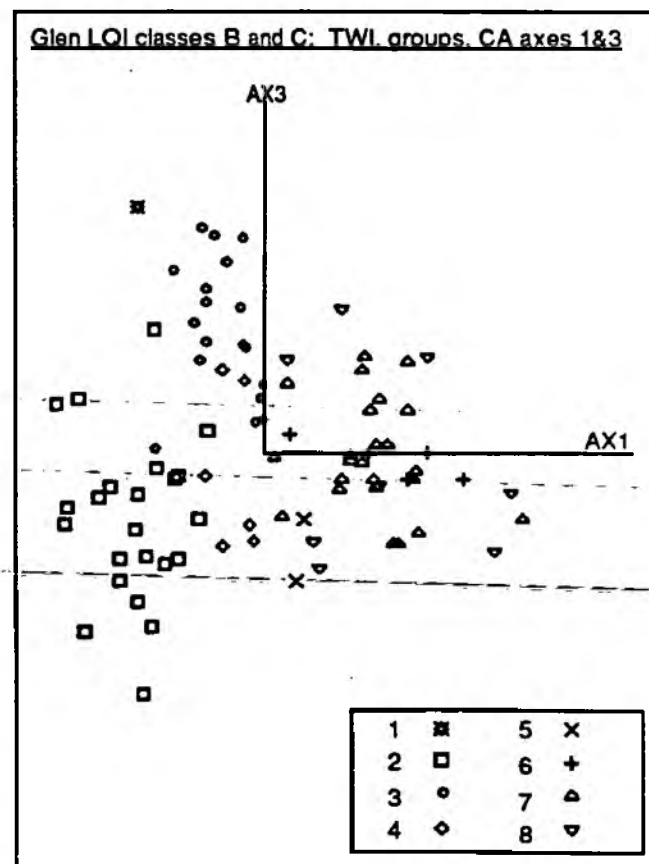
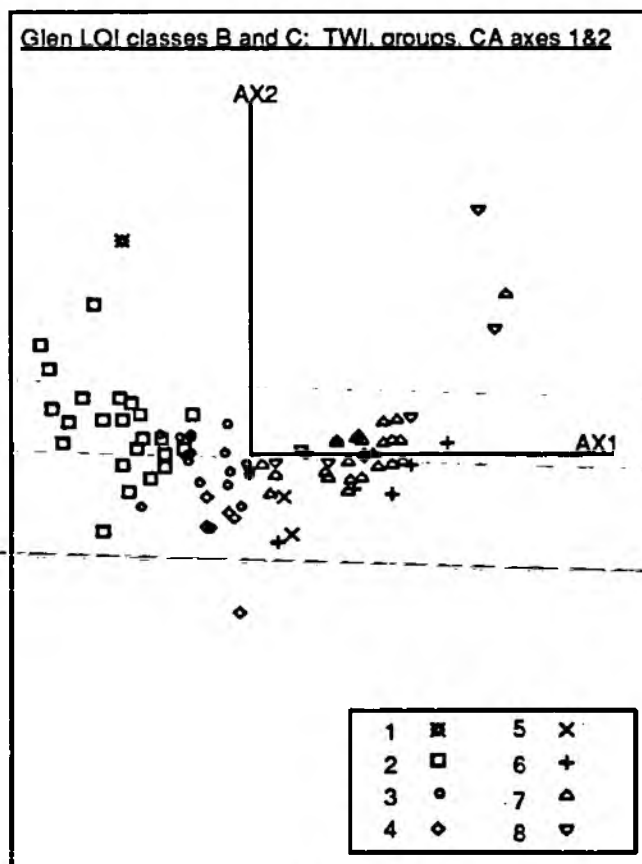
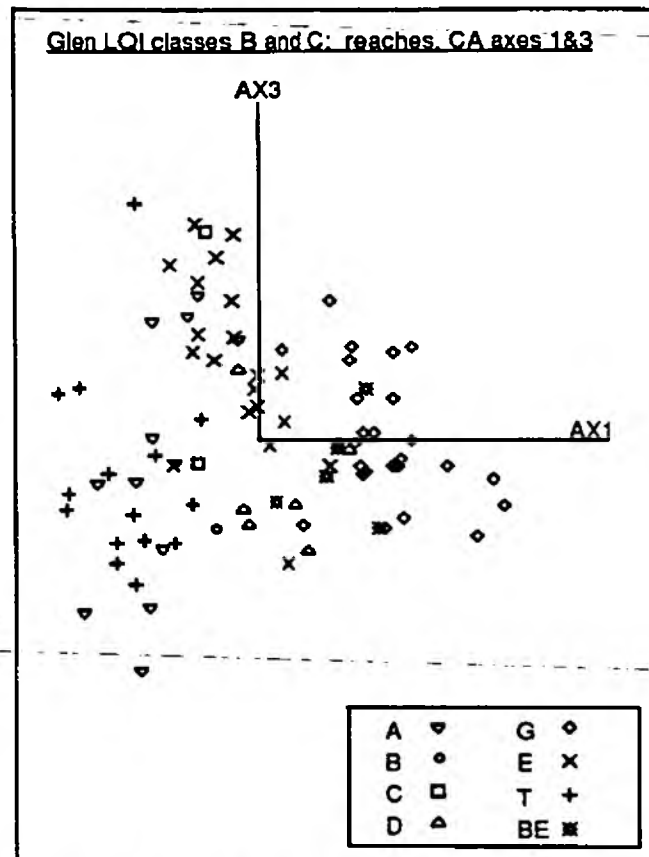
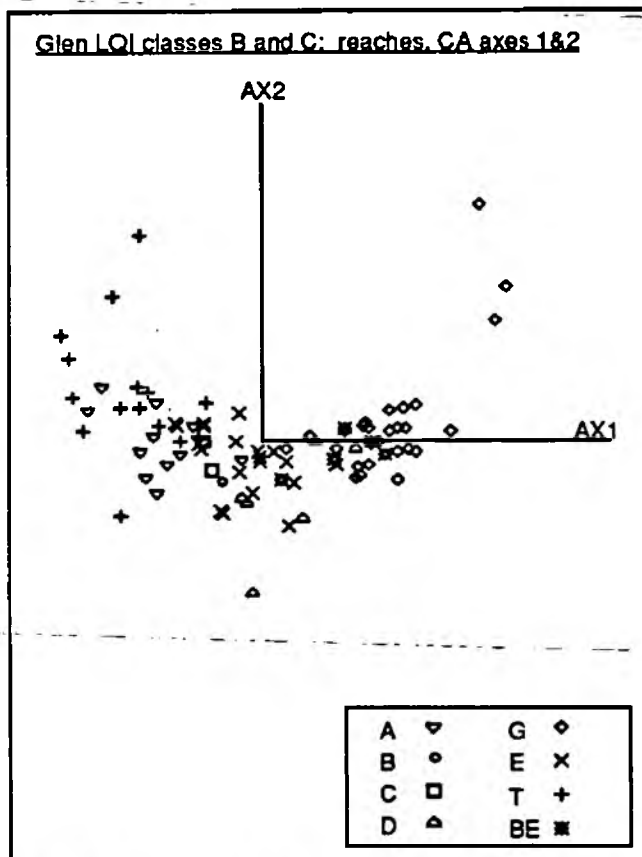


Figure B25. Ordination based on correspondence analysis of family data for samples in LQI classes B and C, axes 1&2 and 1&3, with reaches and TWINSpan end groups (see figure 21) highlighted.

5.8.3 Implications of within LQI class differences

On the evidence of the within-LQI class classifications above it appears that several different types of community can be defined within each class. It may be predicted that a class B site owes its characteristic fauna to a range of environmental conditions, and that were such a site to improve to class A status, due to, for example, an improvement in water quality, the other environmental constraints at that site would determine what type of class A community would develop.

By extracting from the data those class B and C samples from sites which at the next sampling occasion were class A or above, and comparing the TWINSPAN groups into which these fell with the TWINSPAN group of the subsequent class A sample, it was possible to assess whether class B and C communities of one TWINSPAN group always developed into the same type of class A community. The eight original TWINSPAN end groups were re-grouped into three, these being based on the most significant divisions (subjectively defined from the previous classifications - see Sub-sections 5.8.1 and 5.8.2) which broadly represent the "upper", "lower" and "intermediate" type faunas. Figure B26 shows the frequency with which each transition took place. It is clear that there is a strong link between the nature of the communities from the two bands of LQI class. The taxa associated with the transitions are listed in Table B4.

The variability within LQI classes and the reach-determined nature of this variation has clear implications where conservation measures are undertaken not only to improve biological quality but also to encourage a particular kind of invertebrate community or even specific (perhaps rare) taxa. Water quality improvement alone at a class B site may ensure improvement to class A status, but does not guarantee that, for instance, damselflies, which occur frequently in class A sites, will be present. Reference to the fauna of other class A sites within that reach, however, may enable an accurate prediction of the kind of community to be expected (see section 5.4.4).

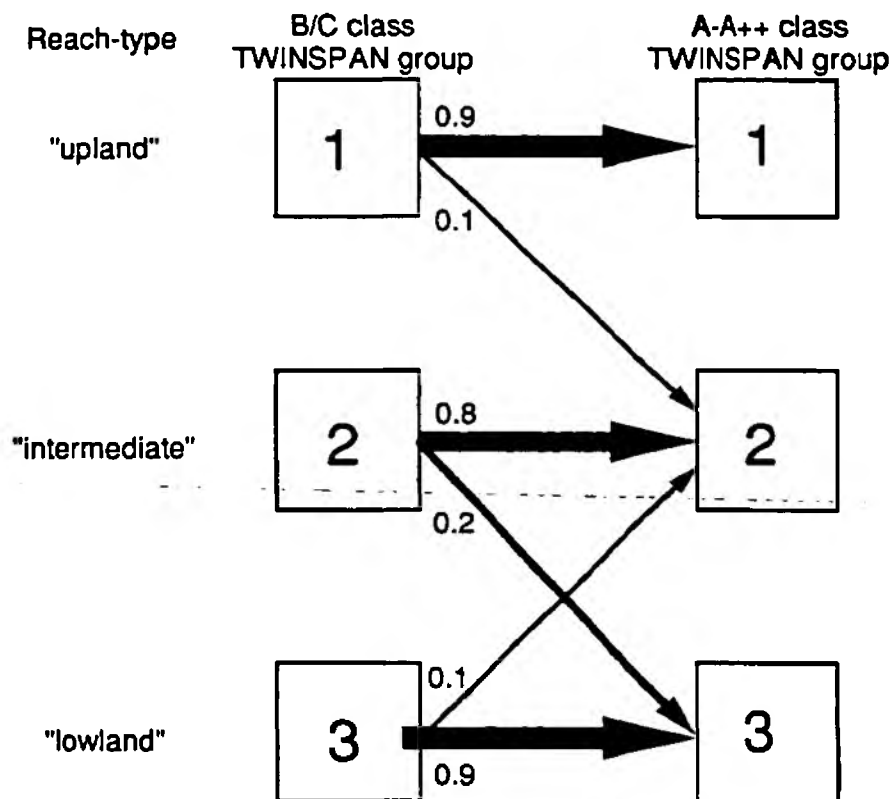


Figure B26. Frequency (numbers by arrows) of transition of LQI class B and C samples of TWINSpan groups 1-3 to LQI class A-A++ samples of TWINSpan groups 1-3.

Table B4. Taxa associated with changes from LQI class B/C to A-A++

Tham and middle West Glen (B/C gp.1 > A-A++ gp.3)		East Glen and "intermediate" sites (B/C gp.2 > A-A++ gp.2)		lower main Glen and Bourne Eau (B/C gp.3 > A-A++ gp.1)	
taxa gained	% samples	taxa gained	% samples	taxa gained	% samples
HYDRACARINA	50	Piscicolidae	80	Leptoceridae	60
Helodidae	50	Dendrocoelidae	50	Dytiscidae	40
Leptoceridae	50	COPEPODA	50	Gerridae	40
Leptophlebiidae	50	Leptophlebiidae	50	Hydrobiidae	30
Planariidae	30	Ephemeridae	50	Lymnaeidae	30
COPEPODA	30	Sericostomatidae	50		
Polycentropidae	30	Dytiscidae	50		
Limnephilidae	30	Corixidae	50		
taxa lost		taxa lost		taxa lost	
Sialidae	50	Asellidae	50		
Hydroptilidae	50	Tipulidae	50		
		Hydrobiidae	50		
		Planorbidae	50		
		Ancylidae	50		

6 CONCLUSIONS

6.1 Biological records

The data set of biological records for the River Glen system is probably a good representative of many for similar systems with a history of water quality, and quantity, problems and general conservation interest. It reflects not only changes in the invertebrate communities of the system, but the development of biological monitoring itself. The records begin in the mid-1970's when biological indices were first widely used, and when the severe drought of 1976 caused particular concerns about damage to, and recovery of, biological communities. The most widely used Biological index at the time was the Trent Biotic Index, and the level of identification used in invertebrate monitoring was that needed to derive TBI scores and little more. The later development of the BMWP system, the LQI system and later still the recognition of the importance of seasonally consistent, routine sampling and species level differences demanded improvements in the level of identification and frequency of sampling: all reflected in the detail of the data set.

These long-term changes in the nature of the invertebrate records inevitably effect their interpretation, particularly in two ways: First, where the level of identification varies, for comparisons to be valid only the highest taxonomic groups can be used, and much detail in "better" records is wasted. Secondly, as the motivation for monitoring has changed from response to particular incidents to routine quality assessment, the average quality observed and the sites targeted will have changed.

If long term assessments of biological records are considered of value, then the above problems should be considered especially carefully when either planning a new or altering an old monitoring program.

6.2 Site and reach level differences in Invertebrate communities

The invertebrate fauna of the Glen system is typical of lowland-type rivers, being rich in a range of taxa including many insect orders, molluscs and crustaceans. This invertebrate diversity reflects the water chemistry and diversity of substrate, flow characteristics and macrophytes. Classification and ordination of the family level data revealed distinct reach and site level differences in communities, but also great similarities. Representatives of the Trichoptera, Coleoptera, Mollusca and Crustacea were found in the majority of samples, with changes at the family level distinguishing sites as opposed to whole orders or classes as can be found in comparisons between rivers. Reach-scale differences in community were clear, with "upland", "lowland" reaches and an "intermediate" zone around the confluence of the West and East Glens being clearly distinguished. These reach-types defined on the basis of the invertebrate fauna corresponded closely to known physical differences at this scale, for examples sedimentological differences. By the nature of the faunal differences it appeared that water quality was only one of a number of physical-attributes varying between sites and reaches.

The analytical tools used to investigate site and reach differences - TWINSpan and correspondence analysis - proved successful in combination. TWINSpan is very useful for identifying assemblages of taxa characterising groups of samples and reaches, but is less helpful in assessing the magnitude of between-group differences, especially where these are principally differences relating to presence or absence of taxa. Ordination is essential for comparing the separation of groups and identifying aberrant samples.

From the classification of the three reach-types, lists of taxa associated with each could be produced, which may be regarded as the total communities which any site within those reaches could potentially possess.

6.3 Biological scores

Biological scores including LQI proved efficient at identifying within site differences in fauna: In the site case-studies samples of similar score were consistently grouped together; only occasionally did temporal differences override score differences, and on these occasions long term changes in habitat were inferred. At the site and reach level, however, differences were greater between sites and reaches than between score groups, indicating that factors other than water quality were structuring the communities. Within LQI classes, for instance, site and reach differences were as clear as within the whole data set. Also, these between site and reach differences were as great within the lower LQI classes (B and C) as between the highest (A - A++). This is rather surprising as it was assumed that there would be greater uniformity in the lower-quality sites, with a limited range of common taxa. What may in fact be happening is that where the fauna is reduced it is a more specialised community which remains, being more highly adapted to the particular site-type, ie. it implies that specialism and sensitivity to water quality are not necessarily linked.

The diversity within sites of high and moderate quality (as defined by LQI) emphasises the importance of between-site differences in factors other than water quality: Hydrology, hydraulics, substrate, macrophytes and other biotic and abiotic characteristics, in determining the composition of invertebrate communities, and has clear implications for river management.

6.4 Long-term changes

Examination of changes in biological scores over the 1976-91 period for the whole data set indicated that biological quality rose from 1976 to the late 1980's, with a slight fall in 1991. More detailed analysis of individual sites proved that these did not all show the same pattern of changes. Whilst most sites followed the pattern described for the whole data set, some showed a continuous increase in scores, some no change and a few a progressive decrease. In addition, changes in BMWP were a result of either changes in the quality (ASPT) or number of taxa, or both. The implication of this is that changes in invertebrate diversity are occurring as a result of factors in addition to changes in water quality. Some long term changes are likely to be due to "structural" changes in the habitat, perhaps related to weed-cutting and dredging regimes, whereas other, shorter-term changes reflect more variable influences, the most significant of these is likely to be hydrology. A further examination of the biological data in conjunction with the hydrological records could therefore be very enlightening.

The five "case-studies" revealed interesting relationships between temporal changes in invertebrate community and biological score. In the case of Banthorpe Lodge on the West Glen the fauna of the 1991 sample closely resembled that of the late 1970's samples, with which it also shared a low score, suggesting that the community responded to drought in a similar manner on both occasions. However, at Surfleet Seas End on the main Glen the relatively high scoring 1979 sample showed more faunal similarities with the other 1970's samples, despite their low scores, than with the high scoring 1980's samples; while the low scoring 1991 sample resembled the other later samples rather than low scoring 1970's ones. This could be due to saline intrusion during the 1970's period. The Glen at Kates Bridge showed a different pattern again, with the low-scoring 1991 sample resembling faunistically neither the 1980's samples it was close to temporally, nor the 1970's samples it was close to in score. It therefore appears that the fauna of different sites behave differently under stress: Either changing to a new, adapted and quite different community (if, for example, the management regime changes or there is a permanent change in discharge due to abstraction or augmentation), or to a reduced form of their normal community, with only the most sensitive taxa eliminated. Such differences may depend on the nature of the stress, its duration (whether it results in a permanent change in habitat), and also on the nature and source of recolonisation after such a stress event.

6.5 Seasonal differences

Evidence for seasonal changes in frequency of collection was found for a number of taxa, predictably families of univoltine insects. Although no significant seasonal changes in BMWP or number of taxa were observed in the whole data set, small but significant changes were observed in ASPT: a distinct trough was observed in September / October. This could be a result of the fact that most high-scoring taxa are insect nymphs and larvae, which do have seasonal peaks and troughs of abundance related to growth, emergence, and egg- or later-stage diapause. However, an attempt to relate numbers of the three orders of insects containing most of these taxa (Plecoptera, Ephemeroptera and Trichoptera) failed to show any significant pattern similar to that of ASPT. The trough in ASPT was later than that expected if the predominance of these taxa were indeed the cause. An alternative possibility is that annual lowest flows, which frequently coincide with this September / October period, are influencing the taxa present and / or collected.

7 RECOMMENDATIONS

Analysis of the existing invertebrate records for the River Glen system has revealed that the invertebrate community is responsive to:

- reach-scale habitat conditions, and
- physical and biotic, as well as chemical, variations at sites within the reach-scale constraints.

These findings suggest that there is great potential for expanding the use of biological monitoring beyond the setting and monitoring of water quality objectives (for which it was designed and appears to perform satisfactorily) to assess the potential "best" invertebrate community on a reach-specific basis and set targets if required based on this potential, and by improved and coordinated measurement of a range of habitat factors to monitor the effects of these on the invertebrate community and its ability to achieve the predicted potential. The following strategy (illustrated in Figure B27) is designed to provide the necessary information for such an expanded use of biological monitoring:

7.1 Preliminary reach assessment

An initial biological survey of the river system incorporating the maximum number of sites possible and at least three seasonal collections should be carried out in order to identify reach types. Existing invertebrate records could be included in this initial assessment. The choice of sampling sites should aim to include as many habitat types as possible, particularly those subjectively identified as "habitat rich". From this preliminary survey reaches would be defined on the basis of the fauna using a combination of analytical techniques including classification and ordination of sites as described in section 4. For each reach a list of taxa which could potentially be supported in that reach would be compiled.

7.2 Selection of routine monitoring sites

Within each reach a number of routine sites should be selected, including those which fulfil the following criteria (within the normal limits eg. accessibility):

- sites of the highest habitat quality
- sites susceptible to ecological damage (pollution, low flows, any other locally important factors)
- sites reflecting the range of management regimes in the reach

Routine monitoring of these sites should provide a continually-updated picture of the maximum and minimum biological quality of the reach, reflecting natural and anthropogenic influences on the invertebrate community.

7.3 Biological monitoring - frequency

Sampling carried out in spring, summer and autumn (as at present) appears to provide adequate seasonal coverage. Emergence of some insect taxa can take place within a few days, making it preferable for all sites within a reach to be sampled over the same few days each year, thus allowing accurate between-site and between-year comparisons.

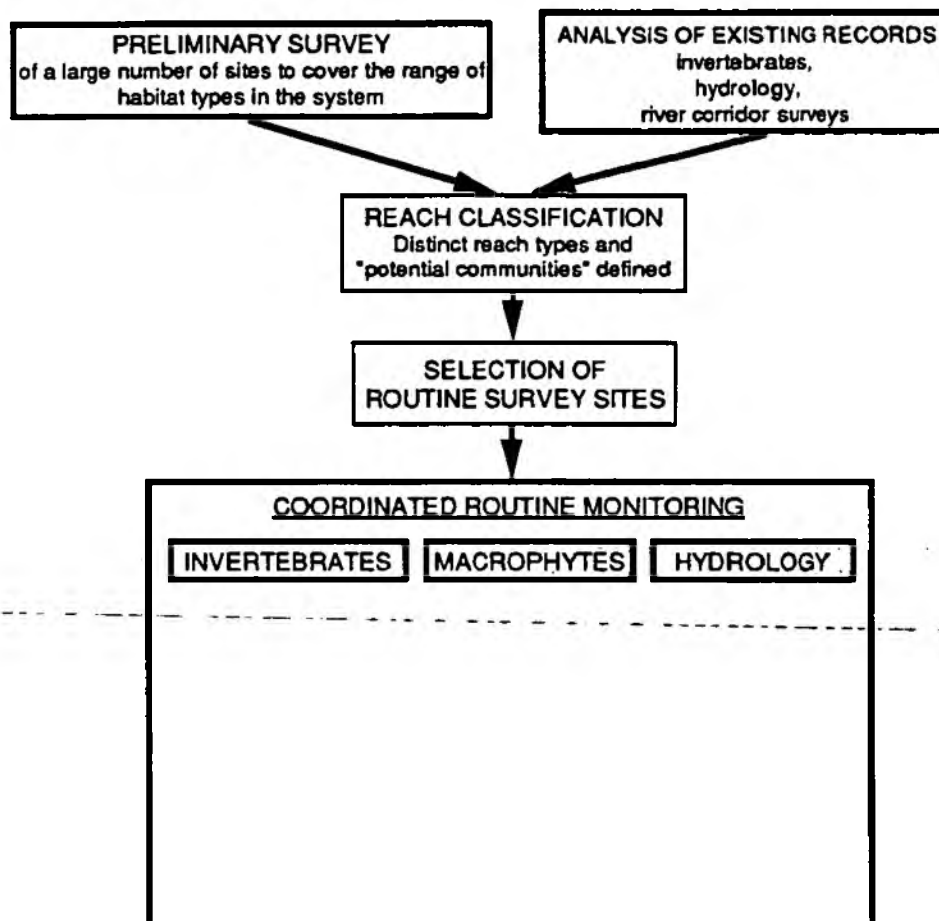


Figure B27 Strategy for reach assessment, coordinated biological and hydrological recording and integration of additional information to allow the monitoring of short- and long-term change.

7.4 Biological monitoring - field procedure

7.4.1 Invertebrate sampling

The upstream and downstream limits of each site should be clearly defined, and sampling using the kick and sweep method carried out between these limits on each occasion, with the effect that a reduction in wet width will result in a smaller area being sampled. Sampling effort should be divided between micro-habitats in proportion to their area within the site.

7.4.2 Invertebrate identification

If identification to species level is considered desirable and practicable for some taxa then this should be consistently used for these taxa. Varying identification between family and species level necessitates that the species level information be wasted when between-sample comparisons are made. For some taxa, species level identification is easy during one season but impossible during others, in which case separate recording forms might be considered for the spring, summer and autumn samples which would allow between-year if not between season comparisons for these groups.

7.5 Associated routine monitoring

In order to relate between-site and between sample changes to the physical and other biotic site characteristics, the assessment of a suite of variables should be performed at the same time as the invertebrate sampling:

- macrophyte cover and type - a record of the percentage cover of the site and dominant species would be more informative than a list of all species presence / absence alone.
- substrate composition - an index of dominant substrate type and amount of surface siltation.
- hydraulic characteristics - coordinating routine current metering surveys with the invertebrate monitoring would provide vital information on site depth and flow velocity distributions, which could be related to discharge and / or water level information from continuous monitoring stations.

8 REFERENCES

Hill, MO (1979) TWINSpan - A FORTRAN Program for Arranging Multivariate Data in an Ordered Two-Way Table by Classification of the Individuals and Attributes. *Ecology and Systematics*, Cornell University. Ithica, New York 14850.

Extence, CA (1989) Biological survey of the East and West Glens. Internal report for Anglian Water.

Extence, CA and Ferguson, JD (1989) Aquatic invertebrate surveys as a water quality management tool in the Anglian Water Region. *Regulated Rivers: Research and Management* 4 139-146.

Ter Braak, CJF (1988) CANOCO - a FORTRAN program for canonical community ordination by [partial] [detrended] [canonical] correspondence analysis, principal components analysis and redundancy analysis (version 2.1). Agricultural Mathematics group, Ministry of Agriculture and Fisheries. Wageningen.

APPENDICES

APPENDIX B1. TAXA LISTS FOR ALL SAMPLES

Appendix 1.

TAXON	EG8.	EPH1.	EMT.	EMU.	EMD.	EMG.	EBR.	ET1.1	ET1.4.
BRYOZOA									
COELENTERATA									
POLYZOA									
PORIFERA									
<i>Dugesia polychroa/lugubris</i>									
<i>Dugesia ligna</i>									
<i>Polysia</i> sp.									
<i>Dendrocoelum lacteum</i>									
NEMATODA									
NEMATOMORPHA									
OLIGOCHAETA									
Piscolidae									
<i>Glossiphonia complanata</i>									
<i>Glossiphonia heteroclita</i>									
<i>Helobdella stagnalis</i>									
<i>Hemicleps marginata</i>									
<i>Theromyzon tessulatum</i>									
Erpobdellidae									
OSTRACODA									
CLADOCERA									
COPEPODA									
<i>Asellus aquaticus</i>									
<i>Asellus mendianus</i>									
<i>Crangonyx</i> sp.									
<i>Gammarus</i> sp.									
<i>Gammarus zaddachi</i>									
Corophidae									
<i>Neomysis integer</i>									
ANISOPTERA									
Coenagnidae									
<i>Calopteryx splendens</i>									
Nemouridae									
Leuctidae									
<i>Baetis</i> sp.									
<i>Centropilum</i> sp.									
<i>Cloeon</i> sp.									
Leptophlebiidae									
<i>Ephemerella ignita</i>									
Ephemerae									
Caenidae									
<i>Stalis lutana</i>									
Rhyacophidae									
Polycentropidae									
Psychomyidae									
Hydropsychidae									
Hydropsulidae									
Phryganeidae									
Limnephilidae									
Molannidae									
Leptocendae									
Goeridae									
<i>Lepidostoma hirtum</i>									
Lepidostomatidae									
Sarcostomatidae									
<i>Agapetus</i> sp.									
<i>Glossosoma</i> sp.									
<i>Nymphula nympheta</i>									
<i>Elmis aenea</i>									
<i>Limnius volkman</i>									
<i>Oulimnius</i> sp.									
<i>Riolus</i> sp.									
Halipidae									
Dytiscidae									
Gyrinidae									
Hydrophilidae									
Helodidae									
Dryopidae									
Chrysomelidae									
Mesoveliidae									
Velidae									
Hydrometidae									
Conidae									
Gemidae									
Notonectidae									
Dicranota l.									
Tipulidae l.									
Pedicia l.									
Simuliidae									
Chironomidae									
Ceratopogonidae									
<i>Culex</i> sp.									
Dixidae									
<i>Pericoma</i> sp. l.									
<i>Psychoda</i> sp. l.									
Psychopteridae									
Stratiomyidae									
Tabanidae									
Muscidae									
<i>Ernstia</i> sp.									
Empididae									
Unionidae									
Sphaeriidae									
Neridae									
Valvatidae									
<i>Potamopyrgus jenkinsi</i>									
<i>Hydrobia</i> sp.									
<i>Bythinia leachi</i>									
<i>Bythinia tentaculata</i>									
<i>Lymnaea auriculata</i>									
<i>Lymnaea glabra</i>									
<i>Lymnaea palustris</i>									
<i>Lymnaea peregra</i>									
<i>Lymnaea stagnalis</i>									
<i>Lymnaea truncatula</i>									
Physidae									
Planorbidae									
Vivipandae									
<i>Ancyus fluviatilis</i>									
<i>Acroloxus lacustris</i>									
Zonitidae									
Succineidae									
HYDRACARINA									

Appendix 1.

[illegible]

Appendix 1.

[illegible]

Appendix 1.

TAXON	W.ED.	W.B.L.	11/88	11/76	10/78	05/75	08/79	02/83	11/83	08/84	07/85	05/86	06/87	11/88	05/89	11/89	02/90	07/90	05/91
BRYOZOA																			
COELENTERATA																			
POLYZOA																			
PORIFERA																			
<i>Dugesia polychroa/lugubris</i>																			
<i>Dugesia ligna</i>																			
<i>Polycelis</i> sp.																			
<i>Dendrocoelum lacteum</i>																			
NEMATODA																			
NEMATOMORPHA																			
OLIGOCHAETA																			
Pisicoidae																			
<i>Glossiphonia complanata</i>																			
<i>Glossiphonia heterochita</i>																			
<i>Helobdella stagnalis</i>																			
<i>Hemiclepsis marginata</i>																			
<i>Theromyzon tessulatum</i>																			
Erpobdellidae																			
OSTRACODA																			
CLADOCERA																			
COPEPODA																			
<i>Asellus aquaticus</i>																			
<i>Asellus mendianus</i>																			
<i>Crangonyx</i> sp.																			
<i>Gammarus</i> sp.																			
<i>Gammarus zaddachi</i>																			
Corophidae																			
<i>Neomysis integer</i>																			
ANISOPTERA																			
Coenagrionidae																			
<i>Calopteryx splendens</i>																			
Nemouridae																			
Leuctridae																			
<i>Baetis</i> sp.																			
<i>Centropilum</i> sp.																			
<i>Cloeon</i> sp.																			
Leptophlebiidae																			
<i>Ephemerella ignita</i>																			
Ephemerae																			
Caenidae																			
<i>Salix lutana</i>																			
Rhyacophilidae																			
Polycentropidae																			
Psychomyidae																			
Hydropsychidae																			
Hydroptilidae																			
Phryganeidae																			
Limnephilidae																			
Molannidae																			
Leptoceridae																			
Goeridae																			
<i>Lepidostoma hirtum</i>																			
Lepidostomatidae																			
Sencostomatidae																			
<i>Agapetus</i> sp.																			
<i>Glossosoma</i> sp.																			
<i>Nymphula nymphalea</i>																			
<i>Elms aenea</i>																			
<i>Limnias volkman</i>																			
<i>Oulimnius</i> sp.																			
<i>Riotus</i> sp.																			
Halipidae																			
Oytidae																			
Gynidae																			
Hydrophilidae																			
Helodidae																			
Dryopidae																			
Chrysomelidae																			
Mesoveliidae																			
Velidae																			
Hydrometridae																			
Conidae																			
Geridae																			
Notonectidae																			
Dicranota l.																			
Tipulidae l.																			
Pedicia l.																			
Simuliidae																			
Chironomidae																			
Ceratopogonidae																			
<i>Culex</i> sp.																			
Dixidae																			
<i>Pencoma</i> sp. l.																			
<i>Psychoda</i> sp. l.																			
Ptychoporidae																			
Strabomyidae																			
Tabanidae																			
Muscidae																			
<i>Eristalis</i> sp.																			
Empididae																			
Unionidae																			
Sphaeridae																			
Neridae																			
Valvatidae																			
<i>Potamopyrgus jenkinsi</i>																			
<i>Hydrobia</i> sp.																			
<i>Bythinea leachi</i>																			
<i>Bythinea tentaculata</i>																			
<i>Lymnaea auncularia</i>																			
<i>Lymnaea glabra</i>																			
<i>Lymnaea palustris</i>																			
<i>Lymnaea peregra</i>																			
<i>Lymnaea stagnalis</i>																			
<i>Lymnaea truncatula</i>																			
Physidae																			
Planorbidae																			
Vivipandae																			
<i>Ancylus fluviatilis</i>																			
<i>Acroloxus lacustris</i>																			
Zonitidae																			
Succineidae																			
HYDRACARINA																			

Appendix 1.

[illegible]

[illegible]

Appendix 1.

TAXON	B.SU.	B.SD.	B.RL.	B.TU.	B.YD.
BRYOZOA					
COELENTERATA					
POLYZOA					
PORIFERA					
<i>Dugesia polychroa/lugubris</i>					
<i>Dugesia bignna</i>					
<i>Polycelis</i> sp.					
<i>Dendrocoelum lacteum</i>					
NEMATODA					
NEMATOMORPHA					
OLIGOCHAETA					
<i>Piscolidae</i>					
<i>Glossiphonia complanata</i>					
<i>Glossiphonia heteroclita</i>					
<i>Helobdella stagnalis</i>					
<i>Hemicleipsis marginata</i>					
<i>Theromyzon tessellatum</i>					
<i>Eupobdellidae</i>					
OSTRACODA					
CLADOCERA					
COPEPODA					
<i>Asellus aquaticus</i>					
<i>Asellus mendianus</i>					
<i>Crangonyx</i> sp.					
<i>Gammarus</i> sp.					
<i>Gammarus zaddachi</i>					
<i>Corophidae</i>					
<i>Neomysis integer</i>					
ANISOPTERA					
<i>Coenagnidae</i>					
<i>Calopteryx splendens</i>					
<i>Nemouridae</i>					
<i>Leuctidae</i>					
<i>Baetis</i> sp.					
<i>Centropilum</i> sp.					
<i>Cloeon</i> sp.					
<i>Lepidophlebiae</i>					
<i>Ephemerella ignita</i>					
<i>Ephemerae</i>					
<i>Caenidae</i>					
<i>Sialis lutana</i>					
<i>Rhyacophilidae</i>					
<i>Polycentropidae</i>					
<i>Psychomyidae</i>					
<i>Hydropsychidae</i>					
<i>Hydroptilidae</i>					
<i>Phryganeidae</i>					
<i>Limnephilidae</i>					
<i>Molannidae</i>					
<i>Leptoceridae</i>					
<i>Goenidae</i>					
<i>Lepidostoma hirtum</i>					
<i>Lepidostomatidae</i>					
<i>Sencostomatidae</i>					
<i>Agapetus</i> sp.					
<i>Glossosoma</i> sp.					
<i>Nymphula nymphalae</i>					
<i>Elmis aenea</i>					
<i>Limnias volkmanni</i>					
<i>Oulimnius</i> sp.					
<i>Riolus</i> sp.					
<i>Halipidae</i>					
<i>Dytiscidae</i>					
<i>Gyrinidae</i>					
<i>Hydrophilidae</i>					
<i>Helodidae</i>					
<i>Dryopidae</i>					
<i>Chrysomelidae</i>					
<i>Mesoveliidae</i>					
<i>Velidae</i>					
<i>Hydrometridae</i>					
<i>Corixidae</i>					
<i>Geridae</i>					
<i>Notonectidae</i>					
<i>Dicranola</i> l.					
<i>Tipulidae</i> l.					
<i>Pedicia</i> l.					
<i>Simuliidae</i>					
<i>Chironomidae</i>					
<i>Ceratopogonidae</i>					
<i>Culex</i> sp.					
<i>Oxidae</i>					
<i>Pericoma</i> sp. l.					
<i>Psychoda</i> sp. l.					
<i>Psychoptendae</i>					
<i>Stratiomyidae</i>					
<i>Tabanidae</i>					
<i>Muscidae</i>					
<i>Ensiatis</i> sp.					
<i>Empididae</i>					
<i>Unionidae</i>					
<i>Sphaeridae</i>					
<i>Neritidae</i>					
<i>Valvatidae</i>					
<i>Potamopyrgus jenkinsi</i>					
<i>Hydrobia</i> sp.					
<i>Bythinia leachi</i>					
<i>Bythinia tentaculata</i>					
<i>Lymnaea auricularia</i>					
<i>Lymnaea glabra</i>					
<i>Lymnaea palustris</i>					
<i>Lymnaea peregra</i>					
<i>Lymnaea stagnalis</i>					
<i>Lymnaea truncatula</i>					
<i>Physidae</i>					
<i>Planorbidae</i>					
<i>Vivipandae</i>					
<i>Ancylus fluviatilis</i>					
<i>Acrolorus lacustris</i>					
<i>Zonitidae</i>					
<i>Succineidae</i>					
HYDRACARINA					

Appendix 1.

TAXON	T.CH.	T.CU.	T.CD.	T.GU.	T.GD.	T.LU.	T.LD.
BRYOZOA							
COELENTERATA							
POLYZOA							
PORIFERA							
<i>Dugesia polychroa/lugubris</i>							
<i>Dugesia ligna</i>							
<i>Polycelis</i> sp.							
<i>Dendrocoelum lacteum</i>							
NEMATODA							
NEMATOMORPHA							
OLIGOCHAETA							
Pisicoidae							
<i>Glossiphonia complanata</i>							
<i>Glossiphonia heteroclita</i>							
<i>Helobdella stagnalis</i>							
<i>Hemicleipsis marginata</i>							
<i>Theromyzon tessulatum</i>							
Erpobdellidae							
OSTRACODA							
CLADOCERA							
COPEPODA							
<i>Asellus aquaticus</i>							
<i>Asellus meridianus</i>							
<i>Crangonyx</i> sp.							
<i>Gammarus</i> sp.							
<i>Gammarus zaddachi</i>							
Corophidae							
<i>Neomysis integer</i>							
ANISOPTERA							
Coenagrionidae							
<i>Calopteryx splendens</i>							
Nemouridae							
Leuctidae							
<i>Baetis</i> sp.							
<i>Centropetrum</i> sp.							
<i>Cloeon</i> sp.							
Leptophlebiidae							
<i>Ephemerella ignita</i>							
Ephemeridae							
Caenidae							
<i>Stalioa lutana</i>							
Rhyacophilidae							
Polycentropidae							
Psychomyidae							
Hydropsychidae							
Hydroptilidae							
Phryganidae							
Limnephilidae							
Molannidae							
Leptocendae							
Goeridae							
<i>Lepidostoma hirtum</i>							
Lepidostomatidae							
Sencostomatidae							
<i>Agapetus</i> sp.							
<i>Glossosoma</i> sp.							
<i>Nymphula nymphaloides</i>							
<i>Elmis aenea</i>							
<i>Limnius volkmani</i>							
<i>Oulimnius</i> sp.							
<i>Riolus</i> sp.							
Halplidae							
Dytiscidae							
Gyrinidae							
Hydrophilidae							
Helodidae							
Dryopidae							
Chrysomelidae							
Mesoveliidae							
Velidae							
Hydrometidae							
Corixidae							
Geridae							
Noloneclidae							
Dicranota l.							
Tipulidae l.							
Pedicia l.							
Simuliidae							
Chironomidae							
Ceratopogonidae							
<i>Culex</i> sp.							
Dixidae							
<i>Pericoma</i> sp. l.							
<i>Psychoda</i> sp. l.							
Ptychoptera							
Stratiomyidae							
Tabanidae							
Muscidae							
<i>Eristalis</i> sp.							
Empididae							
Unonidae							
Sphaeridae							
Neriidae							
Valvatidae							
<i>Polamopyrgus jenkinsi</i>							
<i>Hydrobia</i> sp.							
<i>Bythinia leachi</i>							
<i>Bythinia tentaculata</i>							
<i>Lymnea auricularia</i>							
<i>Lymnea glabra</i>							
<i>Lymnea palustris</i>							
<i>Lymnea peregra</i>							
<i>Lymnea stagnalis</i>							
<i>Lymnea truncatula</i>							
Physidae							
Planorbidae							
Vivipandae							
<i>Ancylus fluviatilis</i>							
<i>Acrolux lacustris</i>							
Zonitidae							
Succineidae							
HYDRACARINA							

APPENDIX B2. BIOTIC SCORES FOR ALL SAMPLES

Appendix 2. RIVER GLEN BIOLOGICAL SCORES

SAMPLE	BMWP	NO.SCRG.TAXA	ASPT	HABITAT TYPE	LOI
E.LN.11/88	48	12	4.0	RR	D
E.LN.5/89	86	19	4.5	RR	A+
E.LN.8/89	31	9	3.4	RR	E
E.LN.3/90	41	12	3.4	RR	E
E.LN.8/90	47	13	3.6	RR	D
E.LN.11/90	35	10	3.5	RR	E
E.LN.3/91	12	5	2.4	RR	G
E.HW.11/88	75	18	4.2	R	D
E.BB.9/77	60	15	4.0	RR	C
E.BB.10/78	54	15	3.8	RR	C
E.BB.10/79	54	14	3.9	RR	C
E.BB.11/82	52	13	4.0	RR	C
E.BB.6/83	63	14	4.5	RR	A
E.BB.11/83	75	17	4.4	RR	B
E.BB.11/88	64	15	4.3	RR	B
E.EL.11/88	82	16	3.9	RR	C
E.EH.11/76	23	6	3.8	R	F
E.EH.10/78	86	21	4.1	R	D
E.EH.10/79	86	20	4.3	R	D
E.EH.11/83	82	20	4.6	R	B
E.EH.8/84	97	21	4.6	R	B
E.EH.7/85	69	18	4.3	R	D
E.EH.6/87	69	15	4.6	R	C
E.EH.11/88	82	19	4.3	R	D
E.EH.4/89	80	18	4.4	R	D
E.EH.11/89	76	18	4.8	R	C
E.EH.2/90	81	19	4.3	R	D
E.EH.7/90	48	13	3.7	R	E
E.EH.10/90	77	19	4.1	R	D
E.EH.3/91	78	18	4.3	R	D
E.GB.11/82	86	19	4.5	R	D
E.GB.11/89	71	16	4.4	R	D
E.GB.7/90	48	13	3.7	R	E
E.PH.9/77	53	14	3.8	RR	C
E.PH.11/82	58	14	4.1	RR	B
E.PH.11/88	82	19	4.3	RR	A
E.MT.2/88	47	12	3.9	R	E
E.MU.2/88	65	15	4.3	R	D
E.MU.7/88	53	12	4.4	R	E
E.MD.2/88	75	15	5.0	R	C
E.MD.7/88	40	11	3.6	R	E
E.MG.11/78	45	10	4.5	R	E
E.MG.9/77	44	12	3.7	R	E
E.MG.11/82	66	16	4.1	R	D
E.MG.11/83	81	18	4.5	R	D
E.MG.2/88	64	14	4.6	R	C
E.MG.11/88	75	17	4.4	R	D
E.BR.10/78	97	22	4.4	R	C
E.BR.10/79	84	20	4.2	R	D
E.BR.2/83	78	17	4.6	R	C
E.BR.11/83	121	24	5.0	R	A
E.BR.8/84	121	23	5.3	R	A+
E.BR.7/85	141	28	5.0	R	A
E.BR.6/86	130	23	5.7	R	A++
E.BR.6/87	96	19	5.1	R	A
E.BR.8/87	110	22	5.0	R	B
E.BR.11/88	88	19	4.9	R	C
E.BR.4/89	107	22	4.9	R	B
E.BR.11/89	61	15	4.1	R	D
E.BR.2/90	55	15	3.7	R	E
E.BR.7/90	61	15	4.1	R	D
E.BR.10/90	66	17	3.9	R	D
E.BR.3/91	70	16	4.4	R	D
ET1.1.6/83	18	3	6.0	R	B
ET1.4.6/83	24	5	4.8	R	E
ET1.4.11/88	55	13	4.2	R	E

Appendix 2. RIVER GLEN BIOLOGICAL SCORES

SAMPLE	BMWP	NO.SCRQ.TAXA	ASPT	HABITAT TYPE	LOI
W.BP.11/88	62	14	4.4	R	D
W.BF.10/78	84	20	4.2	R	D
W.BF.5/79	71	17	4.2	R	D
W.BF.8/79	88	17	4.0	R	D
W.BF.11/83	84	18	4.7	R	C
W.BF.11/88	74	18	4.8	R	C
W.BC.11/88	78	17	4.5	R	D
W.BC.5/89	101	22	4.8	R	B
W.BC.7/89	88	21	4.2	R	D
W.BC.3/90	82	18	4.6	R	C
W.BC.8/90	74	18	4.1	R	D
W.BC.11/90	32	9	3.6	R	E
W.BC.3/91	56	13	4.3	R	E
W.CG.10/78	34	10	3.4	R	F
W.CG.5/79	83	14	4.5	R	D
W.CG.8/79	39	11	3.5	R	F
W.CG.11/82	9	4	2.3	R	I
W.CG.11/83	63	14	4.5	R	D
W.CG.11/88	6	3	2.0	R	I
W.CG.7/89	30	8	3.8	R	F
W.SW.11/88	87	15	4.5	R	D
W.SW.7/89	46	11	4.2	R	E
W.CR.11/88	50	12	4.2	R	E
W.LU.3/85	78	18	4.3	R	D
W.LD.11/76	73	18	4.1	R	D
W.LD.9/77	87	19	4.6	R	C
W.LD.5/79	102	21	4.9	R	B
W.LD.8/79	84	17	4.8	R	C
W.LD.11/83	181	30	5.4	R	A++
W.LD.9/84	138	29	4.8	R	A
W.LD.3/85	117	24	4.9	R	B
W.LD.7/85	112	24	4.7	R	B
W.LD.6/86	127	22	5.8	R	A++
W.LD.6/87	118	22	5.4	R	A
W.LD.10/88	112	23	4.9	R	B
W.LD.11/88	113	24	4.7	R	B
W.LD.5/89	114	22	5.2	R	A
W.LD.7/89	121	25	4.8	R	A
W.LD.3/90	113	21	5.4	R	A
W.LD.8/90	73	18	4.8	R	C
W.LD.11/90	133	25	5.3	R	A+
W.LD.3/91	108	20	5.4	R	A
W.EU.11/76	95	18	5.3	R	A
W.EU.10/78	80	18	4.4	R	D
W.EU.9/77	92	20	4.6	R	B
W.EU.10/79	105	20	5.3	R	A
W.EU.11/82	137	25	5.5	R	A++
W.EU.2/11/83	122	22	5.5	R	A++
W.EU.3/11/83	144	27	5.3	R	A+
W.ED.11/88	147	28	5.3	R	A+
W.BL.11/76	34	9	3.8	R	E
W.BL.10/78	86	21	4.1	R	D
W.BL.5/79	72	18	4.5	R	D
W.BL.8/79	70	18	4.4	R	D
W.BL.2/83	100	22	4.5	R	C
W.BL.11/83	131	24	5.5	R	A++
W.BL.8/84	130	25	5.2	R	A+
W.BL.7/85	130	26	5.0	R	A
W.BL.6/86	162	26	6.2	R	A++
W.BL.8/87	127	21	6.0	R	A++
W.BL.11/88	161	30	5.4	R	A++
W.BL.6/89	112	22	5.1	R	A
W.BL.11/89	150	25	8.0	R	A++
W.BL.2/90	178	32	5.5	R	A++
W.BL.7/90	169	32	5.3	R	A++
W.BL.5/91	87	18	4.8	R	C

Appendix 2. RIVER GLEN BIOLOGICAL SCORES

SAMPLE	BMWP	NO.SCRQ.TAXA	ASPT	HABITAT TYPE	LQI
G.KB.6/76	72	18	4.0	R	D
G.KB.11/76	62	16	3.9	R	D
G.KB.9/77	75	16	4.2	R	D
G.KB.10/78	94	22	4.3	R	C
G.KB.10/89	110	23	4.6	R	B
G.KB.11/82	120	25	4.8	R	B
G.KB.11/83	128	26	4.9	R	A
G.KB.4/89	134	27	5.0	R	A
G.KB.11/89	144	31	4.6	R	A
G.KB.2/90	137	28	4.9	R	A
G.KB.7/90	158	32	4.9	R	A+
G.KB.10/90	138	30	4.6	R	A
G.KB.3/91	120	27	4.4	R	C
G.TF.11/76	17	5	3.4	RR	F
G.TF.9/77	34	9	3.8	RR	D
G.TF.10/78	75	20	3.8	RR	C
G.TF.10/79	121	26	4.7	RR	A++
G.TF.11/83	67	20	4.4	RR	A
G.TF.7/85	45	10	4.5	RR	B
G.TF.6/86	61	13	4.7	RR	A
G.TF.6/87	104	20	5.2	RR	A++
G.TE.10/78	98	22	4.5	RR	A+
G.TE.5/79	80	18	4.4	RR	B
G.TE.8/79	99	23	4.3	RR	A
G.TE.11/82	82	20	4.1	RR	A
G.TE.11/83	62	16	3.9	RR	C
G.TE.7/85	84	20	4.2	RR	A
G.TE.6/86	50	11	4.5	RR	B
G.TE.6/87	85	19	4.5	RR	A+
G.TE.4/89	69	15	4.6	RR	A
G.TE.11/89	61	20	4.1	RR	B
G.TE.2/90	94	23	4.1	RR	A
G.TE.7/90	110	25	4.4	RR	A+
G.TE.10/90	81	19	4.3	RR	A
G.TE.3/91	82	16	3.9	RR	C
G.GG.6/78	45	10	4.5	RR	B
G.GG.11/78	23	6	3.8	RR	E
G.GG.9/77	23	6	3.8	RR	E
G.GG.10/78	54	13	4.2	RR	B
G.GG.10/79	85	21	4.0	RR	B
G.GG.11/82	73	19	3.8	RR	C
G.GG.11/83	72	18	4.0	RR	C
G.PK.6/78	18	5	3.8	RR	E
G.PK.11/76	23	5	4.6	RR	C
G.PK.9/77	26	7	3.7	RR	D
G.PK.10/78	69	17	4.1	RR	B
G.PK.5/79	69	17	4.1	RR	B
G.PK.8/79	98	22	4.4	RR	A
G.PK.11/82	60	16	3.8	RR	C
G.PK.11/83	76	18	4.2	RR	B
G.SF.5/89	82	19	4.3	RR	A
G.SF.10/89	92	21	4.4	RR	A
G.SF.3/90	68	17	4.0	RR	C
G.SF.8/90	79	20	4.0	RR	C
G.SF.10/90	115	25	4.6	RR	A++
G.SF.4/91	71	18	3.9	RR	C
G.SS.6/78	9	3	3.0	RR	H
G.SS.11/78	8	2	4.0	RR	F
G.SS.9/77	26	7	3.7	RR	D
G.SS.10/78	41	11	3.7	RR	D
G.SS.10/79	61	14	4.4	RR	B
G.SS.11/83	50	12	4.2	RR	C
G.SS.7/84	58	15	3.9	RR	C
G.SS.8/85	63	16	3.9	RR	C
G.SS.8/86	37	10	3.7	RR	D
G.SS.6/87	66	15	4.4	RR	B
G.SS.5/89	67	17	3.9	RR	C
G.SS.10/89	79	19	4.2	RR	B
G.SS.3/90	93	23	4.0	RR	B
G.SS.8/90	97	23	4.2	RR	A
G.SS.10/90	94	22	4.3	RR	A
G.SS.4/91	70	18	3.9	RR	C

Appendix 2. RIVER GLEN BIOLOGICAL SCORES

SAMPLE	BMWP	NO.SCRQ.TAXA	ASPT	HABITAT TYPE	LQI
T.CH.4/82	52	10	5.2	R	C
T.CH.4/84	53	12	4.4	R	E
T.CH.4/87	50	11	4.5	R	E
T.CU.4/84	35	9	3.9	R	E
T.CU.4/87	45	10	4.5	R	E
T.CD.4/82	87	18	4.8	R	C
T.CD.4/84	67	15	4.5	R	D
T.CD.4/87	67	14	4.8	R	C
T.GU.4/82	90	18	5.0	R	C
T.GU.4/84	69	15	4.6	R	C
T.GU.3/85	91	18	5.1	R	S
T.GU.3/90	111	22	5.0	R	B
T.GU.3/91	108	20	5.4	R	A
T.GD.3/85	80	17	4.7	R	C
T.LU.10/78	83	19	4.4	R	D
T.LU.10/79	66	14	4.7	R	C
T.LU.11/83	102	22	4.6	R	B
T.LD.9/77	54	12	4.5	R	E
T.LD.4/82	112	21	5.3	R	A
T.LD.4/84	92	18	5.1	R	A
T.LD.3/85	64	14	4.6	R	C
T.LD.4/87	88	17	5.2	R	B
T.LD.11/88	94	20	4.7	R	B
T.LD.5/89	102	21	4.9	R	B
T.LD.7/89	92	21	4.4	R	C
T.LD.3/90	88	21	4.2	R	D
T.LD.8/90	91	19	4.8	R	B
T.LD.11/90	150	29	5.2	R	A+
T.LD.3/91	44	11	4.0	R	E

B.BU.11/78	17	8	2.8	RR	G
B.BU.10/78	52	15	3.5	RR	D
B.BU.10/79	47	14	3.4	RR	E
B.BD.9/77	33	9	3.7	RR	D
B.BD.10/79	51	15	3.4	RR	D
B.BD.11/82	63	17	3.7	RR	C
B.BD.11/83	55	15	3.7	RR	C
B.RL.10/79	61	17	3.6	RR	C
B.RL.11/82	56	16	3.5	RR	D
B.RL.2/83	36	11	3.3	RR	E
B.RL.11/83	52	15	3.5	RR	D
B.TU.4/89	72	17	4.2	RR	B
B.TU.11/89	66	17	3.9	RR	C
B.TD.2/90	101	22	4.6	RR	A++
B.TD.7/90	94	21	4.5	RR	A+
B.TD.3/91	107	24	4.5	RR	A++

APPENDIX 3 TWINSPAN PREFERENTIAL TAXA

APPENDIX 3.1

TWINSPAN PREFERENTIAL TAXA FOR THE CLASSIFICATION OF ALL SAMPLES

Appendix 3.1

TWINSPAN "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of all samples (Section 5.4, Figures 5 and 6). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 249) I.E. GROUP *
 Eigenvalue .170 at iteration 2
 INDICATORS, together with their SIGN
 Limn ephil(+), Elmi nthil(+), Hydr opsy1(+), Simu liid1(+), Tipu lidal(+), Goer idael(+), Ephe merel(+)
 Coen agril(-), Rhya coph1(+), Valv atid1(-), Ancy lidal(-), Plan aril1(+), Cori xidal(-), Lept ophil(+)

NEGATIVE PREFERENTIALS

OSTR ACOD1(40, 7) CLAD OCER1(41, 4) Coen agril(58, 2) Valv atid1(49, 3)

POSITIVE PREFERENTIALS

Dend rocol(22, 25) Lept ophil(9, 33) Ephe merel(7, 38) Ephe meril(3, 21) Rhya coph1(3, 34) Poly cent1(11, 25)
 Hydr opsy1(24, 55) Hydr optil(29, 33) Limn ephil(14, 64) Goer idael(2, 35) Elmi nthil(72, 84) Tipu lidal(56, 68)
 Simu liid1(21, 55) Ancy lidal(0, 27)

DIVISION 2 (N= 162) I.E. GROUP *0

Eigenvalue .132 at iteration 1
 INDICATORS, together with their SIGN
 Valv atid1(-), Phys idael(-), Plan aril1(-), Coen agril(-), CLAD OCER1(-), Ase1 lidal(-), Plan orbil(-)
 Pisc icoll(-), OSTR ACOD1(-), Baet idael(-), Elmi nthil(-), Gamm arid1(-), Tipu lidal(+), Hydr obil1(-)

NEGATIVE PREFERENTIALS

Plan aril1(53, 22) Pisc icoll(27, 4) OSTR ACOD1(34, 14) CLAD OCER1(32, 9) Ase1 lidal(60, 41) Coen agril(45, 13)
 Hydr optil(23, 6) Noto nect1(20, 10) Valv atid1(49, 0) Phys idael(43, 6)

DIVISION 3 (N= 87) I.E. GROUP *1

Eigenvalue .108 at iteration 2
 INDICATORS, together with their SIGN
 Ephe meril(-), Cori xidal(-), Plan orbil(-), Lept ocer1(-), Phys idael(-), Caen idael(-), Spha erid1(-)
 Dend rocol(-), Poly cent1(-), Rhya coph1(+), Ancy lidal(+), Noto nect1(-)

NEGATIVE PREFERENTIALS

Dend rocol(15, 10) Ephe meril(10, 3) Poly cent1(15, 10) Lept ocer1(21, 16) Seri cost1(11, 6) Gyri nid1(6, 1)
 Cori xidal(24, 15) Noto nect1(9, 1) Phys idael(15, 2) Plan orbil(26, 22)

POSITIVE PREFERENTIALS

Rhya coph1(4, 30) Ancy lidal(3, 24)

NON-PREFERENTIALS

Plan aril1(22, 49) Olig ochal(20, 59) Pisc icoll(13, 20) Glos siph1(24, 49) Erpo bdell1(27, 42) COPE PODAL(6, 11)
 Ase1 lidal(10, 27) Gamm arid1(20, 56) Baet idael(25, 49) Lept ophil(15, 18) Ephe merel(15, 23) Caen idael(26, 29)
 Sial idael(13, 14) Hydr opsy1(20, 35) Hydr optil(13, 20) Limn ephil(20, 44) Goer idael(15, 20) Elmi nthil(20, 56)
 Hall plid1(19, 31) Dyti acid1(22, 44) Helo didal(3, 12) Tipu lidal(20, 48) Simu liid1(15, 40) Chir onom1(25, 58)
 Cera topol(10, 17) Spha erid1(20, 33) Hydr obil1(19, 42) Lymn aeid1(10, 31) HYDR ACAR1(22, 48)

DIVISION 4 (N= 67) I.E. GROUP *00

Eigenvalue .086 at iteration 3
 INDICATORS, together with their SIGN
 Noto nect1(-), COPE PODAL(+), Lept ocer1(-), CLAD OCER1(+), Gerr idael(-), Hydr optil(+)

NEGATIVE PREFERENTIALS

Sial idael(6, 7) Lept ocer1(13, 10) Hydr omet1(4, 1) Gerr idael(7, 3) Noto nect1(14, 6) Unio nid1(5, 3)

POSITIVE PREFERENTIALS

CLAD OCER1(3, 29) COPE PODAL(1, 29) Hydr optil(2, 21)

DIVISION 5 (N= 95) I.E. GROUP *01

Eigenvalue .142 at iteration 2
 INDICATORS, together with their SIGN
 Glos siph1(+), Erpo bdell1(+), Spha erid1(+), Caen idael(+), Elmi nthil(+), Hydr obil1(+), Plan orbil(+)
 Tipu lidal(+), COPE PODAL(-), CLAD OCER1(-)

NEGATIVE PREFERENTIALS

CLAD OCER1(9, 0) COPE PODAL(12, 16)

POSITIVE PREFERENTIALS

Plan aril1(1, 21) Glos siph1(2, 63) Erpo bdell1(5, 62) Ase1 lidal(5, 36) Caen idael(2, 45) Sial idael(1, 23)
 Hydr opsy1(0, 23) Lept ocer1(0, 26) Elmi nthil(4, 49) Tipu lidal(3, 38) Cera topol(3, 24) Spha erid1(1, 45)
 Hydr obil1(3, 49) Plan orbil(3, 46)

DIVISION 6 (N= 20) I.E. GROUP *10

Eigenvalue .084 at iteration 2
 INDICATORS, together with their SIGN
 Psyc homil(-), Ephe merel(+), Poly cent1(+), Sial idael(-), Coen agril(-), Valv atid1(-)

NEGATIVE PREFERENTIALS

OSTR ACOD1(2, 3) CLAD OCER1(1, 3) Coen agril(2, 0) Agri idael(2, 2) Sial idael(4, 9) Mola nnid1(1, 2)
 Helo didal(2, 1) Psyc homil(3, 0) Stra tiom1(1, 0) Taba nid1(1, 0) Musc idael(1, 1) Valv atid1(2, 0)

POSITIVE PREFERENTIALS

Pisc icoll(1, 12) COPE PODAL(0, 6) Ephe merel(0, 15) Poly cent1(0, 15) Psyc homyl(0, 5) Gyri nid1(0, 6)

Appendix 3.1

DIVISION 7 (N= 59) I.E. GROUP *11
Eigenvalue .097 at iteration 2
INDICATORS, together with their SIGN
Ancy lidal(+) Hydr optil(+) Aael lidal(-) Goer idael(+) Plan orbil(-)

NEGATIVE PREFERENTIALS

Aael lidal(15, 12) Sial idael(9, 5) Lept ocerl(10, 6) Psyc nodil(4, 4) Plan orbil(13, 9)

POSITIVE PREFERENTIALS

Dend rocol(1, 9) Pisc icoll(3, 17) Rhya cophl(5, 25) Hydr optil(0, 20) Goer idael(2, 18) Glos aaeol(1, 9)
Ancy lidal(1, 23)

DIVISION 8 (N= 16) I.E. GROUP *000

Eigenvalue .108 at iteration 3
INDICATORS, together with their SIGN

Plan aril(+)

NEGATIVE PREFERENTIALS

Ephe merel(2, 0)

POSITIVE PREFERENTIALS

Plan aril(0, 11) Glos siphil(1, 11) Erpo bdell(0, 8) OSTR ACODl(0, 7) Sial idael(1, 5) Phry ganel(0, 3)
Elmi nthil(1, 5) Gyri nidal(0, 3) Hydr ophil(0, 3) Helo didal(0, 3) Hydr ometl(0, 4) Spha eridl(1, 8)
Phya idael(2, 9)

DIVISION 9 (N= 51) I.E. GROUP *001

Eigenvalue .081 at iteration 2
INDICATORS, together with their SIGN

Coen agril(-) Erpo bdell(+) Cori xidal(-) Phys idael(-) Caen idael(-) Dend rocol(+) Hydr optil(+)
Poly centl(+)

NEGATIVE PREFERENTIALS

Coen agril(30, 2) Caen idael(23, 2) Cori xidal(37, 5)

POSITIVE PREFERENTIALS

Dend rocol(2, 6) Erpo bdell(17, 13) Sial idael(4, 3) Poly centl(1, 5) Hydr optil(12, 9) Tipu lidal(6, 5)
Simu liidl(0, 3) Musc idael(1, 3) Phys idael(19, 13)

DIVISION 10 (N= 23) I.E. GROUP *010

Eigenvalue .260 at iteration 3
INDICATORS, together with their SIGN

Dyti scidl(+)

NEGATIVE PREFERENTIALS

Erpo bdell(3, 2)

POSITIVE PREFERENTIALS

CLAD OCERl(1, 8)

DIVISION 11 (N= 72) I.E. GROUP *011

Eigenvalue .096 at iteration 2
INDICATORS, together with their SIGN

Cera topol(+) Lept ocerl(+) Spha eridl(+) Simu liidl(-) OSTR ACODl(-) Hydr opsyil(+) Dend rocol(-)
Phya idael(-) Sial idael(+) HYDR ACARl(+) Coen agril(+) COPE PODAl(-) Baet idael(-)

NEGATIVE PREFERENTIALS

Dend rocol(7, 5) OSTR ACODl(9, 3) Simu liidl(10, 5) Phys idael(5, 1)

POSITIVE PREFERENTIALS

Coen agril(0, 11) Sial idael(3, 20) Hydr opsyil(2, 21) Lept ocerl(1, 25) Cera topol(1, 23) Spha eridl(6, 39)

DIVISION 12 (N= 4) I.E. GROUP *100

Eigenvalue .120 at iteration 4
INDICATORS, together with their SIGN

Simu liidl(+)

NEGATIVE PREFERENTIALS

Goer idael(1, 1) Cera topol(1, 1) Taba nidal(1, 0)

POSITIVE PREFERENTIALS

Dend rocol(0, 2) Pisc icoll(0, 1) OSTR ACODl(0, 2) CLAD OCERl(0, 1) Coen agril(0, 2) Agri idael(0, 2)
Ephe meril(0, 2) Hydr opsyil(0, 2) Hydr optil(0, 2) Mola nnidl(0, 1) Seri costl(0, 1) Halli plidl(0, 2)
Helo didal(0, 2) Noto nectl(0, 1) Simu liidl(0, 3) Stra tioml(0, 1) Musc idael(0, 1) Valv atidl(0, 2)
Phya idael(0, 3)

DIVISION 13 (N= 24) I.E. GROUP *101

Eigenvalue .093 at iteration 3
INDICATORS, together with their SIGN

Seri costl(-) Poly centl(+)

NEGATIVE PREFERENTIALS

Rhya cophl(3, 1) Psyc homyl(4, 1) Seri costl(10, 0) Gyri nidal(4, 2) Ancy lidal(3, 0)

POSITIVE PREFERENTIALS

PORI FERAl(0, 3) CLAD OCERl(0, 3) COPE PODAl(2, 4) Poly centl(3, 12) Hydr optil(3, 8) Simu liidl(4, 8)
Cera topol(1, 7)

Appendix 3.1

DIVISION 14 (N= 18) I.E. GROUP *110

Eigenvalue .120 at iteration 2

INDICATORS, together with their SIGN

Lept ophil(+) COPE PODAL(+) Cera topol(-) Pisc icoll(+)

NEGATIVE PREFERENTIALS

Poly centl(2, 0) Cera topol(7, 0) Psyc hodil(3, 1) Tapa nical(2, 0) Lymn aeidl(5, 2)

POSITIVE PREFERENTIALS

Pisc icoll(0, 3) COPE PODAL(1, 4) Lept ophil(0, 7) Goer idael(0, 2) Seri costl(0, 2) Helo didal(0, 3)
Cori xidal(1, 4)

.....

DIVISION 15 (N= 41) I.E. GROUP *111

Eigenvalue .101 at iteration 2

INDICATORS, together with their SIGN

Spha eridl(-) Ephe merel(-) Caen idael(-) Dend rocol(-) Asel lidal(-) Expo bdell(-) Ancy lidal(-)
Plan orbil(-) Cori xidal(-) Lept ocerl(-) Simu liidl(-) Sial idael(-) Limn ephil(+)
Lymn aeidl(-) Helo didal(+)

NEGATIVE PREFERENTIALS

Dend rocol(9, 0) Asel lidal(10, 2) Ephe merel(13, 4) Caen idael(14, 5) Sial idael(5, 0) Lept ocerl(6, 0)
Cori xidal(8, 2) Spha eridl(16, 6) Plan orbil(8, 1)

POSITIVE PREFERENTIALS

Hel o didal(2, 7)

.....

APPENDIX 3.2

TWINSPAN PREFERENTIAL TAXA FOR THE CLASSIFICATIONS OF THE CASE-STUDY SITES

3.2.1 West Glen at Essendine

3.2.2 West Glen at Banthorpe Lodge

3.2.3 Glen at Kates Bridge

3.2.4 Glen at Surfleet Seas End

3.2.4 East Glen at Braceborough

Appendix 3.2.1 Indicator and preferential taxa for WEU

TWINSpan "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of WEU samples (Section 5.6.1, Figure 11). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 7) I.E. GROUP *

Eigenvalue .222 at iteration 2
INDICATORS, together with their SIGN
Hydr opayl(-)

NEGATIVE PREFERENTIALS

Lept ophil(3, 0) Ephe merel(3, 0) Ephe meril(5, 0) Seri costl(3, 0) Dend rocol(4, 0) Hydr opayl(6, 0)
Dyti scidl(5, 0) Cori xidal(6, 0) Noto nectl(2, 0) Simu liidl(3, 0) Pisc icoll(3, 0) Glos siph(5, 0)
Erpo bdeil(6, 0) Cera topol(3, 0)

POSITIVE PREFERENTIALS

Limn ephil(2, 1) Hydr optil(2, 1) Hali plidl(2, 1) Tipu lidal(2, 1) Sial idael(2, 1) Aseil lidal(0, 1)
Hydr obil(0, 1) Lymn aeidl(1, 1) Phys idael(2, 1) COEL ENTEL(0, 1)

DIVISION 2 (N= 6) I.E. GROUP *0

Eigenvalue .211 at iteration 1
INDICATORS, together with their SIGN
Dend rocol(+)

NEGATIVE PREFERENTIALS

Seri costl(2, 1) Gyri nidal(1, 0) Hydr ophil(1, 0) Tipu lidal(1, 1) Sial idael(2, 0) COPE PODAL(1, 0)

POSITIVE PREFERENTIALS

Lept ophil(0, 3) Mola nnidl(0, 1) Rhya cophl(0, 1) Limn ephil(0, 2) Hydr optil(0, 2) Plan ariil(1, 4)
Dend rocol(0, 4) Hali plidl(0, 2) Noto nectl(0, 2) Simu liidl(0, 3) Pisc icoll(0, 3) Lymn aeidl(0, 1)
Phys idael(0, 2) Plan orbil(0, 4) PORI FERAL(0, 1) HYDR ACARl(1, 4)

DIVISION 4 (N= 2) I.E. GROUP *00

Eigenvalue .219 at iteration 0
INDICATORS, together with their SIGN
Ephe merel(-)

NEGATIVE PREFERENTIALS

Ephe merel(1, 0) Plan ariil(1, 0) Hydr ophil(1, 0) Tipu lidal(1, 0)

POSITIVE PREFERENTIALS

Lept ocerl(0, 1) Gyri nidal(0, 1) COPE PODAL(0, 1) Cera topol(0, 1) HYDR ACARl(0, 1)

DIVISION 5 (N= 4) I.E. GROUP *01

Eigenvalue .145 at iteration 6
RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .026 INSTEAD OF .003 (THE TOLERANCE)
INDICATORS, together with their SIGN
Lept ophil(-)

NEGATIVE PREFERENTIALS

Lept ophil(3, 0) Ephe merel(2, 0) Ephe meril(3, 0) Mola nnidl(1, 0) Lept ocerl(3, 0) Seri costl(1, 0)
Rhya cophl(1, 0) Limn ephil(2, 0) Noto nectl(2, 0) Tipu lidal(1, 0) Simu liidl(3, 0) Pisc icoll(3, 0)
Lymn aeidl(1, 0) Phys idael(2, 0) PORI FERAL(1, 0) Cera topol(2, 0)

POSITIVE PREFERENTIALS

Hydr optil(1, 1) Hali plidl(1, 1)

DIVISION 10 (N= 3) I.E. GROUP *010

Eigenvalue .178 at iteration 6
RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .045 INSTEAD OF .003 (THE TOLERANCE)
INDICATORS, together with their SIGN
Ephe merel(+)

NEGATIVE PREFERENTIALS

Mola nnidl(1, 0) Seri costl(1, 0) Noto nectl(1, 1) Glos siph(1, 1) Phys idael(1, 1) Cera topol(1, 1)

POSITIVE PREFERENTIALS

Ephe merel(0, 2) Rhya cophl(0, 1) Poly centl(0, 2) Limn ephil(0, 2) Hydr optil(0, 1) Hali plidl(0, 1)
Dyti scidl(0, 2) Tipu lidal(0, 1) Lymn aeidl(0, 1) Chir onoml(0, 2) PORI FERAL(0, 1)

Appendix 3.2.2 Indicator and preferential taxa for WBL

TWINSpan "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of WBL samples (Section 5.6.2, Figure 11). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 16) I.E. GROUP *

Eigenvalue .164 at iteration 1

INDICATORS, together with their SIGN

Goer idael(-) Ephe meril(-) Seri costl(-)

NEGATIVE PREFERENTIALS

Caen idael(10, 2) Spha eridl(10, 2) Goer idael(9, 0) Limn ephil(6, 1) Hydr opsy(8, 1) Cori xidal(8, 1)

Ephe meril(8, 0) Lept ocerl(7, 1) Seri costl(7, 0) Poly centl(6, 1) Phya idael(6, 1) Hydr optil(5, 1)

Lept ophil(5, 0) Pisc icoll(5, 0) Sial idael(5, 0) Ancy lidal(4, 0) Lepi dostl(3, 0) Agri idael(3, 0)

POSITIVE PREFERENTIALS

Cera topol(4, 4) COPE PODal(1, 2) Stra tioml(0, 2) Musc idael(0, 2)

DIVISION 2 (N= 11) I.E. GROUP *0

Eigenvalue .132 at iteration 1

INDICATORS, together with their SIGN

Dend rocol(+) Poly centl(+)

NEGATIVE PREFERENTIALS

Plan orbil(6, 2) Hydr obil(6, 2) Cori xidal(6, 2) Seri costl(5, 2) Lymn aedl(4, 1) Phya idael(5, 1)

Asel lidal(4, 0) Ancy lidal(3, 1) Agri idael(3, 0) Mola nnidl(2, 0) Melo didal(2, 0)

POSITIVE PREFERENTIALS

Simu liidl(2, 4) Dend rocol(1, 5) Poly centl(1, 5) Rhya cophl(1, 3)

DIVISION 3 (N= 5) I.E. GROUP *1

Eigenvalue .308 at iteration 1

INDICATORS, together with their SIGN

Gamm aridl(-)

NEGATIVE PREFERENTIALS

Gamm aridl(4, 0) Baet idael(4, 0) Caen idael(2, 0) Glos siph(4, 0) Spha eridl(2, 0)

Hydr obil(4, 0) Plan ariil(3, 0) HYDR ACARl(4, 0) Heli plidl(2, 0) Ephe meril(2, 0) Limn ephil(1, 0)

Hydr opsy(1, 0) Cori xidal(1, 0) Cera topol(4, 0) Lept ocerl(1, 0) Poly centl(1, 0) Phya idael(1, 0)

Rhya cophl(2, 0) Hydr optil(1, 0) Asel lidal(2, 0) Psyc hodil(1, 0) Stra tioml(2, 0) Musc idael(2, 0)

Glos sosol(1, 0) COEL ENTEL(1, 0) Taba nidal(1, 0) Nemo uridl(1, 0) Hydr ophil(1, 0)

POSITIVE PREFERENTIALS

Dyti acidl(2, 1) Dend rocol(1, 1) Lymn aedl(1, 1) COPE PODal(1, 1) OSTR ACODl(0, 1)

DIVISION 4 (N= 6) I.E. GROUP *00

Eigenvalue .181 at iteration 1

INDICATORS, together with their SIGN

Caen idael(+)

NEGATIVE PREFERENTIALS

Ephe meril(1, 2) Lept ocerl(1, 2) Dend rocol(1, 0) Hydr optil(1, 1) Pisc icoll(1, 2) Sial idael(1, 2)

Ancy lidal(1, 2) Leuc tridl(1, 0) Mola nnidl(1, 1) Melo didal(1, 1) Veli idael(1, 0) Gerr idael(1, 0)

POSITIVE PREFERENTIALS

Caen idael(0, 5) HYDR ACARl(0, 3) Simu liidl(0, 2) Goer idael(0, 5) Limn ephil(0, 4) Cera topol(0, 2)

Asel lidal(0, 4) Lept ophil(0, 3) Lepi dostl(0, 2) Agri idael(0, 3)

DIVISION 5 (N= 5) I.E. GROUP *01

Eigenvalue .221 at iteration 1

INDICATORS, together with their SIGN

Tipu lidal(-)

NEGATIVE PREFERENTIALS

Tipu lidal(3, 0) Simu liidl(3, 1) Ephe meril(3, 1) Limn ephil(3, 1) Seri costl(2, 0) Lymn aedl(1, 0)

Rhya cophl(-3, 0) Ancy lidal(-1, 0) Lepi dostl(-1, 0) Leuc tridl(-1, 0) Glos sosol(1, 0) Gyri nidal(-1, 0)

POSITIVE PREFERENTIALS

Glos siph(1, 2) HYDR ACARl(1, 2) Cera topol(0, 2) Ephe meril(1, 2) Phya idael(0, 1) Pisc icoll(0, 2)

COPE PODal(0, 1) Veli idael(0, 1) PORI FERAl(0, 1)

DIVISION 6 (N= 4) I.E. GROUP *10

Eigenvalue .248 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .007 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

Cori xidal(+)

NEGATIVE PREFERENTIALS

Caen idael(2, 0) Ephe meril(2, 0) Limn ephil(1, 0) Hydr opsy(1, 0) Lept ocerl(1, 0) Dend rocol(1, 0)

Hydr optil(1, 0) COPE PODal(1, 0) Psyc hodil(1, 0) Stra tioml(2, 0) Glos sosol(1, 0) Taba nidal(1, 0)

Nemo uridl(1, 0) Hydr ophil(1, 0)

POSITIVE PREFERENTIALS

Dyti acidl(1, 1) Spha eridl(1, 1) Heli plidl(1, 1) Cori xidal(0, 1) Poly centl(0, 1) Lymn aedl(0, 1)

Phya idael(0, 1) Rhya cophl(1, 1) Asel lidal(1, 1) Musc idael(1, 1) COEL ENTEL(0, 1)

Appendix 3.2.2

DIVISION 9 (N= 5) I.E. GROUP *001
Eigenvalue .198 at iteration 1
INDICATORS, together with their SIGN
Simu liid1(-)

NEGATIVE PREFERENTIALS

Plan arii1(2, 1) HYDR ACAR1(2, 1) Simu liid1(2, 0) Cera topol(2, 0) Hydr optil(1, 0) Psyc hodil(1, 0)
Mole nnid1(1, 0) Helo didal(1, 0) Coen agril(1, 0) Noto nect1(1, 0)

POSITIVE PREFERENTIALS

Ephe merel(0, 2) Ephe meril(1, 3) Seri cost1(1, 3) Poly cent1(0, 1) Lymn aeid1(0, 3) Rhys coph1(0, 1)
Pisc icoll(0, 2) Lepi dost1(0, 2) OSTR ACOD1(0, 1) Gyri nid1(0, 1)

DIVISION 10 (N= 3) I.E. GROUP *010
Eigenvalue .269 at iteration 2
INDICATORS, together with their SIGN
Plan orbil(-)

NEGATIVE PREFERENTIALS

Plan orbil(1, 0) Dyti acid1(1, 1) Glos siph1(1, 0) Spha erid1(1, 1) Hydr obil1(1, 0) Hali plid1(1, 1)
Hydr opsy1(1, 1) Cori xidal(1, 0) Seri cost1(1, 1) Lymn aeid1(1, 0) Hydr optil(1, 1) Gyri nid1(1, 0)

POSITIVE PREFERENTIALS

Plan arii1(0, 2) HYDR ACAR1(0, 1) Goer idael(0, 2) Ephe meril(0, 1) Lept ocer1(0, 2) Lept oph1(0, 1)
Sial idael(0, 1) Ancy lidal(0, 1) Lepi dost1(0, 1) Leuc trid1(0, 1) Glos sosol(0, 1)

DIVISION 11 (N= 2) I.E. GROUP *011
Eigenvalue .236 at iteration 0
INDICATORS, together with their SIGN
Plan orbil(-)

NEGATIVE PREFERENTIALS

Plan orbil(1, 0) Hydr obil1(1, 0) Simu liid1(1, 0) Ephe merel(1, 0) Sial idael(1, 0) Veli idael(1, 0)

POSITIVE PREFERENTIALS

Linn eph1(0, 1) Cori xidal(0, 1) Phys idael(0, 1) Hydr optil(0, 1) Lept oph1(0, 1) COPE PODA1(0, 1)
PORI FERAL(0, 1)

DIVISION 12 (N= 3) I.E. GROUP *100
Eigenvalue .362 at iteration 2
INDICATORS, together with their SIGN
Caen idael(+)

NEGATIVE PREFERENTIALS

Tipu lidal(1, 1) Ephe merel(1, 1) Hydr opsy1(1, 0) Dend rocol(1, 0) Rhys coph1(1, 0) Asei lidal(1, 0)
Hydr oph1(1, 0)

POSITIVE PREFERENTIALS

Caen idael(0, 2) Dyti acid1(0, 1) Spha erid1(0, 1) Plan arii1(0, 2) Hali plid1(0, 1) Simu liid1(0, 2)
Linn eph1(0, 1) Lept ocer1(0, 1) Hydr optil(0, 1) COPE PODA1(0, 1) Psyc hodil(0, 1) Stra tiom1(0, 2)
Musc idael(0, 1) Glos sosol(0, 1) Taba nid1(0, 1) Nemo urid1(0, 1)

Appendix 3.2.3 Indicator and preferential taxa for GKB

TWINSpan "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of GKB samples (Section 5.6.3, Figure 13). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 13) I.E. GROUP *

Eigenvalue .118 at iteration 2

INDICATORS, together with their SIGN

Simu liid1(-) Gyri nidal(+)

NEGATIVE PREFERENTIALS

Lept ophil(2, 0) Goer idael(2, 0) Psyc homyl(2, 0) Simu liid1(8, 0) PORI FERAL(3, 0)

POSITIVE PREFERENTIALS

Ephe meril(1, 1) Ephe meril(2, 2) Mola nnid1(0, 1) Coen agril(2, 2) Gyri nidal(0, 3) Vell idael(0, 1)

Hydr omet1(0, 2) Gerr idael(0, 2) Noto nect1(1, 3) Sial idael(4, 3) Valv atid1(3, 2) Musc idael(3, 2)

.....

DIVISION 2 (N= 10) I.E. GROUP *0

Eigenvalue .129 at iteration 1

INDICATORS, together with their SIGN

Sial idael(-)

NEGATIVE PREFERENTIALS

Lept ophil(2, 0) Ephe meril(2, 0) Lept ocer1(4, 3) Agri idael(1, 0) Limn ephil(3, 2) Dend rocol(4, 2)

Elmi nthil(4, 3) Helo didal(1, 0) Tipu lidal(4, 3) Sial idael(4, 0) Valv atid1(2, 1) NEMA TOMO1(1, 0)

Psyc hodil(1, 0) Stra tiom1(1, 0) Musc idael(2, 1)

POSITIVE PREFERENTIALS

Goer idael(0, 2) Pisc icoll(1, 3) Lynn asid1(2, 6) PORI FERAL(0, 3) CLAD OCER1(1, 4) COPE PODAL(1, 3)

Cera topol(0, 4)

.....

DIVISION 3 (N= 3) I.E. GROUP *1

Eigenvalue .076 at iteration 0

INDICATORS, together with their SIGN

Ephe meril(-)

NEGATIVE PREFERENTIALS

Ephe meril(1, 0) Ephe meril(2, 0) Poly cent1(2, 0) Limn ephil(1, 0) Hydr opsy1(2, 0) Vell idael(1, 0)

Pisc icoll(1, 0) COPE PODAL(1, 0)

POSITIVE PREFERENTIALS

Mola nnid1(0, 1) Coen agril(1, 1) Hydr omet1(1, 1) Gerr idael(1, 1) Tipu lidal(1, 1) Valv atid1(1, 1)

OSTR ACOD1(1, 1) CLAD OCER1(1, 1) Cera topol(1, 1) Musc idael(1, 1) HYDR ACAR1(1, 1)

.....

DIVISION 4 (N= 4) I.E. GROUP *00

Eigenvalue .093 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .026 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

Hydr optil(+)

NEGATIVE PREFERENTIALS

Lept ophil(1, 1) Ephe meril(1, 1) Poly cent1(1, 1) Hydr opsy1(1, 1) COPE PODAL(1, 0)

POSITIVE PREFERENTIALS

Agri idael(0, 1) Psyc homyl(0, 1) Coen agril(0, 1) Hydr optil(0, 3) Helo didal(0, 1) Pisc icoll(0, 1)

Valv atid1(0, 2) Lynn asid1(0, 2) NEMA TOMO1(0, 1) CLAD OCER1(0, 1) Psyc hodil(0, 1) Stra tiom1(0, 1)

Musc idael(0, 2)

.....

DIVISION 5 (N= 6) I.E. GROUP *01

Eigenvalue .191 at iteration 1

INDICATORS, together with their SIGN

Simu liid1(-)

NEGATIVE PREFERENTIALS

Ephe meril(1, 0) Lept ocer1(3, 0) Goer idael(2, 0) Psyc homyl(1, 0) Poly cent1(3, 0) Gamm arid1(4, 1)

Hydr optil(3, 0) Hydr opsy1(2, 0) Elmi nthil(3, 0) Simu liid1(4, 0) Pisc icoll(3, 0) Glos siph1(4, 1)

Phys idael(4, 0) Plan orbil(4, 1) Musc idael(1, 0)

POSITIVE PREFERENTIALS

Limn ephil(1, 1) Coen agril(0, 1) Dend rocol(1, 1) Hall plid1(2, 2) Noto nect1(0, 1) Valv atid1(0, 1)

OSTR ACOD1(2, 2) CLAD OCER1(2, 2) Cera topol(2, 2)

.....

DIVISION 6 (N= 2) I.E. GROUP *10

Eigenvalue .213 at iteration 0

INDICATORS, together with their SIGN

Ephe meril(+)

NEGATIVE PREFERENTIALS

Coen agril(1, 0) Tipu lidal(1, 0) Pisc icoll(1, 0) Valv atid1(1, 0) OSTR ACOD1(1, 0) CLAD OCER1(1, 0)

COPE PODAL(1, 0) Cera topol(1, 0) Musc idael(1, 0) HYDR ACAR1(1, 0)

POSITIVE PREFERENTIALS

Ephe meril(0, 1) Limn ephil(0, 1) Vell idael(0, 1) Hydr omet1(0, 1) Gerr idael(0, 1)

.....

Appendix 3.2.3

DIVISION 9 (N= 3) I.E. GROUP *001

Eigenvalue .212 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .016 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

Coen agril(-)

NEGATIVE PREFERENTIALS

Limn ephil(1, 1) Coen agril(1, 0) Hydr opsy1(1, 0) Dyti scidi(1, 1) Melo didal(1, 0) Pisc icoll(1, 0)
Spha eridl(1, 1) Valv acidl(1, 1) Hydr obil1(1, 1) Lymn aeidl(1, 1) Psyc hodil(1, 0) Stra tioml(1, 0)
Musc idael(1, 1)

POSITIVE PREFERENTIALS

Lept ophil(0, 1) Ephe meril(0, 1) Agri idael(0, 1) Psyc homyl(0, 1) Poly cent1(0, 1) Hali plidl(0, 2)
Cori xidal(0, 2) Baet idael(0, 2) NEMA TOMO1(0, 1) CLAD OCER1(0, 1)

DIVISION 10 (N= 4) I.E. GROUP *010

Eigenvalue .092 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .019 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

PORI FERAl(-)

NEGATIVE PREFERENTIALS

Lept ocer1(2, 1) Psyc homyl(1, 0) Poly cent1(2, 1) Hyar optil(2, 1) Dend rocol(1, 0) Erpe bdell(2, 1)
Spha eridl(2, 1) Chir onoml(2, 1) PORI FERAl(2, 0) Musc idael(1, 0)

POSITIVE PREFERENTIALS

Ephe merel(0, 1) Caen idael(1, 2) Limn ephil(0, 1) Elmi nthil(1, 2) Pisc icoll(1, 2)

DIVISION 11 (N= 2) I.E. GROUP *011

Eigenvalue .267 at iteration 0

INDICATORS, together with their SIGN

Caen idael(-)

NEGATIVE PREFERENTIALS

Caen idael(1, 0) Limn ephil(1, 0) Dend rocol(1, 0) Noto nect1(1, 0) Glos siph1(1, 0) Plan orbil(1, 0)
PORI FERAl(1, 0) COPE PODAl(1, 0)

POSITIVE PREFERENTIALS

Gamm aridl(0, 1) Coen agril(0, 1) Tipu lidal(0, 1) Valv acidl(0, 1)

Appendix 3.2.4 Indicator and preferential taxa for GSS.

TWINSpan "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of GSS samples (Section 5.6.4, Figure 14). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 16) I.E. GROUP *
Eigenvalue .187 at iteration 1
INDICATORS, together with their SIGN
Coen agril(-) Lymn aeidl(-) Valv atidl(-) Aael lidal(-)

NEGATIVE PREFERENTIALS

Lept ocer1(2, 0) Rhya coph1(2, 0) Coen agril(9, 2) Unio nidal(2, 0) Plan ari11(8, 1) Elmi nth11(5, 0)
Hali plid1(7, 2) Dyt1 scid1(8, 3) Noto nect1(3, 0) Tipu lidal(2, 0) Pisc icoll(5, 1) Glos siph1(6, 1)
Erpo bdel1(4, 1) Aael lidal(6, 1) Spha eridl(6, 0) Valv atidl(9, 1) Lymn aeidl(8, 1) Phys idae1(3, 0)
Mys1 dae 1(2, 0) Cera topo1(3, 1) Lept ocer2(2, 0) Rhya coph2(2, 0) Coen agril2(9, 2) Unio nida2(2, 0)
Plan ari12(8, 1) Elmi nth12(5, 0) Hali plid2(7, 2) Dyt1 scid2(8, 3) Noto nect2(3, 0) Tipu lida2(2, 0)
Pisc icol2(5, 1) Glos siph2(6, 1) Erpo bdel2(4, 1) Aael lida2(6, 1) Spha erid2(6, 0) Valv atid2(9, 1)
Lymn aeidl2(8, 1) Phys idae2(3, 0) Mys1 dae 2(2, 0) Cera topo2(3, 1)

POSITIVE PREFERENTIALS

CLAD OCER1(3, 5) COPE PODA1(2, 4) CLAD OCER2(3, 5) COPE PODA2(2, 4)

DIVISION 2 (N= 9) I.E. GROUP *0
Eigenvalue .163 at iteration 2
INDICATORS, together with their SIGN
Phys idae1(-) Glos siph1(-) Aael lidal(-) Spha eridl(-)

NEGATIVE PREFERENTIALS

Phys homy1(1, 0) Rhya coph1(2, 0) Deno rocol(1, 0) Helo didal(1, 0) Glos siph1(4, 2)
Erpo bdel1(3, 1) Aael lidal(4, 2) Spha eridl(4, 2) Phys idae1(3, 0) OSTR ACOD1(3, 1) COPE PODA1(2, 0)
Coro phid1(1, 0) Pyra lidal(1, 0) Phys homy2(1, 0) Caen idae2(2, 1) Rhya coph2(2, 0) Dend roco2(1, 0)
Helo dida2(1, 0) Glos siph2(4, 2) Erpo bdel2(3, 1) Aael lida2(4, 2) Spha erid2(4, 2) Phys idae2(3, 0)
OSTR ACOD2(3, 1) COPE PODA2(2, 0) Coro phid2(1, 0) Pyra lida2(1, 0)

POSITIVE PREFERENTIALS

Lept ocer1(0, 2) Gerr idae1(0, 2) Tipu lidal(0, 2) Pisc icoll(1, 4) CLAD OCER1(0, 3) Lept ocer2(0, 2)
Gerr idae2(0, 2) Tipu lida2(0, 2) Pisc icol2(1, 4) CLAD OCER2(0, 3)

DIVISION 3 (N= 7) I.E. GROUP *1
Eigenvalue .255 at iteration 3
INDICATORS, together with their SIGN
Caen idae1(+)

NEGATIVE PREFERENTIALS

Hali plid1(2, 0) Hali plid2(2, 0)

POSITIVE PREFERENTIALS

Caen idae1(0, 2) Poly cent1(0, 1) Coen agril(0, 2) Hydr optil(0, 1) Plan ari11(0, 1) Gerr idae1(1, 1)
Erpo bdel1(0, 1) Aael lidal(0, 1) Valv atidl(0, 1) Hydr obi11(2, 2) OSTR ACOD1(1, 1) COPE PODA1(2, 2)
Cera topo1(0, 1) Caen idae2(0, 2) Poly cent2(0, 1) Coen agril2(0, 2) Hydr opti2(0, 1) Plan ari12(0, 1)
Gerr idae2(1, 1) Erpo bdel2(0, 1) Aael lida2(0, 1) Valv atid2(0, 1) Hydr obi12(2, 2) OSTR ACOD2(1, 1)
COPE PODA2(2, 2) Cera topo2(0, 1)

DIVISION 4 (N= 4) I.E. GROUP *00
Eigenvalue .120 at iteration 6
RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .043 INSTEAD OF .003 (THE TOLERANCE)
INDICATORS, together with their SIGN
Hali plid1(+)

NEGATIVE PREFERENTIALS

Caen idae1(1, 1) COPE PODA1(1, 1) Caen idae2(1, 1) COPE PODA2(1, 1)

POSITIVE PREFERENTIALS

Phys homy1(0, 1) Rhya coph1(0, 2) Hydr optil(0, 1) Unio nidal(0, 1) Dend rocol(0, 1) Hali plid1(0, 3)
Helo didal(0, 1) Noto nect1(0, 1) Pisc icoll(0, 1) Erpo bdel1(0, 3) Phys idae1(0, 3) Coro phid1(0, 1)
Mys1 dae 1(0, 1) Pyra lidal(0, 1) Cera topo1(0, 1) Phys homy2(0, 1) Rhya coph2(0, 2) Hydr opti2(0, 1)
Unio nida2(0, 1) Dend roco2(0, 1) Hali plid2(0, 3) Helo dida2(0, 1) Noto nect2(0, 1) Pisc icol2(0, 1)
Erpo bdel2(0, 3) Phys idae2(0, 3) Coro phid2(0, 1) Mys1 dae 2(0, 1) Pyra lida2(0, 1) Cera topo2(0, 1)

DIVISION 5 (N= 5) I.E. GROUP *01
Eigenvalue .238 at iteration 1
INDICATORS, together with their SIGN
Spha eridl(+)

NEGATIVE PREFERENTIALS

Lept ocer1(2, 0) Unio nidal(1, 0) Elmi nth11(2, 0) Hydr ophi1(1, 0) Meso vel11(1, 0) Gerr idae1(2, 0)
Noto nect1(2, 0) Glos siph1(2, 0) Erpo bdel1(1, 0) OSTR ACOD1(1, 0) HYDR ACAR1(2, 0) Lept ocer2(2, 0)
Unio nida2(1, 0) Elmi nth12(2, 0) Hydr ophi2(1, 0) Meso vel12(1, 0) Gerr idae2(2, 0) Noto nect2(2, 0)
Glos siph2(2, 0) Erpo bdel2(1, 0) OSTR ACOD2(1, 0) HYDR ACAR2(2, 0)

POSITIVE PREFERENTIALS

Caen idae1(0, 1) Hydr optil(0, 1) Spha eridl(0, 2) CLAD OCER1(1, 2) Mys1 dae 1(0, 1) Caen idae2(0, 1)
Hydr opti2(0, 1) Spha erid2(0, 2) CLAD OCER2(1, 2) Mys1 dae 2(0, 1)

Appendix 3.2.4

DIVISION 6 (N= 5) I.E. GROUP *10

Eigenvalue .285 at iteration 1

INDICATORS, together with their SIGN

Pisc icoll(-)

NEGATIVE PREFERENTIALS

Hali plid1(1, 1) Dyti scid1(1, 1) Cori xida1(1, 2) Pisc icoll1(1, 0) Baet idae1(1, 2) Hydr obil1(1, 1)
 Lymn aeid1(1, 0) Plan orb1(1, 2) OSTR ACOD1(1, 0) CLAD OCER1(1, 2) Hali plid2(1, 1) Dyti scid2(1, 1)
 Cori xida2(1, 2) Pisc icoll2(1, 0) Baet idae2(1, 2) Hydr obil2(1, 1) Lymn aeid2(1, 0) Plan orb12(1, 2)
 OSTR ACOD2(1, 0) CLAD OCER2(1, 2)

POSITIVE PREFERENTIALS

Gerr idae1(0, 1) Glos siph1(0, 1) COPE PODA1(0, 2) HYDR ACAR1(0, 3) Gerr idae2(0, 1) Glos siph2(0, 1)
 COPE PODA2(0, 2) HYDR ACAR2(0, 3)

DIVISION 7 (N= 2) I.E. GROUP *11

Eigenvalue .318 at iteration 0

INDICATORS, together with their SIGN

Poly cent1(+)

NEGATIVE PREFERENTIALS

Plan aril1(1, 0) Gerr idae1(1, 0) Valv atid1(1, 0) Plan aril2(1, 0) Gerr idae2(1, 0) Valv atid2(1, 0)

POSITIVE PREFERENTIALS

Poly cent1(0, 1) Hydr opt11(0, 1) Dyti scid1(0, 1) Erpo bdell1(0, 1) Asel lida1(0, 1) OSTR ACOD1(0, 1)
 Cera topo1(0, 1) HYDR ACAR1(0, 1) Poly cent2(0, 1) Hydr opt12(0, 1) Dyti scid2(0, 1) Erpo bdell2(0, 1)
 Asel lida2(0, 1) OSTR ACOD2(0, 1) Cera topo2(0, 1) HYDR ACAR2(0, 1)

DIVISION 9 (N= 3) I.E. GROUP *001

Eigenvalue .255 at iteration 4

INDICATORS, together with their SIGN

Caen idae1(-)

NEGATIVE PREFERENTIALS

Caen idae1(1, 0) Dend rocol1(1, 0) Elmi nth1(1, 1) Dyti scid1(1, 1) Helo dida1(1, 0) Noto nect1(1, 0)
 Lymn aeid1(1, 1) Coro phid1(1, 0) Caen idae2(1, 0) Dend rocol2(1, 0) Elmi nth2(1, 1) Dyti scid2(1, 1)
 Helo dida2(1, 0) Noto nect2(1, 0) Lymn aeid2(1, 1) Coro phid2(1, 0)

POSITIVE PREFERENTIALS

Payc homy1(0, 1) Rhya coph1(0, 2) Hydr opt11(0, 1) Unio nida1(0, 1) Pisc icoll1(0, 1) OSTR ACOD1(0, 2)
 COPE PODA1(0, 1) Mysi dae 1(0, 1) Pyra lida1(0, 1) Cera topo1(0, 1) HYDR ACAR1(0, 2) Payc homy2(0, 1)
 Rhya coph2(0, 2) Hydr opt12(0, 1) Unio nida2(0, 1) Pisc icoll2(0, 1) OSTR ACOD2(0, 2) COPE PODA2(0, 1)
 Mysi dae 2(0, 1) Pyra lida2(0, 1) Cera topo2(0, 1) HYDR ACAR2(0, 2)

DIVISION 10 (N= 3) I.E. GROUP *010

Eigenvalue .254 at iteration 4

INDICATORS, together with their SIGN

Hydr oph1(-)

NEGATIVE PREFERENTIALS

Lept ocer1(1, 1) Gamm arid1(1, 1) Plan aril1(1, 1) Elmi nth12(1, 1) Hali plid1(1, 1) Hydr oph11(1, 0)
 Gerr idae1(1, 1) Noto nect1(1, 1) Tipu lida1(1, 0) Pisc icoll1(1, 1) Glos siph1(1, 1) Erpo bdell1(1, 0)
 Asel lida1(1, 0) OSTR ACOD1(1, 0) Cera topo1(1, 0) Lept ocer2(1, 1) Gamm arid2(1, 1) Plan aril2(1, 1)
 Elmi nth12(1, 1) Hali plid2(1, 1) Hydr oph12(1, 0) Gerr idae2(1, 1) Noto nect2(1, 1) Tipu lida2(1, 0)
 Pisc icoll2(1, 1) Glos siph2(1, 1) Erpo bdell2(1, 0) Asel lida2(1, 0) OSTR ACOD2(1, 0) Cera topo2(1, 0)

POSITIVE PREFERENTIALS

Unio nida1(0, 1) Meao veli1(0, 1) Cori xida1(0, 2) CLAD OCER1(0, 1) HYDR ACAR1(0, 2) Unio nida2(0, 1)
 Meao veli2(0, 1) Cori xida2(0, 2) CLAD OCER2(0, 1) HYDR ACAR2(0, 2)

DIVISION 11 (N= 2) I.E. GROUP *011

Eigenvalue .148 at iteration 0

INDICATORS, together with their SIGN

Caen idae1(+)

NEGATIVE PREFERENTIALS

Cera topo1(1, 0) Cera topo2(1, 0)

POSITIVE PREFERENTIALS

Caen idae1(0, 1) Hydr opt11(0, 1) Tipu lida1(0, 1) Asel lida1(0, 1) Mysi dae 1(0, 1) Caen idae2(0, 1)
 Hydr opt12(0, 1) Tipu lida2(0, 1) Asel lida2(0, 1) Mysi dae 2(0, 1)

DIVISION 13 (N= 4) I.E. GROUP *101

Eigenvalue .334 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .036 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

Cori xida1(+)

NEGATIVE PREFERENTIALS

POSITIVE PREFERENTIALS

Hali plid1(0, 1) Dyti scid1(0, 1) Cori xida1(0, 2) Gerr idae1(0, 1) Baet idae1(0, 2) Glos siph1(0, 1)
 Hydr obil1(0, 1) Plan orb1(0, 2) Olig ocha1(1, 2) COPE PODA1(0, 2) HYDR ACAR1(1, 2) Hali plid2(0, 1)
 Dyti scid2(0, 1) Cori xida2(0, 2) Gerr idae2(0, 1) Baet idae2(0, 2) Glos siph2(0, 1) Hydr obil2(0, 1)
 Plan orb12(0, 2) Olig ocha2(1, 2) COPE PODA2(0, 2) HYDR ACAR2(1, 2)

Appendix 3.2.5 Indicator and preferential taxa for EBR

TWINSpan "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of EBR samples (Section 5.6.5, Figure 15). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 16) I.E. GROUP *

Eigenvalue .174 at iteration 3

INDICATORS, together with their SIGN

Poly cent1(+) COPE PODA1(+)

NEGATIVE PREFERENTIALS

Hydr opsy1(2, 0) Limn eph1(7, 2) Lept ocer1(6, 0) Seri cost1(2, 0) Psyc hodi1(3, 0) Hydr opsy2(2, 0)

Limn eph2(7, 2) Lept ocer2(6, 0) Seri cost2(2, 0) Psyc hodi2(3, 0)

POSITIVE PREFERENTIALS

OSTR ACOD1(0, 3) CLAD OCER1(0, 3) COPE PODA1(0, 4) Leuc trid1(0, 2) Ephe mer1(3, 5) Poly cent1(0, 5)

Hydr opt1(1, 4) Halli plid1(5, 6) Gyri nida1(0, 2) Hydr ophi1(0, 2) Valv atid1(0, 2) Phys idae1(0, 2)

OSTR ACOD2(0, 3) CLAD OCER2(0, 3) COPE PODA2(0, 4) Leuc trid2(0, 2) Ephe mere2(3, 5) Poly cent2(0, 5)

Hydr opt2(1, 4) Halli plid2(5, 6) Gyri nida2(0, 2) Hydr ophi2(0, 2) Valv atid2(0, 2) Phys idae2(0, 2)

DIVISION 2 (N= 10) I.E. GROUP *0

Eigenvalue .227 at iteration 1

INDICATORS, together with their SIGN

Sial idae1(+)

NEGATIVE PREFERENTIALS

NEMA TODA1(1, 0) Pisc icoll(1, 0) Lept ophi1(3, 1) Ephe mer1(3, 0) Ephe mer1(1, 0) Caen idae1(4, 1)

Hydr opsy1(2, 0) Hydr opt1(1, 0) Limn eph1(4, 3) Glos soso1(1, 0) Noto nect1(2, 1) Simu liid1(4, 0)

Hydr obil1(3, 2) HYDR ACAR1(4, 3) NEMA TODA2(1, 0) Pisc icoll2(1, 0) Lept ophi2(3, 1) Ephe mere2(3, 0)

Ephe mer12(1, 0) Caen idae2(4, 1) Hydr opsy2(2, 0) Hydr opt2(1, 0) Limn eph2(4, 3) Glos soso2(1, 0)

Noto nect2(2, 1) Simu liid2(4, 0) Hydr obil2(3, 2) HYDR ACAR2(4, 3)

POSITIVE PREFERENTIALS

Erpo bdel1(2, 6) Asel lida1(2, 6) Sial idae1(0, 6) Lept ocer1(1, 5) Cera topol(0, 5) Psyc hodi1(0, 3)

Erpo bdel2(2, 6) Asel lida2(2, 6) Sial idae2(0, 6) Lept ocer2(1, 5) Cera topo2(0, 5) Psyc hodi2(0, 3)

DIVISION 3 (N= 6) I.E. GROUP *1

Eigenvalue .199 at iteration 2

INDICATORS, together with their SIGN

Cera topol(-)

NEGATIVE PREFERENTIALS

Coen agril1(1, 0) Leuc trid1(2, 0) Lept ophi1(2, 0) Ephe mer1(1, 0) Caen idae1(3, 0) Limn eph1(2, 0)

Goer idae1(2, 0) Gyri nida1(2, 0) Hydr ophi1(2, 0) Cori xida1(4, 1) Gerr idae1(1, 0) Noto nect1(2, 0)

Cera topol(4, 0) Ptyc hopt1(1, 0) Spha erid1(4, 1) Hydr obil1(3, 0) HYDR ACAR1(4, 1) Coen agril2(1, 0)

Leuc trid2(2, 0) Lept ophi2(2, 0) Ephe mer12(1, 0) Caen idae2(3, 0) Limn eph2(2, 0) Goer idae2(2, 0)

Gyri nida2(2, 0) Hydr ophi2(2, 0) Cori xida2(4, 1) Gerr idae2(1, 0) Noto nect2(2, 0) Cera topo2(4, 0)

Ptyc hopt2(1, 0) Spha erid2(4, 1) Hydr obil2(3, 0) HYDR ACAR2(4, 1)

POSITIVE PREFERENTIALS

Plan arii1(1, 2) Dend rocol(0, 1) Hydr opt1(2, 2) Simu liid1(2, 2) Musc idae1(0, 1) Valv atid1(0, 2)

Phys idae1(0, 2) Plan arii2(1, 2) Dend rocol2(0, 1) Hydr opt2(2, 2) Simu liid2(2, 2) Musc idae2(0, 1)

Valv atid2(0, 2) Phys idae2(0, 2)

DIVISION 4 (N= 4) I.E. GROUP *00

Eigenvalue .194 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .045 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

NEMA TODA1(-)

NEGATIVE PREFERENTIALS

NEMA TODA1(1, 0) Erpo bdel1(1, 1) Hydr opt1(1, 0) Goer idae1(1, 1) Tipu lida1(1, 1) NEMA TODA2(1, 0)

Erpo bdel2(1, 1) Hydr opt2(1, 0) Goer idae2(1, 1) Tipu lida2(1, 1)

POSITIVE PREFERENTIALS

Pisc icoll(0, 1) Glos siph1(0, 2) Asel lida1(0, 2) Lept ophi1(0, 3) Ephe mer1(0, 3) Ephe mer1(0, 3)

Hydr opsy1(0, 2) Lept ocer1(0, 1) Seri cost1(0, 1) Glos soso1(0, 1) Halli plid1(0, 2) Dyti scid1(0, 3)

Cori xida1(0, 2) Noto nect1(0, 2) Pisc icoll2(0, 1) Glos siph2(0, 2) Asel lida2(0, 2) Lept ophi2(0, 3)

Ephe mere2(0, 3) Ephe mer12(0, 1) Hydr opsy2(0, 2) Lept ocer2(0, 1) Seri cost2(0, 1) Glos soso2(0, 1)

Halli plid2(0, 2) Dyti scid2(0, 3) Cori xida2(0, 2) Noto nect2(0, 2)

DIVISION 5 (N= 6) I.E. GROUP *01

Eigenvalue .193 at iteration 1

INDICATORS, together with their SIGN

Plan arii1(-)

NEGATIVE PREFERENTIALS

Plan arii1(4, 0) Dend rocol(1, 0) Glos siph1(4, 1) Gamm arid1(4, 0) Lept ophi1(1, 0) Caen idae1(1, 0)

Goer idae1(2, 0) Seri cost1(1, 0) Elmi nth1(4, 0) Cera topol(4, 1) Psyc hodi1(3, 0) Taba nida1(1, 0)

HYDR ACAR1(3, 0) Plan arii2(4, 0) Dend rocol2(1, 0) Glos siph2(4, 1) Gamm arid2(4, 0) Lept ophi2(1, 0)

Caen idae2(1, 0) Goer idae2(2, 0) Seri cost2(1, 0) Elmi nth2(4, 0) Cera topo2(4, 1) Psyc hodi2(3, 0)

Taba nida2(1, 0) HYDR ACAR2(3, 0)

POSITIVE PREFERENTIALS

Baet idae1(2, 2) Helo dida1(0, 1) Noto nect1(0, 1) Culi cida1(0, 1) Hydr obil1(1, 1) Baet idae2(2, 2)

Helo dida2(0, 1) Noto nect2(0, 1) Culi cida2(0, 1) Hydr obil2(1, 1)

Appendix 3.2.5

DIVISION 6 (N= 4) I.E. GROUP *10

Eigenvalue .134 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .025 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

Leuc trid1(-)

NEGATIVE PREFERENTIALS

Plan ari11(1, 0) Glos siph1(2, 1) Leuc trid1(2, 0) Lept oph11(2, 0) Ephe mer1(2, 1) Caen idae1(2, 1)
Poly cent1(2, 1) Hydr opt11(2, 0) Elmi nth11(2, 1) Gerr idae1(1, 0) Noto nect1(2, 0) Tipu lida1(2, 1)
Hydr obi11(2, 1) Plan eri12(1, 0) Glos siph2(2, 1) Leuc trid2(2, 0) Lept oph12(2, 0) Ephe mer2(2, 1)
Caen idae2(2, 1) Poly cent2(2, 1) Hydr opt12(2, 0) Elmi nth12(2, 1) Gerr idae2(1, 0) Noto nect2(2, 0)
Tipu lida2(2, 1) Hydr obi12(2, 1)

POSITIVE PREFERENTIALS

COPE PODA1(1, 2) Asel lida1(1, 2) Coen agr11(0, 1) Ephe mer11(0, 1) Sial idae1(0, 2) Gyri nida1(0, 2)
Ptyc hopt1(0, 1) COPE PODA2(1, 2) Asel lida2(1, 2) Coen agr12(0, 1) Ephe mer12(0, 1) Sial idae2(0, 2)
Gyri nida2(0, 2) Ptyc hopt2(0, 1)

DIVISION 7 (N= 2) I.E. GROUP *11

Eigenvalue .225 at iteration 0

INDICATORS, together with their SIGN

Dend rocol(-)

NEGATIVE PREFERENTIALS

Dend rocol(1, 0) OSTR ACOD1(1, 0) CLAD OCER1(1, 0) Elmi nth11(1, 0) Cori xida1(1, 0) Tipu lida1(1, 0)
Musc idae1(1, 0) HYDR ACAR1(1, 0) Dend roco2(1, 0) OSTR ACOD2(1, 0) CLAD OCER2(1, 0) Elmi nth12(1, 0)
Cori xida2(1, 0) Tipu lida2(1, 0) Musc idae2(1, 0) HYDR ACAR2(1, 0)

POSITIVE PREFERENTIALS

COPE PODA1(0, 1) Sial idae1(0, 1) Spha erid1(0, 1) COPE PODA2(0, 1) Sial idae2(0, 1) Spha erid2(0, 1)

DIVISION 9 (N= 3) I.E. GROUP *001

Eigenvalue .276 at iteration 2

INDICATORS, together with their SIGN

Pisc icoll(-)

NEGATIVE PREFERENTIALS

Pisc icoll(1, 0) Glos siph1(1, 1) Asel lida1(1, 1) Hydr opsy1(1, 1) Lept ocer1(1, 0) Goer idae1(1, 0)
Seri cost1(1, 0) Glos siph2(1, 1) Tipu lida1(1, 0) Hydr obi11(1, 1) Plan orbi1(1, 1) Pisc icoll2(1, 0)
Glos siph2(1, 1) Asel lida2(1, 1) Hydr opsy2(1, 1) Lept ocer2(1, 0) Goer idae2(1, 0) Seri cost2(1, 0)
Glos soso2(1, 0) Tipu lida2(1, 0) Hydr obi12(1, 1) Plan orbi2(1, 1)

POSITIVE PREFERENTIALS

Erpo bdel1(0, 1) Ephe mer11(0, 1) Hali plid1(0, 2) Cori xida1(0, 2) Noto nect1(0, 2) Spha erid1(0, 2)
Lynn seid1(0, 2) Erpo bdel2(0, 1) Ephe mer12(0, 1) Hali plid2(0, 2) Cori xida2(0, 2) Noto nect2(0, 2)
Spha erid2(0, 2) Lynn seid2(0, 2)

DIVISION 10 (N= 4) I.E. GROUP *010

Eigenvalue .181 at iteration 6

RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .009 INSTEAD OF .003 (THE TOLERANCE)

INDICATORS, together with their SIGN

Dend rocol(+)

NEGATIVE PREFERENTIALS

Baet idae1(2, 0) Lept oph11(1, 0) Caen idae1(1, 0) Limn eph11(2, 0) Tipu lida1(3, 0) Psyc hod11(3, 0)
Taba nida1(1, 0) Hydr obi11(1, 0) Lynn seid1(3, 0) Baet idae2(2, 0) Lept oph12(1, 0) Caen idae2(1, 0)
Limn eph12(2, 0) Tipu lida2(3, 0) Psyc hod12(3, 0) Taba nida2(1, 0) Hydr obi12(1, 0) Lynn seid2(3, 0)

POSITIVE PREFERENTIALS

Dend rocol(0, 1) Goer idae1(1, 1) Seri cost1(0, 1) Hali plid1(1, 1) Dend roco2(0, 1) Goer idae2(1, 1)
Seri cost2(0, 1) Hali plid2(1, 1)

DIVISION 11 (N= 2) I.E. GROUP *011

Eigenvalue .310 at iteration 0

INDICATORS, together with their SIGN

Glos siph1(+)

NEGATIVE PREFERENTIALS

Hali plid1(1, 0) Noto nect1(1, 0) Tipu lida1(1, 0) Spha erid1(1, 0) Hali plid2(1, 0) Noto nect2(1, 0)
Tipu lida2(1, 0) Spha erid2(1, 0)

POSITIVE PREFERENTIALS

Glos siph1(0, 1) Limn eph11(0, 1) Melo dida1(0, 1) Cera topol(0, 1) Culi cida1(0, 1) Hydr obi11(0, 1)
Glos siph2(0, 1) Limn eph12(0, 1) Melo dida2(0, 1) Cera topo2(0, 1) Culi cida2(0, 1) Hydr obi12(0, 1)

Appendix 3.2.5

DIVISION 12 (N= 2) I.E. GROUP *100
 Eigenvalue .168 at iteration 0
 INDICATORS, together with their SIGN
 Plan ari11(+)

NEGATIVE PREFERENTIALS

OSTR ACOD1(1, 0) CLAD OCER1(1, 0) Goer idae1(1, 0) OSTR ACOD2(1, 0) CLAD OCER2(1, 0) Goer idae2(1, 0)

POSITIVE PREFERENTIALS

Plan ari11(0, 1) COPE PODA1(0, 1) Asel lida1(0, 1) Limn ephi1(0, 1) Hydr ophi1(0, 1) Gerr idae1(0, 1)
 Simu liid1(0, 1) Plan ari12(0, 1) COPE PODA2(0, 1) Asel lida2(0, 1) Limn ephi2(0, 1) Hydr ophi2(0, 1)
 Gerr idae2(0, 1) Simu liid2(0, 1)

DIVISION 13 (N= 2) I.E. GROUP *101
 Eigenvalue .316 at iteration 0
 INDICATORS, together with their SIGN
 Glos siph1(+)

NEGATIVE PREFERENTIALS

Ephe mere1(1, 0) Ephe meri2(1, 0) Caen idae1(1, 0) Poly cent1(1, 0) Limn ephi1(1, 0) Goer idae1(1, 0)
 Elmi nthi1(1, 0) Tipu lida1(1, 0) Simu liid1(1, 0) Hydr obli1(1, 0) Ephe mere2(1, 0) Ephe meri2(1, 0)
 Caen idae2(1, 0) Poly cent2(1, 0) Limn ephi2(1, 0) Goer idae2(1, 0) Elmi nthi2(1, 0) Tipu lida2(1, 0)
 Simu liid2(1, 0) Hydr obli2(1, 0)

POSITIVE PREFERENTIALS

Glos siph1(0, 1) OSTR ACOD1(0, 1) CLAD OCER1(0, 1) Coen aqri1(0, 1) Hydr ophi1(0, 1) Ptyc hopt1(0, 1)
 Glos siph2(0, 1) OSTR ACOD2(0, 1) CLAD OCER2(0, 1) Coen aqri2(0, 1) Hydr ophi2(0, 1) Ptyc hopt2(0, 1)

APPENDIX 3.3

TWINSPAN PREFERENTIAL TAXA FOR THE CLASSIFICATION OF SAMPLES WITHIN LOI CLASSES A - A++ AND B & C.

3.3.1 LQI classes A - A++

3.3.2 LQI classes B & C

Appendix 3.3.1 LOI classes A-A++

TWINSpan "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of A-A++ class samples (Section 5.8.2, Figures 20 and 21). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 66) I.E. GROUP *
Eigenvalue .188 at iteration 2
INDICATORS, together with their SIGN
Coen agril(-) Hydr opsy1(+) Valv atid1(-) Goer idae1(+) Simu liid1(+)

NEGATIVE PREFERENTIALS

OSTR ACOD1(15, 4) CLAD OCER1(9, 2) Coen agril(22, 1) Hydr ophi1(6, 2) Gerr idae1(8, 4) Noto nect1(13, 9)
Valv atid1(20, 1) Phys idae1(18, 11)

POSITIVE PREFERENTIALS

Dend rocol(4, 17) Lept ophi1(4, 22) Ephe mere1(2, 26) Ephe meril1(2, 20) Rhya coph1(2, 14) Poly cent1(2, 18)
Hydr opsy1(1, 30) Limn ephi1(6, 31) Goer idae1(0, 24) Seri cost1(0, 18) Elmi nth1(13, 38) Tipu lida1(7, 28)
Simu liid1(1, 24) Ancy lida1(0, 11)

DIVISION 2 (N= 27) I.E. GROUP *0
Eigenvalue .090 at iteration 1
INDICATORS, together with their SIGN
Dend rocol(+)

NEGATIVE PREFERENTIALS

Hydr ophi1(6, 0) Unio nida1(5, 0) Hydr ophi2(6, 0) Unio nida2(5, 0)

POSITIVE PREFERENTIALS

Dend rocol(0, 4) OSTR ACOD1(11, 4) CLAD OCER1(5, 4) Agri idae1(0, 1) Ephe meril1(0, 2) Sial idae1(7, 4)
Poly cent1(0, 2) Hydr opsy1(0, 1) Hydr optil1(5, 4) Mola nnid1(1, 1) Elmi nth1(9, 4) Gyri nida1(2, 2)
Tipu lida1(4, 3) Simu liid1(0, 1) Cera topo1(7, 3) Musc idae1(0, 2) Succ ineil1(0, 1) Dend roco2(0, 4)
OSTR ACOD2(11, 4) CLAD OCER2(5, 4) Agri idae2(0, 1) Ephe meri2(0, 2) Sial idae2(7, 4) Poly cent2(0, 2)
Hydr opsy2(0, 1) Hydr opti2(5, 4) Mola nnid2(1, 1) Elmi nthi2(9, 4) Gyri nida2(2, 2) Tipu lida2(4, 3)
Simu liid2(0, 1) Cera topo2(7, 3) Musc idae2(0, 2) Succ inei2(0, 1)

DIVISION 3 (N= 39) I.E. GROUP *1
Eigenvalue .093 at iteration 2
INDICATORS, together with their SIGN
Cori xida1(-) Rhya coph1(+) Ephe meril1(-) Phys idae1(-) Ancy lida1(+)

NEGATIVE PREFERENTIALS

Ephe meril1(17, 3) Sial idae1(10, 3) Cori xida1(22, 5) Noto nect1(8, 1) Phys idae1(11, 0) Ephe meri2(17, 3)
Sial idae2(10, 3) Cori xida2(22, 5) Noto nect2(8, 1) Phys idae2(11, 0)

POSITIVE PREFERENTIALS

Rhya coph1(2, 12) Gloe sosol(0, 4) Helo dida1(2, 5) Ancy lida1(2, 9) Rhya coph2(2, 12) Gloe sosol2(0, 4)
Helo dida2(2, 5) Ancy lida2(2, 9)

DIVISION 4 (N= 23) I.E. GROUP *00
Eigenvalue .100 at iteration 3
INDICATORS, together with their SIGN
Lept ophi1(-)

NEGATIVE PREFERENTIALS

COPE PODA1(2, 5) Lept ophi1(3, 0) Ephe mere1(1, 1) Limn ephi1(2, 3) Helo dida1(1, 2) COPE PODA2(2, 5)
Lept ophi2(3, 0) Ephe mere2(1, 1) Limn ephi2(2, 3) Helo dida2(1, 2)

POSITIVE PREFERENTIALS

OSTR ACOD1(0, 11) CLAD OCER1(0, 5) Sial idae1(0, 7) Hydr optil1(0, 5) Phry ganel(0, 4) Lept ocer1(1, 16)
Hydr ophi1(0, 6) Gerr idae1(0, 7) Tipu lida1(0, 4) Cera topo1(0, 7) Unio nida1(0, 5) Valv atid1(0, 16)
Plan orbi1(0, 19) OSTR ACOD2(0, 11) CLAD OCER2(0, 5) Sial idae2(0, 7) Hydr opti2(0, 5) Phry gane2(0, 4)
Lept ocer2(1, 16) Hydr ophi2(0, 6) Gerr idae2(0, 7) Tipu lida2(0, 4) Cera topo2(0, 7) Unio nida2(0, 5)
Valv atid2(0, 16) Plan orbi2(0, 19)

DIVISION 5 (N= 4) I.E. GROUP *01
Eigenvalue .098 at iteration 6
RA TROUBLE 6 ITERATIONS, AND RESIDUAL IS STILL .023 INSTEAD OF .003 (THE TOLERANCE)
INDICATORS, together with their SIGN
Pisc icoll(-)

NEGATIVE PREFERENTIALS

Pisc icoll(2, 0) COPE PODA1(1, 0) Coen agril(2, 1) Poly cent1(2, 0) Hydr opsy1(1, 0) Phry ganel(1, 0)
Dyti acid1(2, 1) Cera topo1(2, 1) Succ ineil1(1, 0) Pisc icoll2(2, 0) COPE PODA2(1, 0) Coen agril2(2, 1)
Poly cent2(2, 0) Hydr opsy2(1, 0) Phry gane2(1, 0) Dyti acid2(2, 1) Cera topo2(2, 1) Succ inei2(1, 0)

POSITIVE PREFERENTIALS

Agri idae1(0, 1) Lept ophi1(0, 1) Limn ephi1(0, 1) Mola nnid1(0, 1) Hydr omet1(0, 1) Gerr idae1(0, 1)
Tipu lida1(1, 2) Simu liid1(0, 1) Spha erid1(1, 2) Agri idae2(0, 1) Lept ophi2(0, 1) Limn ephi2(0, 1)
Mola nnid2(0, 1) Hydr omet2(0, 1) Gerr idae2(0, 1) Tipu lida2(1, 2) Simu liid2(0, 1) Spha erid2(1, 2)

Appendix 3.3.1

DIVISION 6 (N= 24) I.E. GROUP *10

Eigenvalue .097 at iteration 2
INDICATORS, together with their SIGN
Seri cost1(+) Ephe meril(+)

NEGATIVE PREFERENTIALS

CLAD OCER1(2, 0) Leuc trid1(2, 1) Ephe meril(5, 9) Hydr opt1(3, 5) Gerr idae1(2, 2) Noto nect1(3, 5)
CLAD OCER2(2, 0) Leuc trid2(2, 1) Ephe mere2(5, 9) Hydr opt12(3, 5) Gerr idae2(2, 2) Noto nect2(3, 5)

POSITIVE PREFERENTIALS

Dend rocol(0, 11) Pisc icoll(1, 11) Ephe meril(0, 17) Sial idae1(0, 10) Hydr opsy1(2, 16) Lept ocer1(1, 15)
Seri cost1(0, 12) Gyri nidal(0, 5) Phys idae1(1, 10) Dend roco2(0, 11) Pisc icol2(1, 11) Ephe meri2(0, 17)
Sial idae2(0, 10) Hydr opsy2(2, 16) Lept ocer2(1, 15) Seri cost2(0, 12) Gyri nida2(0, 5) Phys idae2(1, 10)

DIVISION 7 (N= 15) I.E. GROUP *11

Eigenvalue .119 at iteration 2
INDICATORS, together with their SIGN
Cori xidal(-)

NEGATIVE PREFERENTIALS

Dend rocol(3, 3) Pisc icoll(4, 4) COPE PODA1(3, 1) Asel lidal(4, 2) Ephe meril(2, 1) Sial idae1(2, 1)
Seri cost1(3, 3) Halli plid1(5, 2) Gyri nidal(2, 0) Cori xidal(5, 0) Spha erid1(5, 5) Lymn aeid1(5, 2)
Plan orb1(5, 2) Dend roco2(3, 3) Pisc icol2(4, 4) COPE PODA2(3, 1) Asel lida2(4, 2) Ephe meri2(2, 1)
Sial idae2(2, 1) Seri cost2(3, 3) Halli plid2(5, 2) Gyri nida2(2, 0) Cori xida2(5, 0) Spha erid2(5, 5)
Lymn aeid2(5, 2) Plan orb2(5, 2)

POSITIVE PREFERENTIALS

Poly cent1(1, 4) Lept ocer1(1, 6) Glos sosal(0, 4) Poly cent2(1, 4) Lept ocer2(1, 6) Glos soso2(0, 4)

DIVISION 8 (N= 3) I.E. GROUP *000

Eigenvalue .262 at iteration 2
INDICATORS, together with their SIGN
Plan ari1(+)

NEGATIVE PREFERENTIALS

Pisc icoll(1, 0) COPE PODA1(1, 1) Asel lidal(1, 1) Gamm arid1(1, 1) Ephe meril(1, 0) Limn eph1(1, 1)
Lept ocer1(1, 0) Elmi nth1(1, 1) Cori xidal(1, 1) Noto nect1(1, 0) Chir oncm1(1, 1) Phys idae1(1, 0)
HYDR ACAR1(1, 1) Pisc icol2(1, 0) COPE PODA2(1, 1) Asel lida2(1, 1) Gamm arid2(1, 1) Ephe mere2(1, 0)
Limn eph2(1, 1) Lept ocer2(1, 0) Elmi nth2(1, 1) Cori xida2(1, 1) Noto nect2(1, 0) Chir oncm2(1, 1)
Phys idae2(1, 0) HYDR ACAR2(1, 1)

POSITIVE PREFERENTIALS

Plan ari1(0, 2) Glos siph1(0, 2) Erpo bdell(0, 2) Helo dida1(0, 1) Spha erid1(0, 2) Plan ari2(0, 2)
Glos siph2(0, 2) Erpo bdell2(0, 2) Helo dida2(0, 1) Spna erid2(0, 2)

DIVISION 9 (N= 20) I.E. GROUP *001

Eigenvalue .093 at iteration 2
INDICATORS, together with their SIGN
Elmi nth1(+), COPE PODA1(+)

NEGATIVE PREFERENTIALS

Sial idae1(6, 1) Limn eph1(3, 0) Unio nidal(5, 0) Sial idae2(6, 1) Limn eph2(3, 0) Unio nida2(5, 0)

POSITIVE PREFERENTIALS

COPE PODA1(0, 5) Elmi nth1(0, 7) Hydr oph1(1, 5) Hydr omet1(1, 2) Neri tidal(0, 2) COPE PODA2(0, 5)
Elmi nth2(0, 7) Hydr oph2(1, 5) Hydr omet2(1, 2) Neri tida2(0, 2)

DIVISION 10 (N= 2) I.E. GROUP *010

Eigenvalue .137 at iteration 0
INDICATORS, together with their SIGN
COPE PODA1(+)

NEGATIVE PREFERENTIALS

Phry gane1(1, 0) Succ ine1(1, 0) Phry gane2(1, 0) Succ ine2(1, 0)

POSITIVE PREFERENTIALS

COPE PODA1(0, 1) Ephe meril(0, 1) Hydr opsy1(0, 1) Gyri nidal(0, 1) Noto nect1(0, 1) Tipu lida1(0, 1)
Musc idae1(0, 1) Spha erid1(0, 1) COPE PODA2(0, 1) Ephe meri2(0, 1) Hydr opsy2(0, 1) Gyri nida2(0, 1)
Noto nect2(0, 1) Tipu lida2(0, 1) Musc idae2(0, 1) Spha erid2(0, 1)

DIVISION 11 (N= 2) I.E. GROUP *011

Eigenvalue .203 at iteration 0
INDICATORS, together with their SIGN
Coen agr1(-)

NEGATIVE PREFERENTIALS

Coen agr1(1, 0) Mola nnid1(1, 0) Dyti scid1(1, 0) Gyri nidal(1, 0) Hydr omet1(1, 0) Gerr idae1(1, 0)
Noto nect1(1, 0) Cera topo1(1, 0) Musc idae1(1, 0) Coen agr2(1, 0) Mola nnid2(1, 0) Dyti scid2(1, 0)
Gyri nida2(1, 0) Hydr omet2(1, 0) Gerr idae2(1, 0) Noto nect2(1, 0) Cera topo2(1, 0) Musc idae2(1, 0)

POSITIVE PREFERENTIALS

Agri idae1(0, 1) Lept oph1(0, 1) Ephe meril(0, 1) Limn eph1(0, 1) Simu liid1(0, 1) Agri idae2(0, 1)
Lept oph2(0, 1) Ephe meri2(0, 1) Limn eph2(0, 1) Simu liid2(0, 1)

Appendix 3.3.1

DIVISION 12 (N= 5) I.E. GROUP *100
Eigenvalue .212 at iteration 1
INDICATORS, together with their SIGN
Caen idae1(+)

NEGATIVE PREFERENTIALS

Asel lidal(1, 2) Dryo pida(1, 0) Gerr idae1(1, 1) Asel lida2(1, 2) Dryo pida2(1, 0) Gerr idae2(1, 1)

POSITIVE PREFERENTIALS

Plan aril1(0, 3) Pisc icoll(0, 1) Glos siph1(0, 3) Erpo bdel1(0, 3) OSTR ACOD1(0, 1) CLAD OCER1(0, 2)
COPE PODA1(0, 1) Leuc trid1(0, 2) Lept oph11(0, 3) Caen idae1(0, 4) Poly cent1(0, 3) Hydr opsy1(0, 2)
Hydr opt11(0, 3) Lept ocer1(0, 1) Goer idae1(0, 2) Elmi nth11(0, 4) Hali plid1(0, 3) Hydr oph11(0, 1)
Noto nect1(0, 3) Tipu lida1(0, 2) Cera topol(0, 3) Spha erid1(0, 4) Hydr obil1(0, 4) Lymn aeid1(0, 4)
Phys idae1(0, 1) Plan aril2(0, 3) Pisc icol2(0, 1) Glos siph2(0, 3) Erpo bdel2(0, 3) OSTR ACOD2(0, 1)
CLAD OCER2(0, 2) COPE PODA2(0, 1) Leuc trid2(0, 2) Lept oph12(0, 3) Caen idae2(0, 4) Poly cent2(0, 3)
Hydr opsy2(0, 2) Hydr opt12(0, 3) Lept ocer2(0, 1) Goer idae2(0, 2) Elmi nth12(0, 4) Hali plid2(0, 3)
Hydr oph12(0, 1) Noto nect2(0, 3) Tipu lida2(0, 2) Cera topo2(0, 3) Spha erid2(0, 4) Hydr obil2(0, 4)
Lymn aeid2(0, 4) Phys idae2(0, 1)

DIVISION 13 (N= 19) I.E. GROUP *101

Eigenvalue .104 at iteration 2

INDICATORS, together with their SIGN

Plan aril1(-) Phys idae1(-) Hydr opt11(-) Lymn ephil(+) COPE PODA1(+)

NEGATIVE PREFERENTIALS

Plan aril1(11, 2) Ephe merel(7, 2) Hydr opt11(5, 0) Mola nnid1(3, 0) Vell idae1(3, 0) Noto nect1(4, 1)
Simu liid1(6, 2) Phys idae1(8, 2) Plan aril2(11, 2) Ephe mere2(7, 2) Hydr opt12(5, 0) Mola nnid2(3, 0)
Vell idae2(3, 0) Noto nect2(4, 1) Simu liid2(6, 2) Phys idae2(8, 2)

POSITIVE PREFERENTIALS

OSTR ACOD1(0, 3) COPE PODA1(1, 4) Asel lidal(4, 6) Psyc homy1(0, 3) Lepi dost1(0, 2) OSTR ACOD2(0, 3)
COPE PODA2(1, 4) Asel lida2(4, 6) Psyc homy2(0, 3) Lepi dost2(0, 2)

DIVISION 14 (N= 5) I.E. GROUP *110

Eigenvalue .194 at iteration 1

INDICATORS, together with their SIGN

Dend rocol(-)

NEGATIVE PREFERENTIALS

Dend rocol(3, 0) Ephe merel(3, 1) Poly cent1(1, 0) Psyc homy1(1, 0) Hydr opsy1(3, 1) Hydr opt11(2, 0)
Lept ocer1(1, 0) Lepi dost1(1, 0) Gyr1 nidal(2, 0) Chry same1(1, 0) Noto nect1(1, 0) Valv atid1(1, 0)
Dend roco2(3, 0) Ephe mere2(3, 1) Poly cent2(1, 0) Psyc homy2(1, 0) Hydr opsy2(3, 1) Hydr opt12(2, 0)
Lept ocer2(1, 0) Lepi dost2(1, 0) Gyr1 nida2(2, 0) Chry same2(1, 0) Noto nect2(1, 0) Valv atid2(1, 0)

POSITIVE PREFERENTIALS

NEMA TODA1(0, 1) NEMA TOMO1(0, 1) Lept oph11(0, 2) Seri cost1(1, 2) Helo dida1(0, 2) Cera topol(0, 1)
Psyc hod11(0, 1) Taba nidal(0, 1) Musc idae1(0, 1) NEMA TODA2(0, 1) NEMA TOMO2(0, 1) Lept oph12(0, 2)
Seri cost2(1, 2) Helo dida2(0, 2) Cera topo2(0, 1) Psyc hod12(0, 1) Taba nida2(0, 1) Musc idae2(0, 1)

DIVISION 15 (N= 10) I.E. GROUP *111

Eigenvalue .142 at iteration 1

INDICATORS, together with their SIGN

Glos siph1(-)

NEGATIVE PREFERENTIALS

Pisc icoll(4, 0) Glos siph1(7, 0) Asel lidal(2, 0) Lept oph11(6, 1) Cera topol(3, 0) Lymn aeid1(2, 0)
Plan orbil(2, 0) Pisc icol2(4, 0) Glos siph2(7, 0) Asel lida2(2, 0) Lept oph12(6, 1) Cera topo2(3, 0)
Lymn aeid2(2, 0) Plan orbil2(2, 0)

POSITIVE PREFERENTIALS

Dend rocol(1, 2) Leuc trid1(0, 1) Ephe meril(0, 1) Sial idae1(0, 1) Poly cent1(1, 3) Lepi dost1(0, 1)
Seri cost1(1, 2) Hali plid1(1, 1) Psyc hod11(0, 1) Dend roco2(1, 2) Leuc trid2(0, 1) Ephe meri2(0, 1)
Sial idae2(0, 1) Poly cent2(1, 3) Lepi dost2(0, 1) Seri cost2(1, 2) Hali plid2(1, 1) Psyc hod12(0, 1)

Appendix 3.3.2 LOI classes B and C

TWINSpan "indicator" taxa and negative (left-hand groups) and positive (right-hand groups) preferential taxa from the classification of B and C class samples (Section 5.8.2, Figures 23 and 24). N = the number of samples in the division; numbers after preferential taxa indicate the number of samples in the negative and positive groups in which these taxa occur.

DIVISION 1 (N= 88) I.E. GROUP *

Eigenvalue .197 at iteration 2

INDICATORS, together with their SIGN

Elmi nth1(-) Tipu lidal(-) Valv atid1(+) Coen agril1(+) Limn ephil(-) Simu liid1(-) Hydr opsy1(-)

CLAD OCER1(+)

NEGATIVE PREFERENTIALS

Lept ophil1(10, 0) Ephe merel(12, 3) Sial idael(12, 3) Rhya coph1(15, 1) Hydr opsy1(25, 1) Limn ephil(28, 0)

Lept ocer1(17, 6) Goer idael(13, 0) Elmi nth1(46, 13) Tipu lidal(39, 8) Simu liid1(31, 2) Ancy lidal(15, 0)

POSITIVE PREFERENTIALS

OSTR ACOD1(4, 14) CLAD OCER1(1, 19) COPE PODA1(7, 17) Coen agril1(3, 27) Valv atid1(1, 25) Phys idael(5, 18)

DIVISION 2 (N= 48) I.E. GROUP *0

Eigenvalue .129 at iteration 1

INDICATORS, together with their SIGN

Rhya coph1(-) Lept ocer1(+) Plan orbil1(+) Ancy lidal(-) Plan aril1(-) Pisc icoll1(-) Caen idael(+)

Limn ephil(-) Ase1 lidal(+)

NEGATIVE PREFERENTIALS

Plan aril1(21, 10) Pisc icoll1(11, 1) Ephe merel(9, 3) Rhya coph1(15, 0) Limn ephil(19, 9) Goer idael(9, 4)

Glos sosol(5, 0) Ancy lidal(13, 2) Plan aril2(21, 10) Pisc icol2(11, 1) Ephe merel2(9, 3) Rhya coph2(15, 0)

Limn ephil2(19, 9) Goer idael2(9, 4) Glos sosol2(5, 0) Ancy lidal2(13, 2)

POSITIVE PREFERENTIALS

Ase1 lidal(6, 16) Lept ophil1(3, 7) Caen idael(10, 20) Sial idael(4, 8) Lept ocer1(2, 15) Cori xidal(7, 15)

Cera topol(2, 9) Phys idael(0, 5) Plan orbil1(6, 19) Ase1 lidal2(6, 16) Lept ophil2(3, 7) Caen idael2(10, 20)

Sial idael2(4, 8) Lept ocer2(2, 15) Cori xidal2(7, 15) Cera topo2(2, 9) Phys idael2(0, 5) Plan orbil2(6, 19)

DIVISION 3 (N= 40) I.E. GROUP *1

Eigenvalue .097 at iteration 3

INDICATORS, together with their SIGN

Dend rocol1(-) Noto nect1(-) Elmi nth1(-) PORI FERA1(-)

NEGATIVE PREFERENTIALS

PORI FERA1(3, 0) Dend rocol(5, 3) Poly cent1(2, 2) Lept ocer1(2, 4) Pyra lidal(2, 0) Elmi nth1(6, 7)

Noto nect1(5, 2) Simu liid1(2, 0) Musc idael(3, 2) PORI FERA2(3, 0) Dend rocol2(5, 3) Poly cent2(2, 2)

Lept ocer2(2, 4) Pyra lidal2(2, 0) Elmi nth12(6, 7) Noto nect2(5, 2) Simu liid2(2, 0) Musc idael2(3, 2)

POSITIVE PREFERENTIALS

Pisc icoll1(1, 13) Pisc icol2(1, 13)

DIVISION 4 (N= 24) I.E. GROUP *00

Eigenvalue .140 at iteration 2

INDICATORS, together with their SIGN

OSTR ACOD1(-) Nemo urid1(-)

NEGATIVE PREFERENTIALS

OSTR ACOD1(1, 1) COPE PODA1(1, 3) Nemo urid1(1, 1) Lept ophil1(1, 2) OSTR ACOD2(1, 1) COPE PODA2(1, 3)

Nemo urid2(1, 1) Lept ophil2(1, 2)

POSITIVE PREFERENTIALS

Plan aril1(0, 21) Pisc icoll1(0, 11) Glos siph1(0, 20) Erpo bdel1(0, 16) Ase1 lidal(0, 6) Ephe merel(0, 9)

Caen idael(0, 10) Rhya coph1(0, 15) Hydr opsy1(0, 14) Hydr optil1(0, 11) Goer idael(0, 9) Glos sosol(0, 5)

Elmi nth1(0, 22) Hali plid1(0, 14) Cori xidal(0, 7) Spha erid1(0, 13) Hydr obil1(0, 19) Lymn aeid1(0, 15)

Plan orbil1(0, 6) Ancy lidal(0, 13) HYDR ACAR1(0, 18) Plan aril2(0, 21) Pisc icol2(0, 11) Glos siph2(0, 20)

Erpo bdel2(0, 16) Ase1 lidal2(0, 6) Ephe merel2(0, 9) Caen idael2(0, 10) Rhya coph2(0, 15) Hydr opsy2(0, 14)

Hydr optil2(0, 11) Goer idael2(0, 9) Glos sosol2(0, 5) Elmi nth12(0, 22) Hali plid2(0, 14) Cori xidal2(0, 7)

Spha erid2(0, 13) Hydr obil2(0, 19) Lymn aeid2(0, 15) Plan orbil2(0, 6) Ancy lidal2(0, 13) HYDR ACAR2(0, 18)

DIVISION 5 (N= 24) I.E. GROUP *01

Eigenvalue .144 at iteration 1

INDICATORS, together with their SIGN

Phys idael(+)

NEGATIVE PREFERENTIALS

Lept ophil1(7, 0) Taba nidal(4, 0) Lept ophil2(7, 0) Taba nidal2(4, 0)

POSITIVE PREFERENTIALS

Plan aril1(4, 6) Dend rocol(1, 3) OSTR ACOD1(0, 2) Sial idael(4, 4) Poly cent1(0, 3) Psyc homyl(0, 2)

Hydr optil1(1, 5) Goer idael(1, 3) Musc idael(2, 2) Phys idael(0, 5) Plan aril2(4, 6) Dend rocol2(1, 3)

OSTR ACOD2(0, 2) Sial idael2(4, 4) Poly cent2(0, 3) Psyc homyl2(0, 2) Hydr optil2(1, 5) Goer idael2(1, 3)

Musc idael2(2, 2) Phys idael2(0, 5)

Appendix 3.3.2

DIVISION 6 (N= 7) I.E. GROUP *10-

Eigenvalue .197 at iteration 1
INDICATORS, together with their SIGN
Coen agr11(+)

NEGATIVE PREFERENTIALS

Pisc icoll(1, 0) Erpo bdel1(2, 2) OSTR ACOD1(2, 1) COPE PODA1(1, 1) Ephe merel(1, 0) Poly cent1(2, 0)
Hydr opt11(2, 2) Lept ocer1(1, 1) Tipu lida1(1, 0) Simu liid1(2, 0) Cera topol(1, 1) Musc idae1(2, 1)
Lymn aeid1(2, 2) Pisc icol2(1, 0) Erpo bdel2(2, 2) OSTR ACOD2(2, 1) COPE PODA2(1, 1) Ephe mere2(1, 0)
Poly cent2(2, 0) Hydr opt12(2, 2) Lept ocer2(1, 1) Tipu lida2(1, 0) Simu liid2(2, 0) Cera topo2(1, 1)
Musc idae2(2, 1) Lymn aeid2(2, 2)

POSITIVE PREFERENTIALS

Coen agr11(0, 5) Caen idae1(0, 3) Pyra lida1(0, 2) Elmi nth1(1, 5) Noto nect1(0, 5) Spha erid1(1, 5)
Hydr obil1(1, 5) Coen agr12(0, 5) Caen idae2(0, 3) Pyra lida2(0, 2) Elmi nth2(1, 5) Noto nect2(0, 5)
Spha erid2(1, 5) Hydr obil2(1, 5)

DIVISION 7 (N= 33) I.E. GROUP *11

Eigenvalue .105 at iteration 2
INDICATORS, together with their SIGN

Spha erid1(-) Plan ari11(-) Valv atid1(-) Plan orb11(-) Gamm erid1(-) Phys idae1(-) Glos siph1(-)
Hydr obil1(-) Caen idae1(-)

NEGATIVE PREFERENTIALS

Plan ari11(21, 2) Pisc icoll(12, 1) Glos siph1(19, 3) Caen idae1(19, 3) Hydr opt11(9, 1) Spha erid1(17, 0)
Valv atid1(19, 1) Phys idae1(12, 1) Plan orb11(25, 4) Plan ari12(21, 2) Pisc icol2(12, 1) Glos siph2(19, 3)
Caen idae2(19, 3) Hydr opt12(9, 1) Spha erid2(17, 0) Valv atid2(19, 1) Phys idae2(12, 1) Plan orb12(25, 4)

POSITIVE PREFERENTIALS

Ephe-mere1(-0,-2)-Poly-cent1(-0,-2)-Gerr-idae1(-1,-2)-Cera-topol(-7,-5)-Ephe-mere2(-0,-2)-Poly-cent2(-0,-2)-
Gerr-idae2(1, 2) Cera topo2(7, 5)

DIVISION 8 (N= 1) I.E. GROUP *000

DIVISION FAILS - There are too few items

DIVISION 9 (N= 23) I.E. GROUP *001

Eigenvalue .125 at iteration 3
INDICATORS, together with their SIGN

Ancy lida1(+) Ase1 lida1(+) Spha erid1(+) Ephe merel(+)

NEGATIVE PREFERENTIALS

Glos sosol(4, 1) Glos soso2(4, 1)

POSITIVE PREFERENTIALS

Dend rocol(0, 4) Pisc icoll(3, 8) Erpo bdel1(5, 11) Ase1 lida1(0, 6) Ephe merel(0, 9) Sial idae1(0, 4)
Cori xida1(1, 6) Spha erid1(3, 10) Plan orb11(1, 5) Ancy lida1(2, 11) Dend roco2(0, 4) Pisc icol2(3, 8)
Erpo bdel2(5, 11) Ase1 lida2(0, 6) Ephe mere2(0, 9) Sial idae2(0, 4) Cori xida2(1, 6) Spha erid2(3, 10)
Plan orb12(1, 5) Ancy lida2(2, 11)

DIVISION 10 (N= 18) I.E. GROUP *010

Eigenvalue .126 at iteration 2

INDICATORS, together with their SIGN

Simu liid1(+) Cori xida1(-) Cera topol(+) Spha erid1(-) Lept oph11(-) Hydr opsy1(-) Lymn aeid1(+)
Musc idae1(+) Unio nida1(+)

NEGATIVE PREFERENTIALS

Plan ari11(4, 0) Lept oph11(7, 0) Ephe merel(3, 0) Sial idae1(4, 0) Hydr opsy1(8, 1) Cori xida1(10, 1)
Pasc hodi1(3, 0) Spha erid1(12, 2) Plan ari12(4, 0) Lept oph12(7, 0) Ephe mere2(3, 0) Sial idae2(4, 0)
Hydr opsy2(8, 1) Cori xida2(10, 1) Pasc hodi2(3, 0) Spha erid2(12, 2)

POSITIVE PREFERENTIALS

Simu liid1(5, 5) Cera topol(3, 4) Musc idae1(0, 2) Unio nida1(0, 2) Lymn aeid1(5, 4) Simu liid2(5, 5)
Cera topo2(3, 4) Musc idae2(0, 2) Unio nida2(0, 2) Lymn aeid2(5, 4)

DIVISION 11 (N= 6) I.E. GROUP *011

Eigenvalue .211 at iteration 1

INDICATORS, together with their SIGN

Pisc icoll(+)

NEGATIVE PREFERENTIALS

Baet idae1(4, 0) Poly cent1(3, 0) Pasc homy1(2, 0) Goer idae1(3, 0) Hali plid1(3, 0) Cori xida1(4, 0)
Cera topol(2, 0) Baet idae2(4, 0) Poly cent2(3, 0) Pasc homy2(2, 0) Goer idae2(3, 0) Hali plid2(3, 0)
Cori xida2(4, 0) Cera topo2(2, 0)

POSITIVE PREFERENTIALS

Dend rocol(2, 1) Pisc icoll(0, 1) OSTR ACOD1(1, 1) Coen agr11(0, 1) Hydr opsy1(1, 1) Lymn eph11(2, 1)
Helo dida1(0, 1) Pasc hodi1(0, 1) Stra tiom1(0, 1) Musc idae1(1, 1) Valv atid1(0, 1) Lymn aeid1(2, 1)
Dend-roco2(2, 1) Pisc icol2(0, 1) OSTR ACOD2(1, 1) Coen agr12(0, 1) Hydr opsy2(1, 1) Lymn eph12(2, 1)
Helo dida2(0, 1) Pasc hodi2(0, 1) Stra tiom2(0, 1) Musc idae2(1, 1) Valv atid2(0, 1) Lymn aeid2(2, 1)

Appendix 3.3.2

DIVISION 12 (N= 2) I.E. GROUP *100

Eigenvalue .221 at iteration 0

INDICATORS, together with their SIGN

PORI FERA1(+)

NEGATIVE PREFERENTIALS

CLAD OCER1(1, 0) Ephe mere1(1, 0) Elmi nth1(1, 0) Tipu lida1(1, 0) Valv atid1(1, 0) CLAD OCER2(1, 0)
Ephe mere2(1, 0) Elmi nth2(1, 0) Tipu lida2(1, 0) Valv atid2(1, 0)

POSITIVE PREFERENTIALS

PORI FERA1(0, 1) Pisc icol1(0, 1) COPE PODA1(0, 1) Lept ocer1(0, 1) Cera topo1(0, 1) Spha erid1(0, 1)
Hydr obil1(0, 1) PORI FERA2(0, 1) Pisc icol2(0, 1) COPE PODA2(0, 1) Lept ocer2(0, 1) Cera topo2(0, 1)
Spha erid2(0, 1) Hydr obil2(0, 1)

DIVISION 13 (N= 5) I.E. GROUP *101

Eigenvalue .226 at iteration 1

INDICATORS, together with their SIGN

PORI FERA1(-)

NEGATIVE PREFERENTIALS

PORI FERA1(2, 0) Dend rocol1(2, 1) Caen idae1(2, 1) Hydr opsy1(1, 0) Lept ocer1(1, 0) Musc idae1(1, 0)
PORI FERA2(2, 0) Dend rocol2(2, 1) Caen idae2(2, 1) Hydr opsy2(1, 0) Lept ocer2(1, 0) Musc idae2(1, 0)

POSITIVE PREFERENTIALS

Plan ari11(1, 3) Glos siph1(1, 3) OSTR ACOD1(0, 1) COPE PODA1(0, 1) Asel lida1(1, 3) Coro phid1(0, 1)
Pyra lida1(0, 2) Helo dida1(0, 1) Cera topo1(0, 1) Neri tida1(0, 1) Valv atid1(1, 3) Lymn aeid1(0, 2)
Phys idae1(0, 3) Plan ari12(1, 3) Glos siph2(1, 3) OSTR ACOD2(0, 1) COPE PODA2(0, 1) Asel lida2(1, 3)
Coro phid2(0, 1) Pyra lida2(0, 2) Helo dida2(0, 1) Cera topo2(0, 1) Neri tida2(0, 1) Valv atid2(1, 3)
Lymn aeid2(0, 2) Phys idae2(0, 3)

DIVISION 14 (N= 25) I.E. GROUP *110

Eigenvalue .096 at iteration 2

INDICATORS, together with their SIGN

Pisc icol1(+) HYDR ACAR1(-) Glos siph1(-) CLAD OCER1(+) Valv atid1(+) OSTR ACOD1(-) Elmi nth1(-)

NEGATIVE PREFERENTIALS

OSTR ACOD1(8, 1) Elmi nth1(5, 0) HYDR ACAR1(12, 2) OSTR ACOD2(8, 1) Elmi nth2(5, 0) HYDR ACAR2(12, 2)

POSITIVE PREFERENTIALS

Pisc icol1(3, 9) CLAD OCER1(4, 7) Tipu lida1(2, 4) Unio nida1(0, 3) Pisc icol2(3, 9) CLAD OCER2(4, 7)
Tipu lida2(2, 4) Unio nida2(0, 3)

DIVISION 15 (N= 8) I.E. GROUP *111

Eigenvalue .298 at iteration 2

INDICATORS, together with their SIGN

Lymn aeid1(-) Plan orbil(-)

NEGATIVE PREFERENTIALS

Plan ari11(2, 0) Glos siph1(2, 1) Elmi nth1(2, 0) Hydr ophil1(1, 0) Meso vel11(1, 0) Hydr omet1(1, 0)
Gerr idae1(2, 0) Tipu lida1(1, 0) Taba nida1(1, 0) Musc idae1(1, 0) Valv atid1(1, 0) Lymn aeid1(3, 1)
Phys idae1(1, 0) Plan orbil1(3, 1) Plan ari12(2, 0) Glos siph2(2, 1) Elmi nth2(2, 0) Hydr ophil2(1, 0)
Meso vel12(1, 0) Hydr omet2(1, 0) Gerr idae2(2, 0) Tipu lida2(1, 0) Taba nida2(1, 0) Musc idae2(1, 0)
Valv atid2(1, 0) Lymn aeid2(3, 1) Phys idae2(1, 0) Plan orbil2(3, 1)

POSITIVE PREFERENTIALS

Erpo bdel1(0, 3) OSTR ACOD1(0, 2) Ephe mere1(0, 2) Caen idae1(0, 3) Poly cent1(0, 2) Erpo bdel2(0, 3)
OSTR ACOD2(0, 2) Ephe mere2(0, 2) Caen idae2(0, 3) Poly cent2(0, 2)